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FIFTY YEARS OF RESEARCH IN ARDS Respiratory Mechanics in Acute Respiratory Distress Syndrome

William R. Henderson¹*, Lu Chen^{2,3}*, Marcelo B. P. Amato⁴, and Laurent J. Brochard^{2,3}

¹Division of Critical Care Medicine, University of British Columbia, Vancouver, British Columbia, Canada; ²Interdepartmental Division of Critical Care Medicine, University of Toronto, Toronto, Ontario, Canada; ³Keenan Research Centre, Li Ka Shing Knowledge Institute, St. Michael's Hospital, Toronto, Ontario, Canada; and ⁴Cardio-Pulmonary Department, Pulmonary Division, Heart Institute (Incor), University of São Paulo, São Paulo, Brazil

ORCID ID: 0000-0002-7512-1865 (L.C.).

Abstract

Acute respiratory distress syndrome is a multifactorial lung injury that continues to be associated with high levels of morbidity and mortality. Mechanical ventilation, although lifesaving, is associated with new iatrogenic injury. Current best practice involves the use of small VT, low plateau and driving pressures, and high levels of positive end-expiratory pressure. Collectively, these interventions are termed "lung-protective ventilation." Recent investigations suggest that individualized measurements of pulmonary mechanical variables rather than population-based ventilation prescriptions may be used to set the ventilator with the potential to improve outcomes beyond those achieved with standard lung protective ventilation. This review outlines the measurement and application of clinically applicable pulmonary mechanical concepts, such as plateau pressures, driving pressure, transpulmonary pressures, stress index, and measurement of strain. In addition, the concept of the "baby lung" and the utility of dynamic in addition to static measures of pulmonary mechanical variables are discussed.

Keywords: mechanical ventilation; resistance; elastance; esophageal pressure; transpulmonary pressure

The acute respiratory distress syndrome (ARDS) is characterized by the rapid onset of severe hypoxic respiratory failure and alterations in pulmonary mechanics. Three main physiological abnormalities characterize ARDS: hypoxemia; reduced capacity to eliminate CO₂; and reduced lung volumes and compliance. LUNG SAFE (Large Observational Study to Understand the Global Impact of Severe Acute Respiratory Failure), a recent multinational observational study, found current intensive care unit and hospital mortality rates from ARDS of 35–46% (1). Whether all forms of ARDS share a common pathophysiologic course characterized by diffuse alveolar damage and neutrophil recruitment to

the lungs is unclear (2–6), but alterations of lung mechanics are ubiquitous and expose the remaining aerated lung to excessive ventilation. The subsequent local release of toxic mediators damages the capillary endothelium and alveolar epithelium. Once a patient with ARDS is placed under mechanical ventilation (MV), the most common abnormality is one of increased elastance secondary to this small lung size.

After the initial description of ARDS, clinical management historically focused on the oxygenation abnormalities. In an attempt to reverse hypoxemia, large VT values of 12-14 ml/kg, and occasionally as large as 24 ml/kg of body weight, were prescribed (7–9). Similarly,

the use of positive end-expiratory pressure (PEEP) levels as high as 44 cm H_2O were proposed to reverse atelectasis and hypoxemia, as indicated by calculated shunt (10). Concerns were raised about the fear of oxygen toxicity (11), but, for a long period, there was little recognition of the possibility that this level of lung distension might be injurious (8).

Ventilator-induced Lung Injury

MV, although often lifesaving in ARDS, may aggravate or initiate lung injury through

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^{*}Co-first authors.

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Correspondence and requests for reprints should be addressed to Laurent J. Brochard, M.D., Critical Care Medicine, St. Michael's Hospital, 30 Bond Street, Toronto, ON M5B 1W8, Canada. E-mail: brochardl@smh.ca

several mechanisms, collectively termed ventilator-induced lung injury (VILI) (12). Separating VILI into injuries that occur at high lung volumes and at low lung volumes provides a useful schema for potential treatment methods for these injuries (13). VILI occurring at high lung volume is due to regional alveolar overdistension (so called "volutrauma"), associated with high end-inspiratory transpulmonary pressure (PL), but not with high airway pressure (Paw) per se. VILI at low lung volume is due to high local tissue stress and parenchymal shear injury, supposedly caused by repetitive opening and closing of alveoli and distal small airways ("atelectrauma"), occurring when PL suddenly increases at early inspiration (14, 15). VILI is associated with increases in pulmonary and systemic inflammatory mediators that may precipitate multiple organ dysfunction syndrome-a process that has been termed "biotrauma" (16, 17).

To mitigate alveolar distension and VILI at high lung volume, limitation of VT and ventilating pressures are applied (18–22). The use of PEEP is believed to be useful in preventing VILI at low lung volumes associated with atelectrauma (14, 23–28). The combined application of low VT and plateau pressure (Pplat) and high **PEEP** is collectively termed lung-protective ventilation. The exact VT and PEEP combination best suited to each patient needs to be individualized. A management approach that accounts for unique patient characteristics through the measurement of respiratory mechanics, such as driving pressure (ΔP) and PL, or the calculation of strain holds the promise of individualized ventilation and improved outcomes. Recent observational studies and post hoc analysis of randomized controlled trials show that markers of distension, lung deformation, or strain (such as ΔP) are strongly correlated with outcome in ARDS (29). Data also show an underutilization of the simplest measurements assessing respiratory mechanics at the bedside (1). These measurements have limitations, such as the inability to account for regional heterogeneity, chest wall stiffness, patient position, and the effects of spontaneous efforts. Despite these shortcomings, global measures, as described in this review, still form the basis of many of the major advances made in the ventilator management of ARDS, and underpin novel approaches. In this review, we introduce

both the theoretical principles and the technical considerations useful for performing and interpreting these measurements. For interpretation of respiratory mechanics during spontaneous breathing, we refer the reader to recent reviews (30, 31).

Respiratory Mechanics during Passive Ventilation

Static Measurements for the Respiratory System: Rationale and Practice

The equation of motion for the respiratory system, first proposed by Rohrer (32) and based on Newton's third law (33), constitutes the fundamental theory of respiratory mechanics. It characterizes mechanical forces and provides the mathematical foundation for static measurements in clinical practice. During MV, both ventilator and respiratory muscles can apply pressures to the respiratory system. The sum of applied pressures is equal to the sum of opposing pressures, namely, elastic recoil pressure (Pel), flow-resistive pressure (Pres), and inertial pressure (Pin) as follows:

Pvent + Pmus = Pel + Pres + Pin

$$= f_1(\mathbf{V}) + f_2(\dot{\mathbf{V}}) + f_3(\ddot{\mathbf{V}}),$$
(1)

where Pvent is ventilator pressure; Pmus is muscle pressure; f_1 , f_2 , and f_3 are functions describing the relationships between Pel and volume (V), Pres and the rate of change in volume (flow, \dot{V}), and Pin and the rate of change in flow (acceleration, \ddot{V}), respectively. This equation applies during both inspiration and expiration, where inspiratory flow is usually defined as positive and expiratory flow as negative. Ventilator pressure is identical to Paw. By assuming linear relationships in these functions, the motion of the system can be described:

$$Paw + Pmus = Ers \cdot \Delta V + Rrs \cdot \dot{V} + Irs \cdot \ddot{V},$$
(2)

where Ers represents respiratory system elastance (the inverse of compliance), ΔV is the volume difference from the resting volume, **Rrs** represents respiratory system resistance, and Irs represents inertance. Other than in situations such as highfrequency ventilation or coughing, inertance is negligible. During passive ventilation, the respiratory muscles are relaxed and the Pmus is nil. ΔV is the sum of inspired volume (Vinsp) above end-expiratory lung volume (EELV) and EELV above FRC. The product of Ers and EELV above FRC is equivalent to the total Pel at the end of expiration (total positive end-expiratory pressure [PEEPtot], namely, the sum of PEEP and intrinsic PEEP). Equation 2 can then be presented as:

$$Paw = Ers \cdot Vinsp + Rrs \cdot \dot{V} + PEEPtot.$$
(3)

Paw, Vinsp, and \dot{V} are directly monitored or set by the ventilator, whereas Ers, Rrs, and PEEPtot need to be calculated. The use of a constant flow on the ventilator greatly facilitates this calculation. An end-expiratory occlusion (EEO), when Vinsp and V are nil, allows measuring PEEPtot. An endinspiratory occlusion (EIO) after delivering VT allows calculating Paw,EIO, defined as Pplat, and calculating Equations 4 and 5:

$$Paw,EIO = Ers \cdot VT + PEEPtot \tag{4}$$

$$\operatorname{Ers} = (\operatorname{Pplat} - \operatorname{PEEPtot})/\operatorname{Vt.}$$
 (5)

When \dot{V} is constant, Pres will remain approximately constant throughout inspiration. The peak Paw (Ppeak) occurs at the end of inspiration:

$$Ppeak = Ers \cdot VT + Rrs \cdot \dot{V} + PEEPtot.$$
 (6)

Subtracting Equation 4 from Equation 6 allows the calculation of Rrs:

$$Ppeak-Paw, EIO = Rrs \cdot \dot{V}$$
 (7)

$$\frac{\text{Rrs} = (\text{Ppeak} - \text{Pplat})/\dot{\text{V}}.$$
 (8)

Of note, the calculated **Rrs** in Equations 3–8 is **inspiratory resistance**, which can differ from expiratory resistance. The latter can be higher in the absence of PEEP and depends on lung volume (34).

Measuring mechanical properties of the respiratory system requires only limited assumptions and the performance of EEO and EIO maneuvers (Figure 1). As there is no change in VT or V during these maneuvers (by using volume-controlled mode), they are called static measures. Paw under static conditions is equal to alveolar pressure (Palv), but may not reflect Palv in regions with airway closure (frequent in morbidly obese patients [35] or during low PEEP ventilation in supine patients [36–39]). In general, compliance calculated



Figure 1. Static measurements of respiratory mechanics. Essential variables of respiratory mechanics can be measured by performing an end-expiratory occlusion (EEO) and an end-inspiratory occlusion (EIO) for 1–2 seconds or by using volume-controlled ventilation with a constant inspiratory flow and an inspiratory pause time of 0.3 seconds. When EEO and EIO are used, potential leakage needs to be excluded by comparing the difference in VT between the breath with occlusion (*light gray area of flow*) and the one without occlusion (*dark gray area*). $\Delta P = driving pressure as the difference between plateau pressure and positive end-expiratory pressure; <math>\Delta Pel_{,L} =$ the change in lung elastic pressure; $\Delta Pel_{,rs} =$ the change in elastic pressure of respiratory system; Paw = airway pressure; PEEP = positive end-expiratory pressure; PEEPtot = total positive end-expiratory pressure; Pes = esophageal pressure; Pes_Eo = esophageal pressure at end-expiratory occlusion; PL = transpulmonary pressure; PL_Eo = transpulmonary pressure; Pelat, est = estimated plateau pressure; Pres = resistive pressure; VCV = volume-controlled ventilation;

using static Paw (Pplat and PEEPtot) reflects the summed influence of both alveolar units (in parallel) and chest wall (in series). Small airways may also have some compliance, but they contribute with less than 3% of static lung compliance (40, 41).

Static Measurements for Partitioning Lungs and Chest Wall

The measurement of Paw allows the description of the total respiratory system. To differentiate the mechanical properties of the lungs from those of the chest wall requires measuring the pleural pressure (Ppl). The pressure applied on the lungs is the difference between Paw and Ppl, and is called the PL (42). When the respiratory muscles are relaxed, the pressure applied on the chest wall (Pcw) is the difference between Ppl and body surface pressure (Pbs); the pressure applied on the

total respiratory system (Prs) is the difference between Paw and Pbs. Hence,

$$Prs = Paw - Pbs$$

= (Paw - Ppl) + (Ppl - Pbs)
= PL + Pcw. (9)

Pbs is usually atmospheric pressure, which, by convention, is referred to as zero. All of the mechanical properties of the respiratory system can be partitioned into lungs and chest wall by separating pressures. For example, Paw at EIO would be the sum of PL and Pcw:

$$Pplat = PL, EIO + Pcw, EIO.$$
 (10)

The change in the elastic pressure of the respiratory system is the sum of the change in lung elastic pressure and the change in chest wall elastic pressure (Figure 1), and:

$$\operatorname{Ers} = \operatorname{EL} + \operatorname{Ecw},$$
 (11)

where EL and Ecw denote lung and chest wall elastance (the inverse of lung and chest wall compliance). Estimating Ppl with an esophageal balloon catheter has made all these calculations possible (Figure 2) (43, 44). As the esophageal pressure (Pes) obtained through a catheter approximates Ppl, Equations 9 and 10 can be restated by replacing Ppl with Pes. PL is calculated as the difference between Paw and Pes, and reflects the distending pressure of the lung. These measurements are illustrated in Figure 1. Two detailed reviews of the application of Pes, technical instructions, and interpretations have recently been published (43, 44). One important limitation is that the Ppl is not uniform, and a pressure gradient between

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nondependent and dependent parts of the pleural space is enlarged in ARDS. Therefore, even if the Pes accurately measures an intrathoracic pressure, it only reflects the distending pressure at one level of the lung, adjacent to the position of the esophagus (45).

"Static" or "Quasistatic" Measures

The magnitude of elastic pressures generated during tidal ventilation relates to the risk of VILI (29). The resistive pressure component, dissipated across artificial airways and main bronchi, does not seem to cause significant bronchiolar and alveolar wall stress. Thus, when VT is kept constant, the influence of slower or faster inspiratory flow rates, generating higher or lower peak pressures, is expected to be negligible, not changing the effective stress on alveolar walls. Nevertheless, because the lung behavior in mammals is not elastic and linear, as assumed by Equation 3, but instead, *viscoelastic* (46–48), the influence of flow and inspiratory time might be relevant in certain conditions. As described in classical studies (41, 49, 50), a slower inflation, for instance, setting inspiratory flow less than 20 L/min, or the use of long inspiratory pauses (EIO > 0.5 s), would typically promote stress relaxation/adaptation of lung tissues, substantially decreasing the EIO pressure for a given VT (51). The longer the inspiratory pause, the higher the pressure

decay (typically amounting to 2-3 cm H_2O). Multiple phenomena participate in this decay, including surfactant spreading in the liquid lining layer, lung scaffold accommodation, pendelluft between alveolar units, or slow tidal recruitment (50, 52). As a consequence, the recommendation of long inspiratory pauses (2-5 s) may cause marked underestimation of the effective peak pressures reaching alveoli (Palv), even after discounting pressure losses through airways (53, 54). This peak Paly, however, has been directly linked to VILI (55), representing the effective pressure imposed by the ventilator against the respiratory system, being opposed by the elastic tension generated across the alveolar walls and thoracic structures.

It thus makes sense to use a shorter EIO $(\leq 0.5 \text{ s})$ for measuring peak Palv or Pplat, especially to estimate the actual stress applied by the ventilator on lung tissues. This short occlusion would provide estimates of compliance similar to the socalled "dynamic compliance" in classical physiology (56, 57). The term is ambiguous, however ("dynamic compliance" has often referred to calculations using peak Paw, instead of peak Palv), and we prefer to name such estimate as "quasistatic compliance." In brief, whereas an EIO maneuver that is too long may underestimate the risk of VILI, the use of peak proximal Paw would overestimate it, and we propose, instead, the use of short

inspiratory pauses and quasistatic compliance to better define these risks.

Other reasons favor shorter EIO maneuvers: (1) longer occlusions are more affected by imperceptible leaks, increasing errors; (2) when using low VT and inspiratory flows of 50-60 L/min, a 0.3second EIO can be continuously applied, allowing ventilators to display Pplat; and (3) such short, continuously applied inspiratory pauses were shown to increase CO₂ elimination in patients with ARDS (58). Quasistatic or static measurements are relatively similar for the chest wall, an important issue to consider when partitioning the stress across the respiratory system (see subsequent discussion). A long inspiratory pause (2-5 s) may facilitate the calculation of chest wall mechanics by filtering artifacts caused by cardiac oscillation. For equivalent reasons, EEO maneuvers can be applied with a short duration (≤ 1 s), better estimating the effective swings in Palv.

Other methods can estimate compliance and resistance. The "multiple linear regression method" (40, 41) simultaneously finds the values for Ers, Rrs, and PEEPtot that best "explain" the observed tracings of Paw during inspiration and expiration, using fast sampling of flow, volume, and pressure signals (typically ≥ 100 Hz). The method assumes a linear model for the respiratory system (Equation 3). This method works for any controlled ventilatory mode, without the need for any specific maneuver, and matches the quasistatic values obtained during short occlusions, realistically representing the maximum swings in Palv (i.e., the ΔP), or the average values for Ers and Rrs during the whole respiratory cycle (Figure 3) (40, 41).

Airway Resistance and Expiratory Flow Limitation

Small airway (<2 mm in diameter) resistance plays a minimal role in terms of patient-ventilator interaction in ARDS. The values for airway resistance during inspiration have been found to be slightly higher than in normal subjects (59–61), probably secondary to decreased lung size, but the presence of significant resistance or intrinsic PEEP should alert the clinician to the possibility of coincident airway diseases and/or airway closure.

Airflow limitation during expiration, rather than inspiration, has been observed in



Figure 3. Tracings of airway pressure (Paw) in a pig model of acute respiratory distress syndrome (*gray lines*), superimposed with estimated values for alveolar pressure (Palv; *red lines*; Palv was obtained by multiple linear regression technique [MLR] according to the motion equation described in Equation 3). Note that the traditional plateau pressure (Pplat) estimated after a long inspiratory pause causes a slow decay in airway (and alveolar) pressures, due to stress relaxation/adaptation of lung tissues (SR), underestimating peak Palv achieved in previous cycles (under dynamic conditions). Recent studies have suggested that such dynamic swings in Palv (peak minus valley of Palv), representing the effective driving pressure (ΔP), better predict the risk of ventilator-induced lung injury (VILI) than traditional measures. Thus, the traditional Pplat after a long inspiratory pause may underestimate the risk for VILI, justifying the preferential use of short inspiratory pauses (≤ 0.5 s) during measurements of Pplat or ΔP . By using this short procedure, the estimates of Pplat would be closer to Paw immediately after occlusion (P₁) described in classic physiology, and closer to peak Palv estimated by MLR.

patients with ARDS without a history of airway disease (62, 63), particularly under conditions of zero PEEP (34). This may contribute to the development of intrinsic PEEP. Similarly, a negative pressure applied to the expiratory circuit during exhalation failed to augment expiratory flow, suggesting flow limitation and small airway closure probably related to low lung volumes, increased lung weight (superimposed pressures) over the airways, and surfactant deficiency. This was not improved by bronchodilators, but was abolished by a PEEP of 10 cm H₂O. Other authors found intrinsic PEEP slowly increasing after a prolonged EEO, suggesting slow compartments favored by flow limitation (64). The clinical significance of these findings needs further investigation.

Clinical Reference Ranges of Respiratory Mechanics

Interpreting the measured variables of respiratory mechanics requires an

understanding of their physiological meanings as well as reference ranges and/or safe limits. However, the description of respiratory mechanics (particularly for lung and chest wall properties) in large cohorts is lacking.

To provide some reference ranges for clinicians, we derived data from the study by D'Angelo and colleagues (65) of patients undergoing surgery in the supine position (Table 1). Compliances in normal, awake subjects are higher than these ranges (66), but patients undergoing general anesthesia may be more comparable to intensive care unit patients.

The Baby Lung Concept

ARDS was initially conceived of as diffuse homogenous increases in elastance throughout the lung parenchyma. Computer tomography (CT) studies demonstrated that abnormalities in ARDS are mostly regional, with areas of dense consolidation or atelectasis and other areas with normal, or near normal, aeration. This loss of aerated lung is reflected in reduced FRC, which may be as low as 20–30% of theoretical values expected for healthy subjects (67, 68). Gattinoni and colleagues (69), using quantitative CT scan analysis, found that the reduction in global compliance was mostly explained by the loss in aerated volume. This finding was made popular by the concept of "baby lung" (70).

Key insights derived from these observations are that the aerated lung in ARDS has nearly normal regional mechanics and that the ventilated lungs are not stiff, but small (69). Despite normal mechanical properties, the aerated lung in patients with ARDS can have abnormal increases in water permeability and metabolic rate when examined with positron emission tomography (71-73). This suggests that the "normal" lung areas are subject to increased inflammation, potentially in part because they are overventilated to compensate for the great loss of alveolar units. In experimental positron emission tomography/CT studies, simulating long-term MV with low VT and low PEEP, the normal areas, receiving the highest regional ventilation-as compared with collapsed or hyperdistended areaswere the ones presenting greater progression of inflammation (74).

Use of Respiratory Mechanics to Guide Ventilation in ARDS and Minimize VILI at High Lung Volume

Webb and Tierney (24) and, later, the experimental studies of Dreyfuss and colleagues (25) on VILI demonstrated the potential complications associated with high-volume, high-pressure ventilation associated with large swings in PL. To mitigate alveolar distension and volutrauma, rigorous limitation of VT and pressures are recommended (18–22).

VT, Pplat

In the landmark ARDS Network trial, longterm mortality improved when VT was limited to an average of 6 ml/kg of predicted body weight (a surrogate of predicted lung size in normal subjects) and Pplat to less than 30 cm H_2O (18). The Express study

Table 1. Clinical References for Calculation of Respiratory Mechanics

Derived Calculations	Average Value in ARDS*	Estimated <mark>Normal</mark> Values [†]	Explanation of Reference Range
Crs, ml/cm H ₂ O	38	1.6% of VC ml/cm H ₂ O	An average VC in normal subjects is around $\frac{4,000 \text{ ml}}{4,000 \text{ ml}}$ the predicted Crs would be $1.6\% \times 4.000 = 64 \text{ ml/cm H}_{2}O$
CL, ml/cm H ₂ O	55	2.9% of VC ml/cm H₂O	Roughly predicted range: 90–140 ml/cm H ₂ O
Ccw, ml/cm H ₂ O	125	3.6% of VC ml/cm H_2O	Roughly predicted range: 100-200 ml/cm H ₂ O
Ers, cm H ₂ O/L	26	0.62 cm H ₂ O/1% VC (L)	An average VC in normal subjects is 4 L; this gives a predicted Ers as $0.62 \div 4\% = 16$ cm H ₂ O
E∟, cm H₂O/L	18	0.34 cm H ₂ O/1% VC (L)	An average predicted value would be around 9 cm H ₂ O/L
Ecw, cm H ₂ O/L	8	0.28 cm H ₂ O/1% VC (L)	An average predicted value would be around 7 cm H_2O/L
EL/Ers	0.70	0.55	Estimated from predicted values of EL and Ers
Rrs, cm H ₂ O/L/s	—	8–12 at 60 L/min of square- wave inspiratory flow	This reference value is valid for inspiration only and depends on flow rate and the size of endotracheal tube

Definition of abbreviations: ARDS = acute respiratory distress syndrome; Ccw = chest wall compliance; CL = lung compliance; Crs = respiratory system compliance; Ecw = chest wall elastance; EL = lung elastance; EL/Ers = the ratio of lung elastance to respiratory system elastance; Ers = respiratory system elastance; Rrs = respiratory system resistance at inspiration; VC = vital capacity.

*Mean values reported in or derived from Chiumello and colleagues (83), in which 6 ml/kg VT and 5 cm H₂O of positive end-expiratory pressure were used in 24 patients with moderate or severe ARDS.

[†]Mean values reported in or derived from D'Angelo and colleagues (65) on 18 anesthetized paralyzed patients for minor surgery in the supine position. Predicted VC for men (L) = $5.76 \times \text{height}$ (m) $- 0.026 \times \text{age}$ (yr) - 4.34; for women (L) = $4.43 \times \text{height}$ (m) $- 0.026 \times \text{age}$ (yr) - 2.89.

titrated PEEP to reach a Pplat between 28 and 30 cm H_2O at Day 1, increasing PEEPinduced recruitment and limiting overdistension at the same time (26). Compared to a lower PEEP, this strategy reduced the days on ventilator and the days with organ failures, but not mortality. Although the use of small VT and low Pplat has improved mortality in ARDS, there is no clear limit to these variables below which further decreases will not improve outcomes (75).

Pplat is the sum of PEEP or PEEPtot and ΔP . A high Pplat, close to 30 cm H₂O, is an important alarm for the clinician. The incidence of complications, such as pneumothoraces, have markedly decreased since the use of lower Pplat (76). Excessive ΔP , and therefore excessive Pplat, increases the risk of VT-induced strain, and is associated with higher mortality (1, 29). The mechanical effects of high PEEP depends on lung recruitability (77) and can be beneficial (recruitment of previously closed alveoli) or harmful (hyperinflation of previously opened alveoli). Therefore, increase in Pplat resulting from increased PEEP may be associated with different effects on ΔP , making clinical interpretation difficult; when ΔP was stratified, Pplat between around 22-34 cm H₂O was not associated with mortality (1).

ΔΡ

Amato and colleagues (19) proposed that VILI might be due to the swings in pressure

during ventilation rather than an absolute maximum level. This value, known as ΔP , corresponds to the elastic pressure swing, Δ Pel,rs = Δ V × Ers. Δ P equals VT/Crs, and Crs is proportional to FRC (69, 78). ΔP thus describes the relationship between \overline{VT} and the lung volume available to receive the breath. Using a statistical tool known as multilevel mediation analysis to analyze individual data from 3,562 patients with ARDS enrolled in nine randomized trials, it was demonstrated that ΔP was the pulmonary mechanical variable most predictive of 60-day survival in ARDS (1, 29). Another study showed that ΔP and lung stress were closely related (79): a ΔP of 15 predicted a lung stress of 24 cm H₂O, a level that has been associated with VILI (80-83).

Limiting ΔP —possibly keeping it below 14 cm H₂O (1, 29)—can be achieved either by decreasing VT or increasing Crs. VT reduction may require volumes that do not support adequate oxygenation or sufficient CO₂ elimination. Increasing Crs by altering PEEP may not always be achievable. Clinical trials to prospectively test the use of ΔP for optimizing ventilatory management are needed.

Pι

Talmor and coworkers (84–86) tested the principle of titrating PEEP to obtain a positive end-expiratory PL (Figure 1) in a pilot study and in an ongoing multicenter trial. Gattinoni and colleagues (77, 80, 87) used end-inspiratory PL values to guide the upper limit of ventilating pressures and volumes. Gattinoni's calculations were not based on direct measurements of Ppl,EIO, but used the lung elastance-to-Ers ratio, which ignores the value of Ppl at endexpiration at zero PEEP. The estimates for end-inspiratory PL are therefore lower when calculated by the direct measurement of PL, EIO using Pes. Absolute values of Pes are reliable (42, 88), especially after proper correction for esophageal wall compliance and esophageal balloon volume (88), but there is a pressure gradient from the vertebral to the sternal part in the pleural space. So the two methods give different indexes that may reflect different local pressures (45).

Recent studies have also suggested a possibly important role of ΔP for general patients under MV (1), neurological patients (89), patients under extracorporeal membrane oxygenation (90), and patients under general anesthesia (91). The relatively constant value of E_{CW} across patients, even in morbid obesity (92-94), makes Δ Pel,rs a reasonable surrogate of Δ Pel,L in many circumstances. Sometimes, it makes sense to better estimate the cyclic alveolar wall stress through a direct measurement of ΔPel_{L} (95) (instead of Δ Pel,rs), especially in situations of clearly impaired chest wall compliance, abdominal hypertension, or severe scoliosis. The lung ΔP alone (i.e., ΔPL in Figure 1) is the component relevant to VILI (13), and might better surrogate lung strain than ΔP (which includes the elastance of the chest wall and

abdomen). Whether ΔP_L better predicts clinical outcomes than the more easily measured ΔP requires further study (96).

Stress Index

Conventional detection of recruitment and overdistension during MV involves analysis of the change in compliance (represented by changes in the slope of the static pressure-volume curve). Ranieri and colleagues (97) found that, by using constant-flow inflation, the dynamic pressure-volume curve during MV could provide the same information as the static curve. As Pres remains constant during inflation at constant flow, the change in Paw reflects the change in Pel. Because volume is the integral of flow rate over time, the pressure-time curve at a constant flow can be used as a surrogate for the quasistatic pressure-volume curve. Through fitting pressure (either Paw or PL) as a power function of time:

$$pressure = a \cdot time^b + c, \qquad (12)$$

where coefficients *a*, *b*, and *c* are constants, the shape of pressure–time curve can be described by coefficient *b* and termed the "stress index." A convex pattern (b < 0.9) of the pressure–time curve indicates intratidal recruitment, a straight line (0.9 < b < 1.1) indicates linear compliance, and a concave pattern (b > 1.1) indicates hyperinflation (98). An alternative approach has been used by fitting the volume as a quadratic function of pressure, and analyzing the signal of the quadratic coefficient E2 (elastance dependent on volume) to indicate either intratidal recruitment or overdistension (99).

Experimental studies have confirmed these shape thresholds and their association with VILI, but a number of caveats exist regarding interpretation in situations of pleural effusions, high intra-abdominal pressure (100), or heterogeneous lung disease (99). A concave pattern of pressure-time curve may relate to progressive decrease in recruitment rather than specifically overdistension (101). Moreover, the stress index was developed with larger VT values than are currently used, increasing the technical difficulties in obtaining reliable values, especially at high inspiratory flow. Finally, it requires dedicated software. Simply observing the shape of the pressure-time curve during constant flow is sometimes insightful when titrating PEEP or VT at the bedside (Figure 4).

Stress and Strain

Lungs of patients with the same body weight ventilated with the same VT may be subjected to different forces dependent on the size of the functional lung. This is expressed by the mechanical concepts of stress and strain.

Stress is the net force acting on a material structure, for instance, a lung strip connecting the hilum to visceral pleura, causing deformation (i.e., strain), divided by the cross-sectional area of this strip. In respiratory mechanics, lung stress reflects the net distending pressures applied on the lung parenchyma, opposed by the elastic pressures generated by the tensioned alveolar walls and lung scaffold. Stress is equal to PL (Paw – Ppl) under zero flow conditions. Strain represents the deformation experienced by a structure, and is defined as the change in length (Δl) or volume compared with the structure's initial length (l_0) or volume, assumed to be the natural, unstressed condition:

strain =
$$\Delta l/l_0$$
, (13)

and, with respect to lung mechanics:

strain = V_T/FRC . (14)

FRC is here assumed as the nonstressed, resting lung condition. The concept of strain demonstrates how it is possible, with the same ideal body weight and VT, but different FRC (e.g., by disease), to have different risks of injury. The generation of strain above specific levels is associated with clinical and biochemical markers of lung injury. In healthy pigs, MV with strain greater than 1.5-2 was systematically associated with the development of VILI (102). Similarly, in an observational human study, patients with ARDS and high strain showed a fourfold increase of IL-6 and IL-8 concentrations in bronchoalveolar lavage fluid compared with those with normal strain (103). Recently, it was demonstrated that fludeoxyglucose F 18 uptake in the normally aerated tissue (the baby lung) was strongly associated with high strain in those regions (104, 105).

Although measuring FRC and resting lung volume is still challenging (106), altered compliance is a marker of the lung volume reduction caused by disease. As such, ΔP , reflecting the ratio between VTand the functional lung size, provides an indirect approach to estimate VT-induced strain at the bedside.

P_L and Strain

It may be possible to estimate strain from measurements of stress, as the two values are



Figure 4. Stress index. Examination of the late portion of the inspiratory pressure–time curve may provide information on the effect of positive end-expiratory pressure (PEEP) on tidal recruitment. (A) Demonstration of a convex shape, consistent with a stress index less than 1, on a PEEP of 5 cm H₂O. (*B*) Demonstration of a flat shape, consistent with a stress index of 1, on a PEEP of 10 cm H₂O. (*C*) Demonstration of a concave shape, consistent with a stress index greater than 1, on a PEEP of 15 cm H₂O. (*C*) Demonstration of a concave shape, consistent with a stress index greater than 1, on a PEEP of 15 cm H₂O. These patterns are correlated with intratidal alveolar recruitment, stable alveolar mechanics, and tidal hyperinflation, respectively. Paw = airway pressure.

linked by the specific elastance of the lung (ELspec) such that:

strain = stress/ELspec.
$$(15)$$

Thus, if the net distending pressure, PL, is known, strain may be estimated:

strain =
$$PL/ELspec.$$
 (16)

The value of ELSPEC is approximately 13 cm H_2O , a value that varies moderately during disease in the range of tidal ventilation (8, 107). Thus, by measuring transpulmonary stress, strain may be inferred (i.e., $\sim 1/13$ of transpulmonary stress). Recent studies on tissue mechanics, however, have challenged the assumption of a linear relationship between stress and strain, suggesting that bedside estimates of strain are potentially problematic when estimating stresses near the limits of tissue rupture (108).

Use of Respiratory Mechanics to Minimize VILI at Low Lung Volumes

It is supposed that cyclic

recruitment-derecruitment of alveoli may occur at low pressures (109–111). VILI at low lung volume is due to high local tissue stress and parenchymal shear injury supposedly caused by repetitive opening and closing of alveoli and distal small airways (atelectrauma) (14, 15).

In patients with recruitable lung, PEEP increases the amount of aerated lung at end expiration, increasing the number of functional lung units compared with zero end-expiratory pressure, and therefore potentially minimizing strain. Because the concept of strain assumes that there is a resting lung volume (equal to FRC) in which the stresses on alveolar walls are zero, the occurrence of recruitment with PEEP creates a conceptual problem. PEEP has two main effects: (1) unfolding of alveolar walls in previously collapsed alveoli (newly recruited), which become functional, but not necessarily strained; and (2) strain of previously functional and newly functional alveoli. Thus, PEEP will increase the end-expiratory lung volume, generating the so-called PEEPinduced increase in lung volume (V_{PEEP}), but at the same time will increase the "functional FRC" (i.e., the FRC that would be observed if the lung did not recollapse at zero endexpiratory pressure). Part of this VPEEP is an unstressed component that should be subtracted from the true strain. In practice, it is difficult to estimate this newly added (unstressed) lung size, and some simplifications have been proposed (106). Some investigators have added V_{PEEP} ("static strain") to VT ("dynamic strain"), and therefore calculate strain as (VT + V_{PEEP})/FRC (112). Others have calculated strain as VT/EELV, where EELV includes both FRC and V_{PEEP} (113–115). Thus, the latter definition removed the "static" component from the numerator, in accordance with more recent studies showing that static strain forces (from PEEP) may be less injurious than dynamic strain forces (from VT), and may be protective (116, 117).

PEEP-induced lung recruitment has a strong impact on strain calculation: the recruited volume and the new functional FRC generated by PEEP will increase the denominator in strain definitions. Thus, the ability to identify patients with recruitable lung has significant implications with respect to PEEP titration in ARDS. Both CT imaging and pressure-volume curves have demonstrated an acceptable ability to differentiate patients with recruitable lung from those with unrecruitable lung (77, 118–120). Dellamonica and colleagues (106) have recently demonstrated that the comparison of EELV at two different PEEP levels allows the bedside assessment of PEEP-associated lung recruitment. This technique may help guide PEEP titration and strain management in patients with ARDS. For the different methods proposed to titrate PEEP, we refer the reader to another article of the same series (121).

Stress Raisers

The lung parenchyma in ARDS displays apparent heterogeneity in the distribution of consolidation and atelectasis. <u>High regional</u> <u>strain</u> may be exacerbated by the presence of <u>"stress raisers</u>"—<u>interfaces</u> of <u>aerated</u> and <u>nonaerated lung</u> that <u>amplify</u> regional tissue <u>forces</u>.

Mead and coworkers (122, 123) demonstrated that this led to an uneven



Figure 5. A <u>stress raiser</u> is a local area of inhomogeneous tissue that multiplies local stress and strain in the tissues around it when a given stress is directed through it. An equivalent volume of gas is ventilated into normal lung (*A*) and into a region of lung with a <u>stress raiser</u>—a <u>collapsed</u> or <u>consolidated</u> portion that does not participate in ventilation (the *dark gray lung unit* in *C*). An equivalent volume is applied to both lung units in *A* and *C*; however, the <u>portions of lung around the stress raiser (*light gray regions* in *D*) are subjected to greater stress than those around the normally aerated central alveolus in *B*, whereas the size of the collapsed or consolidated portion (the *dark gray lung unit* in *C*) and *D*) does not change size with ventilation. Adapted from Reference 126.</u>

distribution of stress, as well as areas of localized increases in stress (Figure 5). Accordingly, when one of two neighboring lung units decreases in elasticity (due to collapse or consolidation), the stress of the adjacent open unit may increase severalfold (122). The borders between normal and abnormal lung units have been termed "stress raisers"—regions where values of stress greatly exceed global values (123, 124). The concept of stress raisers is well established in structural engineering of inanimate systems (125), and may provide a partial explanation of why ventilating pressures or strains that are benign in healthy subjects may be injurious in patients with ARDS (126). A recent study demonstrated that inhomogeneities responsible for the <u>stress raiser</u> phenomenon represent <u>14–23%</u> of the lung <u>parenchyma in ARDS</u>, and that <u>PEEP</u> decreased their prevalence (123).

Conclusions

The use of small VT, low ΔP , and low Pplat in ARDS is supported by physiological data and controlled trials. The use of <u>PEEP</u> to <u>minimize</u> cyclic <u>derecruitment</u>, atelectrauma, inhomogeneity, and stress raisers has a strong biologic rationale. Despite the improvements in mortality with the use of these concepts, controversy remains with respect to titration of VT and PEEP. Measurement of ΔP , PL, or strain allows individualized VT prescriptions to minimize the risks of VILI. Similarly, bedside assessment of recruitment may allow clinicians to determine patients most likely to benefit from the application of a high PEEP.

Author disclosures are available with the text of this article at www.atsjournals.org.

References

- Bellani G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A, Gattinoni L, van Haren F, Larsson A, McAuley DF, *et al.*; LUNG SAFE Investigators; ESICM Trials Group. Epidemiology, patterns of care, and mortality for patients with acute respiratory distress syndrome in intensive care units in 50 countries. *JAMA* 2016;315:788–800.
- Parsons PE, Eisner MD, Thompson BT, Matthay MA, Ancukiewicz M, Bernard GR, Wheeler AP; NHLBI Acute Respiratory Distress Syndrome Clinical Trials Network. Lower tidal volume ventilation and plasma cytokine markers of inflammation in patients with acute lung injury. *Crit Care Med* 2005;33:1–6. [Discussion, pp. 230–232.]
- Donnelly SC, Strieter RM, Reid PT, Kunkel SL, Burdick MD, Armstrong I, Mackenzie A, Haslett C. The association between mortality rates and decreased concentrations of interleukin-10 and interleukin-1 receptor antagonist in the lung fluids of patients with the adult respiratory distress syndrome. *Ann Intern Med* 1996;125:191–196.
- Belperio JA, Keane MP, Lynch JP III, Strieter RM. The role of cytokines during the pathogenesis of ventilator-associated and ventilatorinduced lung injury. Semin Respir Crit Care Med 2006;27:350–364.
- Windsor AC, Mullen PG, Fowler AA, Sugerman HJ. Role of the neutrophil in adult respiratory distress syndrome. Br J Surg 1993;80:10–17.
- Matthay MA, Zemans RL. The acute respiratory distress syndrome: pathogenesis and treatment. Annu Rev Pathol 2011;6:147–163.
- Falke KJ, Pontoppidan H, Kumar A, Leith DE, Geffin B, Laver MB. Ventilation with end-expiratory pressure in acute lung disease. *J Clin Invest* 1972;51:2315–2323.
- Gattinoni L, Carlesso E, Cadringher P, Valenza F, Vagginelli F, Chiumello D. Physical and biological triggers of ventilator-induced lung injury and its prevention. *Eur Respir J Suppl* 2003;47:15s–25s.
- 9. Petty TL, Newman JH. Adult respiratory distress syndrome. West J Med 1978;128:399–407.
- Kirby RR, Downs JB, Civetta JM, Modell JH, Dannemiller FJ, Klein EF, Hodges M. High level positive end expiratory pressure (PEEP) in acute respiratory insufficiency. *Chest* 1975;67:156–163.
- Nash G, Blennerhassett JB, Pontoppidan H. Pulmonary lesions associated with oxygen therapy and artificial ventilation. N Engl J Med 1967;276:368–374.
- 12. Tremblay LN, Slutsky AS. Ventilator-induced lung injury: from the bench to the bedside. *Intensive Care Med* 2006;32:24–33.
- 13. Slutsky AS, Ranieri VM. Ventilator-induced lung injury. *N Engl J Med* 2013;369:2126–2136.
- Muscedere JG, Mullen JB, Gan K, Slutsky AS. Tidal ventilation at low airway pressures can augment lung injury. *Am J Respir Crit Care Med* 1994;149:1327–1334.
- Gattinoni L, Caironi P. Refining ventilatory treatment for acute lung injury and acute respiratory distress syndrome. *JAMA* 2008;299: 691–693.
- Tremblay LN, Slutsky AS. Ventilator-induced injury: from barotrauma to biotrauma. Proc Assoc Am Physicians 1998;110:482–488.

- Rocco PRM, Dos Santos C, Pelosi P. Pathophysiology of ventilatorassociated lung injury. *Curr Opin Anaesthesiol* 2012;25:123–130.
- The Acute Respiratory Distress Syndrome Network. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. N Engl J Med 2000;342:1301–1308.
- Amato MB, Barbas CS, Medeiros DM, Magaldi RB, Schettino GP, Lorenzi-Filho G, Kairalla RA, Deheinzelin D, Munoz C, Oliveira R, *et al.* Effect of a protective-ventilation strategy on mortality in the acute respiratory distress syndrome. *N Engl J Med* 1998;338: 347–354.
- 20. Stewart TE, Meade MO, Cook DJ, Granton JT, Hodder RV, Lapinsky SE, Mazer CD, McLean RF, Rogovein TS, Schouten BD, et al.; Pressure- and Volume-Limited Ventilation Strategy Group. Evaluation of a ventilation strategy to prevent barotrauma in patients at high risk for acute respiratory distress syndrome. N Engl J Med 1998;338:355–361.
- Brochard L, Roudot-Thoraval F, Roupie E, Delclaux C, Chastre J, Fernandez-Mondéjar E, Clémenti E, Mancebo J, Factor P, Matamis D, et al.; The Multicenter Trial Group on Tidal Volume Reduction in ARDS. Tidal volume reduction for prevention of ventilator-induced lung injury in acute respiratory distress syndrome. Am J Respir Crit Care Med 1998;158:1831–1838.
- 22. Brower RG, Shanholtz CB, Fessler HE, Shade DM, White P Jr, Wiener CM, Teeter JG, Dodd-o JM, Almog Y, Piantadosi S. Prospective, randomized, controlled clinical trial comparing traditional versus reduced tidal volume ventilation in acute respiratory distress syndrome patients. *Crit Care Med* 1999;27:1492–1498.
- 23. Villar J, Kacmarek RM, Pérez-Méndez L, Aguirre-Jaime A. A high positive end-expiratory pressure, low tidal volume ventilatory strategy improves outcome in persistent acute respiratory distress syndrome: a randomized, controlled trial. *Crit Care Med* 2006;34:1311–1318.
- Webb HH, Tierney DF. Experimental pulmonary edema due to intermittent positive pressure ventilation with high inflation pressures: protection by positive end-expiratory pressure. *Am Rev Respir Dis* 1974;110:556–565.
- 25. Dreyfuss D, Soler P, Basset G, Saumon G. High inflation pressure pulmonary edema: respective effects of high airway pressure, high tidal volume, and positive end-expiratory pressure. *Am Rev Respir Dis* 1988;137:1159–1164.
- 26. Mercat A, Richard J-CM, Vielle B, Jaber S, Osman D, Diehl J-L, Lefrant J-Y, Prat G, Richecoeur J, Nieszkowska A, *et al.* Expiratory Pressure (Express) Study Group. Positive end-expiratory pressure setting in adults with acute lung injury and acute respiratory distress syndrome: a randomized controlled trial. *JAMA* 2008;299:646–655.
- 27. Meade MO, Cook DJ, Guyatt GH, Slutsky AS, Arabi YM, Cooper DJ, Davies AR, Hand LE, Zhou Q, Thabane L, *et al.*; Lung Open Ventilation Study Investigators. Ventilation strategy using low tidal volumes, recruitment maneuvers, and high positive end-expiratory pressure for acute lung injury and acute respiratory distress syndrome: a randomized controlled trial. *JAMA* 2008;299:637–645.

- 28. Brower RG, Lanken PN, MacIntyre N, Matthay MA, Morris A, Ancukiewicz M, Schoenfeld D, Thompson BT; National Heart, Lung, and Blood Institute ARDS Clinical Trials Network. Higher versus lower positive end-expiratory pressures in patients with the acute respiratory distress syndrome. N Engl J Med 2004;351:327–336.
- Amato MBP, Meade MO, Slutsky AS, Brochard L, Costa ELV, Schoenfeld DA, Stewart TE, Briel M, Talmor D, Mercat A, *et al.* Driving pressure and survival in the acute respiratory distress syndrome. *N Engl J Med* 2015;372:747–755.
- Brochard L, Slutsky A, Pesenti A. Mechanical ventilation to minimize progression of lung injury in acute respiratory failure. *Am J Respir Crit Care Med* 2017;195:438–442.
- Yoshida T, Fujino Y, Amato MBP, Kavanagh BP. Fifty Years of Research in ARDS. Spontaneous breathing during mechanical ventilation: risks, mechanisms, and management. *Am J Respir Crit Care Med* 2017;195:985–992.
- 32. Rohrer J. Der Zusammenhang der Atemkräfte und ihre Abhängigkeit vom Dehnungszustand der Atmungsorgane. *Pflugers Arch Gesamte Physiol Menschen Tiere* 1916;165:419–444.
- 33. Mead J. Mechanical properties of lungs. Physiol Rev 1961;41:281-330.
- 34. Kondili E, Prinianakis G, Athanasakis H, Georgopoulos D. Lung emptying in patients with acute respiratory distress syndrome: effects of positive end-expiratory pressure. *Eur Respir J* 2002;19:811–819.
- 35. Loring SH, O'Donnell CR, Behazin N, Malhotra A, Sarge T, Ritz R, Novack V, Talmor D. Esophageal pressures in acute lung injury: do they represent artifact or useful information about transpulmonary pressure, chest wall mechanics, and lung stress? J Appl Physiol (1985) 2010;108:515–522.
- Hedenstierna G, Bindslev L, Santesson J, Norlander OP. Airway closure in each lung of anesthetized human subjects. *J Appl Physiol* 1981;50:55–64.
- Hedenstierna G, McCarthy G, Bergström M. Airway closure during mechanical ventilation. *Anesthesiology* 1976;44:114–123.
- Schonfeld SA, Ploysongsang Y. Airway closure and trapped gas during low volume breathing. *Respir Physiol* 1983;51:63–77.
- Rothen HU, Sporre B, Engberg G, Wegenius G, Hedenstierna G. Airway closure, atelectasis and gas exchange during general anaesthesia. *Br J Anaesth* 1998;81:681–686.
- Mancebo J, Calaf N, Benito S. Pulmonary compliance measurement in acute respiratory failure. Crit Care Med 1985;13:589–591.
- Brusasco V, Warner DO, Beck KC, Rodarte JR, Rehder K. Partitioning of pulmonary resistance in dogs: effect of tidal volume and frequency. J Appl Physiol (1985) 1989;66:1190–1196.
- Loring SH, Topulos GP, Hubmayr RD. Transpulmonary pressure: the importance of precise definitions and limiting assumptions. *Am J Respir Crit Care Med* 2016;194:1452–1457.
- 43. Akoumianaki E, Maggiore SM, Valenza F, Bellani G, Jubran A, Loring SH, Pelosi P, Talmor D, Grasso S, Chiumello D, et al.; PLUG Working Group (Acute Respiratory Failure Section of the European Society of Intensive Care Medicine). The application of esophageal pressure measurement in patients with respiratory failure. Am J Respir Crit Care Med 2014;189:520–531.
- 44. Mauri T, Yoshida T, Bellani G, Goligher EC, Carteaux G, Rittayamai N, Mojoli F, Chiumello D, Piquilloud L, Grasso S, *et al.*; PLeUral pressure working Group (PLUG—Acute Respiratory Failure section of the European Society of Intensive Care Medicine). Esophageal and transpulmonary pressure in the clinical setting: meaning, usefulness and perspectives. *Intensive Care Med* 2016;42: 1360–1373.
- 45. Yoshida T, Lima C, Roldan R, Morais CCA, Gomes S, Grieco DL, Richard J-CM, Brochard LJ, Kavanagh BP, Amato MBP. Validation of esophageal pressure by direct measurement of pleural pressure in normal and injured lungs [abstract]. *Am J Respir Crit Care Med* 2017; 195:A7528.
- 46. Similowski T, Levy P, Corbeil C, Albala M, Pariente R, Derenne JP, Bates JH, Jonson B, Milic-Emili J. Viscoelastic behavior of lung and chest wall in dogs determined by flow interruption. *J Appl Physiol* (1985) 1989;67:2219–2229.
- Suki B, Barabási AL, Lutchen KR. Lung tissue viscoelasticity: a mathematical framework and its molecular basis. *J Appl Physiol* (1985) 1994;76:2749–2759.

- Suki B, Bates JH. A nonlinear viscoelastic model of lung tissue mechanics. J Appl Physiol (1985) 1991;71:826–833.
- 49. Bachofen H. Lung tissue resistance and pulmonary hysteresis. J Appl Physiol 1968;24:296–301.
- 50. Fredberg JJ, Stamenovic D. On the imperfect elasticity of lung tissue. J Appl Physiol (1985) 1989;67:2408–2419.
- Barberis L, Manno E, Guérin C. Effect of end-inspiratory pause duration on plateau pressure in mechanically ventilated patients. *Intensive Care Med* 2003;29:130–134.
- Hildebrandt J. Pressure-volume data of cat lung interpreted by a plastoelastic, linear viscoelastic model. J Appl Physiol 1970;28:365–372.
- 53. Santini A, Milesi M, Maraffi T, Pugni P, Andreis D, Cavenago M, Gattinoni M, Protti A. Role of static and dynamic driving airway pressure in the development of ventilator-induced lung injury. *Intensive Care Med Exp* 2016;4:A677.
- 54. Mezidi M, Yonis H, Aublanc M, Lissonde F, Louf-Durier A, Perinel S, Tapponnier R, Richard JC, Guérin C. Effect of end-inspiratory plateau pressure duration on driving pressure. *Intensive Care Med* 2016;4:A1040.
- 55. Protti A, Maraffi T, Milesi M, Votta E, Santini A, Pugni P, Andreis DT, Nicosia F, Zannin E, Gatti S, *et al.* Role of strain rate in the pathogenesis of ventilator-induced lung edema. *Crit Care Med* 2016; 44:e838–e845.
- 56. D'Angelo E, Calderini E, Torri G, Robatto FM, Bono D, Milic-Emili J. Respiratory mechanics in anesthetized paralyzed humans: effects of flow, volume, and time. J Appl Physiol (1985) 1989;67:2556–2564.
- 57. Sullivan KJ, Mortola JP. Dynamic lung compliance in newborn and adult cats. *J Appl Physiol (1985)* 1986;60:743–750.
- Devaquet J, Jonson B, Niklason L, Si Larbi A-G, Uttman L, Aboab J, Brochard L. Effects of inspiratory pause on CO₂ elimination and arterial Pco₂ in acute lung injury. *J Appl Physiol (1985)* 2008;105: 1944–1949.
- Pesenti A, Pelosi P, Foti G, D'Andrea L, Rossi N. An interrupter technique for measuring respiratory mechanics and the pressure generated by respiratory muscles during partial ventilatory support. *Chest* 1992;102:918–923.
- Wright PE, Bernard GR. The role of airflow resistance in patients with the adult respiratory distress syndrome. *Am Rev Respir Dis* 1989; 139:1169–1174.
- 61. Pesenti A, Pelosi P, Rossi N, Virtuani A, Brazzi L, Rossi A. The effects of positive end-expiratory pressure on respiratory resistance in patients with the adult respiratory distress syndrome and in normal anesthetized subjects. *Am Rev Respir Dis* 1991;144:101–107.
- 62. Koutsoukou A, Armaganidis A, Stavrakaki-Kallergi C, Vassilakopoulos T, Lymberis A, Roussos C, Milic-Emili J. Expiratory flow limitation and intrinsic positive end-expiratory pressure at zero positive end-expiratory pressure in patients with adult respiratory distress syndrome. *Am J Respir Crit Care Med* 2000;161:1590–1596.
- 63. Koutsoukou A, Bekos B, Sotiropoulou C, Koulouris NG, Roussos C, Milic-Emili J. Effects of positive end-expiratory pressure on gas exchange and expiratory flow limitation in adult respiratory distress syndrome. *Crit Care Med* 2002;30:1941–1949.
- 64. Vieillard-Baron A, Prin S, Schmitt J-M, Augarde R, Page B, Beauchet A, Jardin F. Pressure–volume curves in acute respiratory distress syndrome: clinical demonstration of the influence of expiratory flow limitation on the initial slope. *Am J Respir Crit Care Med* 2002;165: 1107–1112.
- 65. D'Angelo E, Robatto FM, Calderini E, Tavola M, Bono D, Torri G, Milic-Emili J. Pulmonary and chest wall mechanics in anesthetized paralyzed humans. *J Appl Physiol (1985)* 1991;70:2602–2610.
- 66. Estenne M, Yernault JC, De Troyer A. Rib cage and diaphragm–abdomen compliance in humans: effects of age and posture. J Appl Physiol (1985) 1985;59:1842–1848.
- Rylander C, Högman M, Perchiazzi G, Magnusson A, Hedenstierna G. Functional residual capacity and respiratory mechanics as indicators of aeration and collapse in experimental lung injury. *Anesth Analg* 2004;98:782–789.
- Heinze H, Eichler W. Measurements of functional residual capacity during intensive care treatment: the technical aspects and its possible clinical applications. *Acta Anaesthesiol Scand* 2009;53: 1121–1130.

- Gattinoni L, Pesenti A, Avalli L, Rossi F, Bombino M. Pressure–volume curve of total respiratory system in acute respiratory failure: computed tomographic scan study. *Am Rev Respir Dis* 1987;136:730–736.
- Gattinoni L, Marini JJ, Pesenti A, Quintel M, Mancebo J, Brochard L. The "baby lung" became an adult. *Intensive Care Med* 2016;42: 663–673.
- Kaplan JD, Calandrino FS, Schuster DP. A positron emission tomographic comparison of pulmonary vascular permeability during the adult respiratory distress syndrome and pneumonia. *Am Rev Respir Dis* 1991;143:150–154.
- Sandiford P, Province MA, Schuster DP. Distribution of regional density and vascular permeability in the adult respiratory distress syndrome. *Am J Respir Crit Care Med* 1995;151:737–742.
- 73. Bellani G, Messa C, Guerra L, Spagnolli E, Foti G, Patroniti N, Fumagalli R, Musch G, Fazio F, Pesenti A. Lungs of patients with acute respiratory distress syndrome show diffuse inflammation in normally aerated regions: a [18F]-fluoro-2-deoxy-D-glucose PET/CT study. *Crit Care Med* 2009;37:2216–2222.
- 74. Borges JB, Costa ELV, Bergquist M, Lucchetta L, Widström C, Maripuu E, Suarez-Sipmann F, Larsson A, Amato MBP, Hedenstierna G. Lung inflammation persists after 27 hours of protective Acute Respiratory Distress Syndrome Network Strategy and is concentrated in the nondependent lung. *Crit Care Med* 2015;43:e123–e132.
- Hager DN, Krishnan JA, Hayden DL, Brower RG; ARDS Clinical Trials Network. Tidal volume reduction in patients with acute lung injury when plateau pressures are not high. *Am J Respir Crit Care Med* 2005;172:1241–1245.
- 76. Boussarsar M, Thierry G, Jaber S, Roudot-Thoraval F, Lemaire F, Brochard L. Relationship between ventilatory settings and barotrauma in the acute respiratory distress syndrome. *Intensive Care Med* 2002;28:406–413.
- 77. Gattinoni L, Caironi P, Cressoni M, Chiumello D, Ranieri VM, Quintel M, Russo S, Patroniti N, Cornejo R, Bugedo G. Lung recruitment in patients with the acute respiratory distress syndrome. *N Engl J Med* 2006;354:1775–1786.
- Lumb A, Nunn J, editors. Nunn's applied respiratory physiology. Edinburgh, London: Churchill Livingstone/Elsevier; 2010.
- 79. Chiumello D, Carlesso E, Brioni M, Cressoni M. Airway driving pressure and lung stress in ARDS patients. *Crit Care* 2016;20:276.
- Chiumello D, Carlesso E, Cadringher P, Caironi P, Valenza F, Polli F, Tallarini F, Cozzi P, Cressoni M, Colombo A, *et al.* Lung stress and strain during mechanical ventilation for acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2008;178:346–355.
- Ranieri VM, Brienza N, Santostasi S, Puntillo F, Mascia L, Vitale N, Giuliani R, Memeo V, Bruno F, Fiore T, *et al.* Impairment of lung and chest wall mechanics in patients with acute respiratory distress syndrome: role of abdominal distension. *Am J Respir Crit Care Med* 1997;156:1082–1091.
- Grasso S, Terragni P, Birocco A, Urbino R, Del Sorbo L, Filippini C, Mascia L, Pesenti A, Zangrillo A, Gattinoni L, *et al*. ECMO criteria for influenza A (H1N1)–associated ARDS: role of transpulmonary pressure. *Intensive Care Med* 2012;38:395–403.
- 83. Grasso S, Mascia L, Del Turco M, Malacarne P, Giunta F, Brochard L, Slutsky AS, Marco Ranieri V. Effects of recruiting maneuvers in patients with acute respiratory distress syndrome ventilated with protective ventilatory strategy. *Anesthesiology* 2002;96: 795–802.
- 84. Talmor D, Sarge T, Malhotra A, O'Donnell CR, Ritz R, Lisbon A, Novack V, Loring SH. Mechanical ventilation guided by esophageal pressure in acute lung injury. *N Engl J Med* 2008;359:2095–2104.
- Talmor D, Sarge T, O'Donnell CR, Ritz R, Malhotra A, Lisbon A, Loring SH. Esophageal and transpulmonary pressures in acute respiratory failure. *Crit Care Med* 2006;34:1389–1394.
- 86. Fish E, Novack V, Banner-Goodspeed VM, Sarge T, Loring S, Talmor D. The Esophageal Pressure–Guided Ventilation 2 (EPVent2) trial protocol: a multicentre, randomised clinical trial of mechanical ventilation guided by transpulmonary pressure. *BMJ Open* 2014;4: e006356.
- Gattinoni L, Chiumello D, Carlesso E, Valenza F. Bench-to-bedside review: chest wall elastance in acute lung injury/acute respiratory distress syndrome patients. *Crit Care* 2004;8:350–355.

- Mojoli F, Iotti GA, Torriglia F, Pozzi M, Volta CA, Bianzina S, Braschi A, Brochard L. *In vivo* calibration of esophageal pressure in the mechanically ventilated patient makes measurements reliable. *Crit Care* 2016;20:98.
- 89. Tejerina E, Pelosi P, Muriel A, Peñuelas O, Sutherasan Y, Frutos-Vivar F, Nin N, Davies AR, Rios F, Violi DA, *et al.*; for VENTILA group. Association between ventilatory settings and development of acute respiratory distress syndrome in mechanically ventilated patients due to brain injury. *J Crit Care* 2017;38:341–345.
- 90. Serpa Neto A, Schmidt M, Azevedo LCP, Bein T, Brochard L, Beutel G, Combes A, Costa ELV, Hodgson C, Lindskov C, et al.; ReVA Research Network and the PROVE Network Investigators. Associations between ventilator settings during extracorporeal membrane oxygenation for refractory hypoxemia and outcome in patients with acute respiratory distress syndrome: a pooled individual patient data analysis : mechanical ventilation during ECMO. Intensive Care Med 2016;42:1672–1684.
- 91. Neto AS, Hemmes SNT, Barbas CSV, Beiderlinden M, Fernandez-Bustamante A, Futier E, Gajic O, El-Tahan MR, Ghamdi AA, Günay E, et al.; PROVE Network Investigators. Association between driving pressure and development of postoperative pulmonary complications in patients undergoing mechanical ventilation for general anaesthesia: a meta-analysis of individual patient data. Lancet Respir Med 2016;4:272–280.
- Pirrone M, Fisher D, Chipman D, Imber DAE, Corona J, Mietto C, Kacmarek RM, Berra L. Recruitment maneuvers and positive endexpiratory pressure titration in morbidly obese ICU patients. *Crit Care Med* 2016;44:300–307.
- 93. Hedenstierna G, Santesson J. Breathing mechanics, dead space and gas exchange in the extremely obese, breathing spontaneously and during anaesthesia with intermittent positive pressure ventilation. *Acta Anaesthesiol Scand* 1976;20:248–254.
- Chiumello D, Colombo A, Algieri I, Mietto C, Carlesso E, Crimella F, Cressoni M, Quintel M, Gattinoni L. Effect of body mass index in acute respiratory distress syndrome. *Br J Anaesth* 2016;116: 113–121.
- Baedorf Kassis E, Loring SH, Talmor D. Mortality and pulmonary mechanics in relation to respiratory system and transpulmonary driving pressures in ARDS. *Intensive Care Med* 2016;42:1206–1213.
- 96. Chen L, Xu M, Chen G-Q, Soliman I, Rittayamai N, Sklar M, Shklar O, Martins C, Greco P, Every H, et al. Respiratory mechanics in acute respiratory distress syndrome: variables and indexes associated with clinical outcome [abstract]. Am J Respir Crit Care Med 2016;193: A1839.
- Ranieri VM, Giuliani R, Fiore T, Dambrosio M, Milic-Emili J. Volume–pressure curve of the respiratory system predicts effects of PEEP in ARDS: "occlusion" versus "constant flow" technique. *Am J Respir Crit Care Med* 1994;149:19–27.
- Ranieri VM, Zhang H, Mascia L, Aubin M, Lin CY, Mullen JB, Grasso S, Binnie M, Volgyesi GA, Eng P, *et al*. Pressure-time curve predicts minimally injurious ventilatory strategy in an isolated rat lung model. *Anesthesiology* 2000;93:1320–1328.
- 99. Carvalho AR, Spieth PM, Pelosi P, Vidal Melo MF, Koch T, Jandre FC, Giannella-Neto A, de Abreu MG. Ability of dynamic airway pressure curve profile and elastance for positive end-expiratory pressure titration. *Intensive Care Med* 2008;34:2291–2299.
- 100. Formenti P, Graf J, Santos A, Gard KE, Faltesek K, Adams AB, Dries DJ, Marini JJ. Non-pulmonary factors strongly influence the stress index. *Intensive Care Med* 2011;37:594–600. [Published erratum appears in *Intensive Care Med* 37:727.]
- 101. Hickling KG. The pressure–volume curve is greatly modified by recruitment: a mathematical model of ARDS lungs. Am J Respir Crit Care Med 1998;158:194–202.
- 102. Protti A, Cressoni M, Santini A, Langer T, Mietto C, Febres D, Chierichetti M, Coppola S, Conte G, Gatti S, *et al*. Lung stress and strain during mechanical ventilation: any safe threshold? *Am J Respir Crit Care Med* 2011;183:1354–1362.
- 103. González-López A, García-Prieto E, Batalla-Solís E, Amado-Rodríguez L, Avello N, Blanch L, Albaiceta GM. Lung strain and biological response in mechanically ventilated patients. *Intensive Care Med* 2012;38:240–247.

- 104. Bellani G, Guerra L, Musch G, Zanella A, Patroniti N, Mauri T, Messa C, Pesenti A. Lung regional metabolic activity and gas volume changes induced by tidal ventilation in patients with acute lung injury. *Am J Respir Crit Care Med* 2011;183:1193–1199.
- 105. Wellman TJ, Winkler T, Costa ELV, Musch G, Harris RS, Zheng H, Venegas JG, Vidal Melo MF. Effect of local tidal lung strain on inflammation in normal and lipopolysaccharide-exposed sheep*. *Crit Care Med* 2014;42:e491–e500.
- 106. Dellamonica J, Lerolle N, Sargentini C, Beduneau G, Di Marco F, Mercat A, Richard JCM, Diehl JL, Mancebo J, Rouby JJ, et al. PEEP-induced changes in lung volume in acute respiratory distress syndrome: two methods to estimate alveolar recruitment. *Intensive Care Med* 2011;37:1595–1604.
- 107. De Robertis E, Liu JM, Blomquist S, Dahm PL, Thörne J, Jonson B. Elastic properties of the lung and the chest wall in young and adult healthy pigs. *Eur Respir J* 2001;17:703–711.
- 108. Suki B, Bates JHT. Lung tissue mechanics as an emergent phenomenon. *J Appl Physiol (1985)* 2011;110:1111–1118.
- 109. Bruhn A, Bugedo D, Riquelme F, Varas J, Retamal J, Besa C, Cabrera C, Bugedo G. Tidal volume is a major determinant of cyclic recruitment–derecruitment in acute respiratory distress syndrome. *Minerva Anestesiol* 2011;77:418–426.
- 110. Terragni PP, Rosboch G, Tealdi A, Como E, Menaldo E, Davini O, Gandini G, Herrmann P, Mascia L, Quintel M, et al. Tidal hyperinflation during low tidal volume ventilation in acute respiratory distress syndrome. Am J Respir Crit Care Med 2007;175:160–166.
- 111. Terragni PP, Del Sorbo L, Mascia L, Urbino R, Martin EL, Birocco A, Faggiano C, Quintel M, Gattinoni L, Ranieri VM. Tidal volume lower than 6 ml/kg enhances lung protection: role of extracorporeal carbon dioxide removal. *Anesthesiology* 2009; 111:826–835.
- 112. Protti A, Chiumello D, Cressoni M, Carlesso E, Mietto C, Berto V, Lazzerini M, Quintel M, Gattinoni L. Relationship between gas exchange response to prone position and lung recruitability during acute respiratory failure. *Intensive Care Med* 2009;35:1011–1017.
- 113. Brunner JX, Wysocki M. Is there an optimal breath pattern to minimize stress and strain during mechanical ventilation? *Intensive Care Med* 2009;35:1479–1483.
- Mentzelopoulos SD, Roussos C, Zakynthinos SG. Prone position reduces lung stress and strain in severe acute respiratory distress syndrome. *Eur Respir J* 2005;25:534–544.

- 115. Paula LF, Wellman TJ, Winkler T, Spieth PM, Güldner A, Venegas JG, Gama de Abreu M, Carvalho AR, Vidal Melo MF. Regional tidal lung strain in mechanically ventilated normal lungs. J Appl Physiol 2016; 121:1335–1347.
- 116. Protti A, Votta E, Gattinoni L. Which is the most important strain in the pathogenesis of ventilator-induced lung injury: dynamic or static? *Curr Opin Crit Care* 2014;20:33–38.
- 117. Protti A, Andreis DT, Monti M, Santini A, Sparacino CC, Langer T, Votta E, Gatti S, Lombardi L, Leopardi O, *et al*. Lung stress and strain during mechanical ventilation: any difference between statics and dynamics? *Crit Care Med* 2013;41:1046–1055.
- Rouby J-J, Puybasset L, Nieszkowska A, Lu Q. Acute respiratory distress syndrome: lessons from computed tomography of the whole lung. *Crit Care Med* 2003;31(4 suppl):S285–S295.
- 119. Jonson B, Richard JC, Straus C, Mancebo J, Lemaire F, Brochard L. Pressure–volume curves and compliance in acute lung injury: evidence of recruitment above the lower inflection point. Am J Respir Crit Care Med 1999;159:1172–1178.
- 120. Lu Q, Vieira SR, Richecoeur J, Puybasset L, Kalfon P, Coriat P, Rouby JJ. A simple automated method for measuring pressure-volume curves during mechanical ventilation. *Am J Respir Crit Care Med* 1999;159:275–282.
- 121. Sahetya SK, Goligher EC, Brower RG. Fifty years of research in ARDS: setting positive end-expiratory pressure in the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2017;195: 1429–1438.
- 122. Mead J, Takishima T, Leith D. Stress distribution in lungs: a model of pulmonary elasticity. *J Appl Physiol* 1970;28:596–608.
- 123. Cressoni M, Cadringher P, Chiurazzi C, Amini M, Gallazzi E, Marino A, Brioni M, Carlesso E, Chiumello D, Quintel M, et al. Lung inhomogeneity in patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 2014;189:149–158.
- 124. Rausch SMK, Haberthür D, Stampanoni M, Schittny JC, Wall WA. Local strain distribution in real three-dimensional alveolar geometries. *Ann Biomed Eng* 2011;39:2835–2843.
- 125. Gross D, Hauger W, Schröder J, Wall W, Bonet J. Engineering mechanics 2. Berlin, Heidelberg: Springer; 2011.
- 126. Retamal J, Bergamini BC, Carvalho AR, Bozza FA, Borzone G, Borges JB, Larsson A, Hedenstierna G, Bugedo G, Bruhn A. Non-lobar atelectasis generates inflammation and structural alveolar injury in the surrounding healthy tissue during mechanical ventilation. *Crit Care* 2014;18:505.