

EDITORIAL

Open Access



# Predicting critical illness mortality and personalizing therapy: moving to multi-dimensional data

Zudin A. Puthuchery<sup>1,2\*</sup> and Paul Wischmeyer<sup>3,4</sup>

See related research by Looijaard *et al.*, <http://ccforum.biomedcentral.com/articles/10.1186/s13054-016-1563-3>.

**Keywords:** Skeletal muscle, Intensive care, Mortality

Predicting mortality has been a corner piece of critical care research and practise dating back to the first descriptions of the Acute Physiology and Chronic Health Evaluation score in 1981 [1]. With an increasing burden of non-communicable disease in modern society, pre-existing functional status seems to an important contributor to outcome prediction. Poor physical function is an important predictor of mortality in ambulant diseases. The construct of frailty [2], translated from older patient care into critical care, has provided a useful language to discuss pre-morbid functional status.

Whilst frailty can be established from history taking or hospital coding, both of these methods have clinical and research methodological disadvantages. Wilhelmus *et al.* [3] now offer an alternative approach in a retrospective analysis of computerized tomography scans. Muscle quality as defined by Hounsfield units at the level of L3 was investigated in 491 patients, with a threshold set to define intramuscular adipose tissue and visceral adipose tissue. Higher skeletal muscle density (i.e. better quality) was associated with lower 6-month mortality and shorter hospital length of stay after correction for muscle mass and severity of illness.

Skeletal muscle quality is recognized as a marker of function in healthy individuals [4] and critically ill patients [5]. Alterations are seen with aging [4], immobilization [6], chronic disease states [7] and critical illness [8]. These conditions demonstrate qualitative changes in muscle

structure as a result of increasing collagen and lipid deposition [6]. Intramyocellular lipid accumulation is additionally a hallmark of metabolic diseases, and may exacerbate tissue metabolic derangements in the critically ill.

A limitation of these data is the inability to relate either chronic disease states and poor muscle quality, or muscle quality and functional outcomes. However, decreased skeletal muscle density as an independent predictor of mortality raises the possibility of its use in multi-dimensional scoring systems such as the NUTRIC score [9] or as an alternative marker of chronic poor physiological reserve in the APACHE system.

A number of key roles for early trajectory assessments exist. First, novel early outcome predictors are needed to guide patient and family expectations and decision-making. These need to not only predict risk of death, but also disability, so a patient's wishes may be honoured and a realistic appraisal of functional outcomes can be made. It is vital we improve upon our ability to inform patients and families early in critical illness on the likelihood of significant morbidity. It is possible that admission skeletal muscle quality and quantity may be key to this discussion in the future. Ongoing testing via lean body mass ultrasound [8] and other modalities [10] may also be vital to continued discussions of prognosis. Second, these tests of muscle quality should assist in guiding therapy. A recent post-ICU recovery consensus conference indicated that a major gap exists in understanding how to effectively and efficiently screen patients for specific post-ICU impairments to determine the need for further diagnostic work-up and treatment [11]. Thirdly, the current controversy around personalizing

\* Correspondence: Zudin.puthuchery.09@ucl.ac.uk

<sup>1</sup>Institute of Sports and Exercise Health, University College London Hospitals, 1st Floor, 170 Tottenham Court Road, London W1T 7HA, UK

<sup>2</sup>Division of Critical Care, University College London Hospitals, London, UK  
Full list of author information is available at the end of the article

nutrition delivery in the ICU to optimize outcome [12] has begun to be addressed by early studies validating the role of the aforementioned NUTRIC score in nutrition risk prediction [9]. High malnutrition risk patients may benefit to a greater degree than those with lower risk. A key addition to this prediction of nutrition risk may be muscle quantity and quality at ICU admission. The ability of the muscle to utilize substrate such as lipid and overall glycogen content [10] may be key in delivering personalized nutrition to improve outcomes. Patients with low muscle quality and quantity may have greater and different specific nutritional requirements. Conversely, increased muscle myosteatosis as defined by decreased skeletal muscle density or increased intermuscular adipose tissue may indicate impaired muscle substrate utilization as implied by Wilhelmus et al. [3]. This may indicate that nutrition delivery needs to account for impaired substrate (lipid) utilization and/or measures need to be taken to improve muscle lipid uptake/utilization (e.g. carnitine [13]). Finally, exercise and reduction of immobility are essential to reduce impaired muscle substrate metabolism and thus improve poor muscle quality.

These assessments may be a key innovation prior to major surgery or cancer therapy. Patients with poor skeletal muscle quality could then be enrolled in prehabilitative exercise/nutrition programmes to improve skeletal muscle quality and quantity [14]. Clinical trials systematically evaluating muscle quality and quantity measures via CT scan and ultrasound could then be performed to assess interventions and target ideal methods to optimize patients. Further, in the ICU, these techniques need further study to determine the muscle-level effects of individual nutrition (e.g. protein delivery, anabolic agents [10]) and specific ICU-rehabilitation interventions (e.g. in-bed ergometry, functional electrical stimulation [15]). Current functional testing (i.e. Medical Research Council sum score, hand-grip strength, walk testing) is both volitional and not muscle specific, and has significant implementation, interpretation and compliance challenges. Thus, the role of muscle quality and quantity measurement described here deserves additional study and validation to add an additional “dimension” to our prediction of outcome and personalization of care in the ICU.

#### Abbreviations

APACHE: Acute Physiology and Chronic Health Evaluation; CT: Computerized Tomography; ICU: Intensive care unit; NUTRIC: Nutrition Risk in Critically Ill

#### Acknowledgements

None.

#### Funding

Not applicable.

#### Availability of data and materials

Not applicable.

#### Authors' contributions

Both authors contributed equally. Both authors read and approved the final manuscript.

#### Authors' information

See Authors' contributions.

#### Competing interests

The authors declare that they have no competing interests.

#### Consent for publication

Not applicable.

#### Ethics approval and consent to participate

Not applicable.

#### Author details

<sup>1</sup>Institute of Sports and Exercise Health, University College London Hospitals, 1st Floor, 170 Tottenham Court Road, London W1T 7HA, UK. <sup>2</sup>Division of Critical Care, University College London Hospitals, London, UK. <sup>3</sup>Department of Anesthesiology, Duke University of Medicine, Durham, USA. <sup>4</sup>Duke Clinical Research Institute, Duke University of Medicine, Durham, USA.

Published online: 30 January 2017

#### References

- Knaus WA, Zimmerman JE, Wagner DP, et al. APACHE—acute physiology and chronic health evaluation: a physiologically based classification system. *Crit Care Med*. 1981;9(8):591–7.
- Bagshaw SM, McDermid RC. The role of frailty in outcomes from critical illness. *Curr Opin Crit Care*. 2013;19(5):496–503.
- Wilhelmus GPM, Looijaard IMD, Stapel SN, Girbes ARJ, Twisk JWR, Oudemans-van Straaten HM, Weijs PJM. Skeletal muscle quality as assessed by CT-derived skeletal muscle density is associated with 6-month mortality in mechanically ventilated critically ill patients. *Crit Care*. 2016.
- Watanabe Y, Yamada Y, Fukumoto Y, et al. Echo intensity obtained from ultrasonography images reflecting muscle strength in elderly men. *Clin Interv Aging*. 2013;8:993–8.
- Parry SM, El-Ansary D, Cartwright MS, et al. Ultrasonography in the intensive care setting can be used to detect changes in the quality and quantity of muscle and is related to muscle strength and function. *J Crit Care*. 2015; 30(5):1151. e9–51 e14.
- Parry SM, Puthuchery ZA. The impact of extended bed rest on the musculoskeletal system in the critical care environment. *Extrem Physiol Med*. 2015;4:16.
- McNelly AS, Rawal J, Shrikrishna D, et al. An exploratory study of long-term outcome measures in critical illness survivors: construct validity of physical activity, frailty, and health-related quality of life measures. *Crit Care Med*. 2016;44(6):e362–369.
- Puthuchery ZA, Phadke R, Rawal J, et al. Qualitative ultrasound in acute critical illness muscle wasting. *Crit Care Med*. 2015;43(8):1603–11.
- Rahman A, Hasan RM, Agarwala R, et al. Identifying critically-ill patients who will benefit most from nutritional therapy: further validation of the “modified NUTRIC” nutritional risk assessment tool. *Clin Nutr*. 2016;35(1):158–62.
- Wischmeyer PE, San-Millan I. Winning the war against ICU-acquired weakness: new innovations in nutrition and exercise physiology. *Crit Care*. 2015;19 Suppl 3:S6.
- Needham DM, Davidson J, Cohen H, et al. Improving long-term outcomes after discharge from intensive care unit: report from a stakeholders' conference. *Crit Care Med*. 2012;40(2):502–9.
- Wischmeyer PE. Are we creating survivors ... or victims in critical care? Delivering targeted nutrition to improve outcomes. *Curr Opin Crit Care*. 2016;22(4):279–84.
- Bonafe L, Berger MM, Que YA, et al. Carnitine deficiency in chronic critical illness. *Curr Opin Clin Nutr Metab Care*. 2014;17(2):200–9.
- Carli F, Minnella EM. Preoperative functional assessment and optimization in surgical patient. Changing the paradigm. *Minerva Anesthesiol*. 2016. [Epub ahead of print].
- Parry SM, Berney S, Warrillow S, et al. Functional electrical stimulation with cycling in the critically ill: a pilot case-matched control study. *J Crit Care*. 2014;29(4):695. e1–7.

RESEARCH

Open Access



# Skeletal muscle quality as assessed by CT-derived skeletal muscle density is associated with 6-month mortality in mechanically ventilated critically ill patients

Wilhelmus G. P. M. Looijaard<sup>1,2,6\*</sup>, Ingeborg M. Dekker<sup>3</sup>, Sandra N. Stapel<sup>1,2</sup>, Armand R. J. Girbes<sup>1,2</sup>, Jos W. R. Twisk<sup>4</sup>, Heleen M. Oudemans-van Straaten<sup>1,2</sup> and Peter J. M. Weijs<sup>1,3,5</sup>

## Abstract

**Background:** Muscle quantity at intensive care unit (ICU) admission has been independently associated with mortality. In addition to quantity, muscle quality may be important for survival. Muscle quality is influenced by fatty infiltration or myosteatosis, which can be assessed on computed tomography (CT) scans by analysing skeletal muscle density (SMD) and the amount of intermuscular adipose tissue (IMAT). We investigated whether CT-derived low skeletal muscle quality at ICU admission is independently associated with 6-month mortality and other clinical outcomes.

**Methods:** This retrospective study included 491 mechanically ventilated critically ill adult patients with a CT scan of the abdomen made 1 day before to 4 days after ICU admission. Cox regression analysis was used to determine the association between SMD or IMAT and 6-month mortality, with adjustments for Acute Physiological, Age, and Chronic Health Evaluation (APACHE) II score, body mass index (BMI), and skeletal muscle area. Logistic and linear regression analyses were used for other clinical outcomes.

**Results:** Mean APACHE II score was  $24 \pm 8$  and 6-month mortality was 35.6%. Non-survivors had a lower SMD (25.1 vs. 31.4 Hounsfield Units (HU);  $p < 0.001$ ), and more IMAT (17.1 vs. 13.3  $\text{cm}^2$ ;  $p = 0.004$ ). Higher SMD was associated with a lower 6-month mortality (hazard ratio (HR) per 10 HU, 0.640; 95% confidence interval (CI), 0.552–0.742;  $p < 0.001$ ), and also after correction for APACHE II score, BMI, and skeletal muscle area (HR, 0.774; 95% CI, 0.643–0.931;  $p = 0.006$ ). Higher IMAT was not significantly associated with higher 6-month mortality after adjustment for confounders. A 10 HU increase in SMD was associated with a 14% shorter hospital length of stay.

**Conclusions:** Low skeletal muscle quality at ICU admission, as assessed by CT-derived skeletal muscle density, is independently associated with higher 6-month mortality in mechanically ventilated patients. Thus, muscle quality as well as muscle quantity are prognostic factors in the ICU.

**Trial registration:** Retrospectively registered (initial release on 06/23/2016) at ClinicalTrials.gov: NCT02817646.

**Keywords:** Intensive care unit, Computed tomography, CT, Muscle, Muscle quality, Myosteatosis, Skeletal muscle density, Intermuscular adipose tissue, Mortality, Outcome

\* Correspondence: w.looijaard@vumc.nl

<sup>1</sup>Department of Intensive Care Medicine, VU University Medical Center Amsterdam, De Boelelaan 1117, Amsterdam, The Netherlands

<sup>2</sup>Institute for Cardiovascular Research, VU University Medical Center Amsterdam, De Boelelaan 1117, Amsterdam, The Netherlands

Full list of author information is available at the end of the article



## Background

Muscle wasting is a severe complication of critical illness [1]. Puthuchery et al. reported a steady decrease in skeletal muscle mass of almost 20% during the first 10 days of intensive care unit (ICU) admission [2]. Loss of muscle has been associated with longer duration of mechanical ventilation and higher ICU and hospital mortality [3–5]. If patients survive, they exhibit long-term functional disability with a great impact on quality of life for as long as 5 to 8 years after admission [6–8]. However, many patients already have a low muscle quantity upon admission to the ICU. In two retrospective studies as much as 60–70% of patients had low muscle quantity as assessed on computed tomography (CT) scans on ICU admission, and low muscle quantity at ICU admission was associated with a higher mortality [9, 10].

Not only the quantity, but also the quality of muscle seems important [11]. Along with a decline in muscle mass, fatty infiltration of muscles or myosteatosis has been identified as a possible cause of loss of muscle quality [11]. Myosteatosis can be apparent within muscle fibres and evaluated on CT scans by measuring skeletal muscle density (SMD), or between muscle fibres and evaluated on CT scans by measuring the amount of adipose tissue between muscles (also termed intermuscular adipose tissue or IMAT). A lower SMD was associated with increased lipid infiltration in muscle biopsies and poor clinical outcomes in non-ICU populations [12–14]. Additionally, a recent study in critically ill patients using ultrasound of the quadriceps muscle found that not only a decrease in muscle quantity but also increased muscle echogenicity was related to a decrease in muscle function [15]. An increased amount of IMAT as assessed on CT scans has been associated with decreased muscle function and increased (systemic) inflammation in non-ICU populations [16, 17]. The aim of the present study was to investigate if muscle quality, as assessed by CT-derived SMD and IMAT, is associated with mortality independently of muscle quantity and severity of illness. We hypothesized that low SMD and high IMAT at ICU admission are associated with a poor outcome, independent of the quantity of muscle and severity of illness.

## Methods

### Patients and data

This is a retrospective analysis of CT-derived muscle quality at a single time point at ICU admission in critically ill patients admitted to a mixed medical-surgical ICU of a university hospital from September 2003 to April 2013. Patients were included if they were aged 18 years or older, stayed in the ICU for at least 4 days, required mechanical ventilation during their ICU stay, and had an abdominal CT scan made 1 day before or up to 4 days after admission to the ICU. Patients were

excluded if the CT scan was not eligible for analysis, or if data on body weight or height or the Acute Physiological, Age, and Chronic Health Evaluation (APACHE) II score was missing. By searching the hospital information system for any patients meeting inclusion criteria, we expanded our previously reported cohort of ICU patients [9].

Patient data including age, sex, weight, height, admission diagnosis, APACHE II score, length of ventilation (LOV), ICU length of stay (ICU-LOS) and hospital length of stay (hospital-LOS), discharge destination, and ICU and hospital mortality was obtained from the ICU patient data management system (Metavision; IMDsoft, Tel-Aviv, Israel) and the hospital information system (Mirador; iSOFT Nederland BV, Leiden, The Netherlands). If mortality data were not registered, these were collected from the civil registry or from the general practitioner.

### CT scan analysis

The precision of single slice CT scan analysis at the third lumbar vertebra (L3) level is high (inter- and intra-observer variability less than 2% in healthy volunteers) [18]. Both skeletal muscle area ( $r = 0.83$ – $0.99$ ;  $p < 0.01$ ) and IMAT ( $r = 0.39$ – $0.61$ ;  $p < 0.05$ ) at this level are closely related to whole body skeletal muscle and IMAT volumes as assessed by magnetic resonance imaging (MRI) [19–21].

CT scans made 1 day before to 4 days after ICU admission for diagnostic purposes were imported from the hospital radiology system and stored on a secure computer system. Scans were analysed using Slice-O-matic versions 4.3 and 5.0 (TomoVision, Montreal, QC, Canada) by two trained and certified investigators (WGPML and IMD, trained by the Cross Cancer Institute, Edmonton, AB, Canada) who had frequent consultation with each other if there was any doubt about eligibility, landmarking, or analysis.

The CT scans were analysed for eligibility and rejected if the scan quality was too low for analysis or if they contained artefacts, or if muscle was cut off due to windowing. Landmarking was performed by identifying the L3 and isolating the CT slice that depicted the whole vertebra the best. A bony landmark was used to ensure reproducibility and consistency between patients.

Different tissues were identified using boundaries in Hounsfield Units (HU) set to  $-29$  to  $+150$  for muscle,  $-190$  to  $-30$  for IMAT and subcutaneous adipose tissue, and  $-150$  to  $-50$  for visceral adipose tissue [22]. SMD was assessed by the mean radiological muscle attenuation of all muscle visible at the L3 level, measured in HU. The HU scale is a radiological scale describing the density of tissues on CT scans [23]. Lower mean muscle attenuation indicates less dense muscle tissue with more lipid infiltration, e.g. lower SMD, while a higher mean muscle

attenuation indicates denser muscle tissue with less lipid infiltration, e.g. higher SMD [14]. IMAT was assessed by identifying all visible adipose tissue within muscle fascia in  $\text{cm}^2$  [22]. Previously found ICU-specific optimal cut-off points related to hospital mortality were used to define low skeletal muscle area: below  $170 \text{ cm}^2$  for male patients and below  $110 \text{ cm}^2$  for female patients [9]. See Fig. 1 for an example of CT scan analysis.

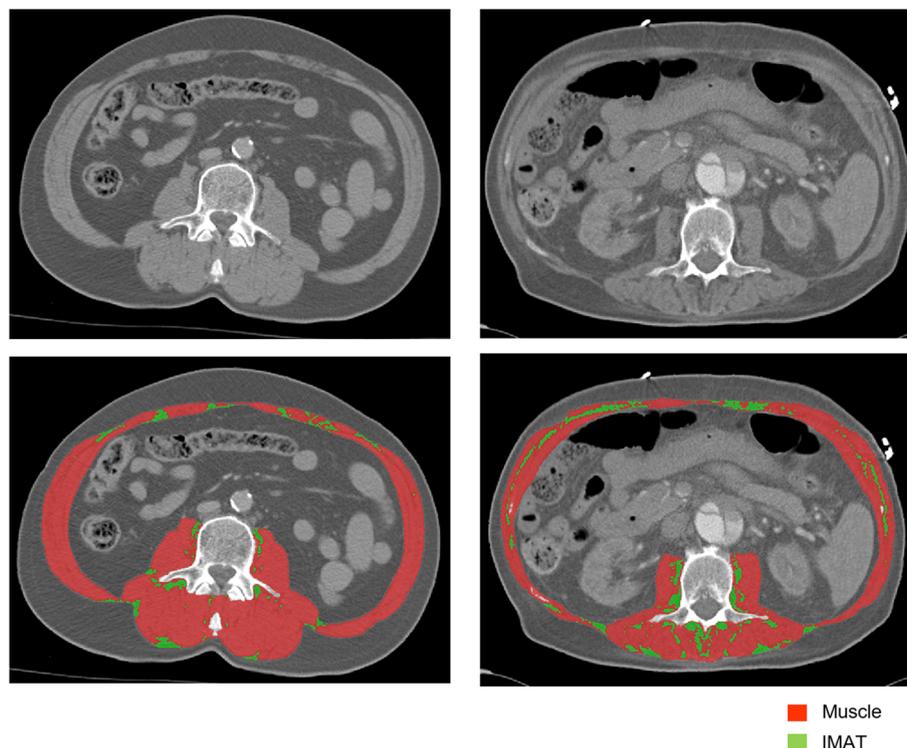
Because muscle quality is important for dealing with recovery after ICU and hospital discharge, we chose 6-month mortality as the primary endpoint. Secondary endpoints were the odds of being discharged from the hospital to home, length of ventilation, and ICU and hospital LOS in survivors.

### Statistics

Independent sample *t* tests were used to compare survivors and non-survivors for normally distributed continuous variables, and Mann-Whitney *U* tests for non-normally distributed continuous variables. Fisher exact and  $\text{Chi}^2$  tests with post-hoc Bonferroni analysis were used to compare survivors and non-survivors for categorical variables. Kaplan-Meier plots were made to visualize the effect of SMD and IMAT (divided into two groups based on the median) on 6-month mortality, with

log-rank tests to compare the survival curves of the two groups. Cox regression analysis was used to evaluate the association between SMD or IMAT (as continuous variables) and 6-month mortality. After univariable analyses, APACHE II score was added to the models to adjust for severity of illness (model 2). In the second adjusted model, body mass index (BMI), and skeletal muscle area were included as well (model 3). Age is included in the APACHE II score and was therefore not separately included in the adjusted models. Additionally, we performed analyses on the subgroup of patients with available data on visceral and subcutaneous adipose tissue in which BMI was substituted with visceral and subcutaneous adipose tissue as a measure of total body fatness (model 4).

Logistic and linear regression analyses were used to evaluate the association between SMD or IMAT and the secondary outcome measures discharge to home, LOV, ICU-LOS, and hospital-LOS in survivors. LOV, ICU-LOS, and hospital-LOS were non-normally distributed and positively skewed; therefore, the analysis was performed on the natural logarithm of the variables. By re-transforming by using the inverse, the influence of a given predictor was calculated as a percentage change in outcome.



**Fig. 1** Example of CT scan analysis. This image shows CT scans at the level of lumbar vertebra 3 of two patients both un-analysed (*upper row*) and analysed (*lower row*). The analysed images show muscle tissue (*red*) and intermuscular adipose tissue (*IMAT, green*). The patient on the left has more muscle ( $165$  vs.  $120 \text{ cm}^2$ ), less IMAT ( $10$  vs.  $19.5 \text{ cm}^2$ ), and higher mean skeletal muscle density ( $42$  vs.  $18$  Hounsfield Units) than the patient on the right

IBM SPSS Statistics 22 (IBM Corp, Armonk, NY, USA) was used for statistical analysis. Values are reported as mean  $\pm$  standard deviation (SD) or median and 25–75% interquartile range (IQR). All statistical tests were two-sided. A  $p < 0.05$  was considered statistically significant.

## Results

A total of 13,434 patients were admitted to the ICU during the study period with a mean APACHE II score of  $17.4 \pm 9.2$ . Six hundred and seventy-eight patients fulfilled inclusion criteria and had their CT scans imported from the radiology system to be analysed for eligibility. CT scans that were found not to be eligible were due to artefacts (78 scans), muscle cut-off (50 scans), or low quality (47 scans). Finally, 491 patients (72%) with complete clinical data and good quality CT scans were included for the statistical analysis. However, due to windowing or artefacts, visceral and/or subcutaneous adipose tissue could not be analysed in 154 patients. We therefore performed subgroup analyses that included visceral and subcutaneous adipose tissue in a subgroup of 337 patients (50%). Figure 2 is the consort diagram showing the inclusion process.

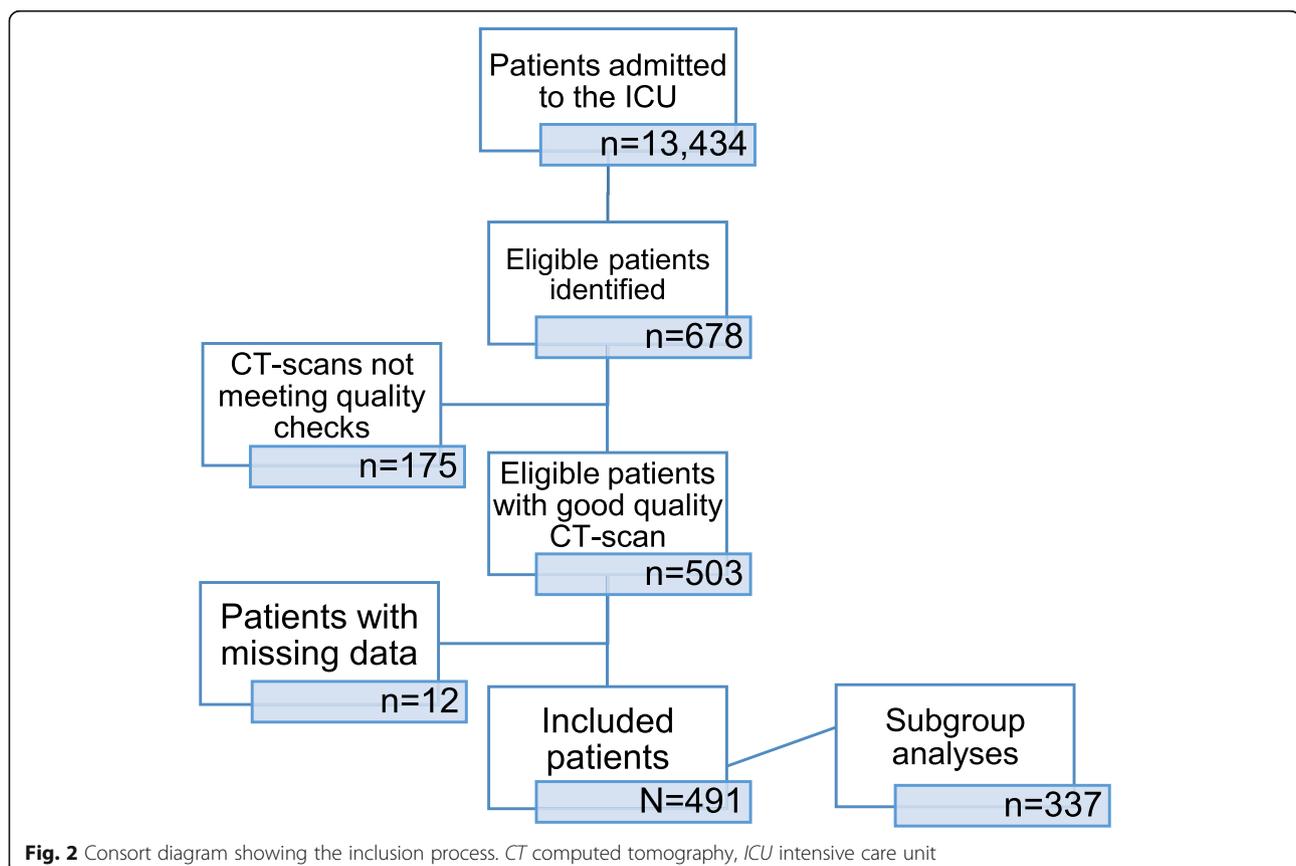
## Patient characteristics

Patient characteristics are presented in Table 1 for 6-month survivors and non-survivors. Outcome measures are presented separately in Table 2. CT scans were mostly made on the day of admission to the ICU. Three hundred and twelve (64.7%) patients had a low skeletal muscle area at ICU admission. Six-month mortality was 35.6%. Non-survivors were older ( $67 \pm 14$  vs.  $55 \pm 18$  years;  $p < 0.001$ ), had a lower BMI ( $24.6 \pm 4.3$  vs.  $25.5 \pm 4.4$  kg/m<sup>2</sup>;  $p = 0.042$ ), higher APACHE II score ( $27 \pm 8$  vs.  $22 \pm 8$ ;  $p < 0.001$ ), and were more often medical patients (62% vs. 43%;  $p < 0.001$ ) than survivors.

Mean SMD at ICU admission was  $29.9 \pm 11.7$  HU. Median IMAT at ICU admission was 13.6 (8.4–24.3) cm<sup>2</sup>, comprising 9.1% of total tissue within muscle fascia (skeletal muscle area plus IMAT) at the L3 level. Non-survivors had a lower skeletal muscle area ( $120.3 \pm 33.0$  vs.  $143.5 \pm 38.9$  cm<sup>2</sup>;  $p < 0.001$ ), lower SMD ( $25.1 \pm 9.4$  vs.  $31.4 \pm 11.7$  HU;  $p < 0.001$ ), and more IMAT ( $17.1$  (10.5–27.1) vs.  $13.3$  (7.9–23.2) cm<sup>2</sup>;  $p = 0.004$ ) than survivors.

## Association between muscle quality and 6-month mortality

Mortality was significantly higher in patients with low muscle quality with SMD values below the median or IMAT values above the median (Fig. 3).



**Table 1** Patient characteristics of all patients and comparison between survivors and non-survivors

	All patients N = 491		Survivors <sup>1</sup> (n = 299)		Non-survivors <sup>1</sup> (n = 165)		P value survivors vs. non-survivors
	Mean/median/n	SD/IQR/%	Mean/median/n	SD/IQR/%	Mean/median/n	SD/IQR/%	
Age, years	58	±18	55	±18	67	±14	<b>&lt;0.001</b>
Sex, male, n (%)	305	62%	191	64%	93	56%	0.135
BMI, kg/m <sup>2</sup>	25.2	±4.3	25.5	±4.4	24.6	±4.3	<b>0.042</b>
Underweight <sup>2</sup> , n (%)	19	4.1%	11	3.7%	8	4.8%	0.291
Normal weight <sup>2</sup> , n (%)	238	51.3%	145	48.5%	93	56.4%	
Overweight <sup>2</sup> , n (%)	158	34.1%	108	36.1%	50	30.3%	
Obesity <sup>2</sup> , n (%)	49	10.6%	35	11.7%	14	8.5%	
APACHE II score	24	±8	22	±8	27	±8	<b>&lt;0.001</b>
Admission category, n (%)							<b>0.001</b>
Medical	248	50.5%	130	43%	102	62%	
Surgical	243	49.5%	169	57%	63	38%	
Admission diagnosis, n (%)							<b>&lt;0.001</b>
Cardiovascular	32	6.5%	18 <sup>a</sup>	6.0%	14 <sup>a</sup>	8.5%	
Metabolic/renal	15	3.1%	8 <sup>a</sup>	2.7%	6 <sup>a</sup>	3.6%	
Neurologic	41	8.4%	19 <sup>a</sup>	6.4%	16 <sup>a</sup>	9.7%	
Post-resuscitation	28	5.7%	16 <sup>a</sup>	5.4%	11 <sup>a</sup>	6.7%	
Post-surgery	149	30.3%	95 <sup>a</sup>	31.8%	50 <sup>a</sup>	30.3%	
Respiratory insufficiency	68	13.8%	40 <sup>a</sup>	13.4%	25 <sup>a</sup>	15.2%	
Sepsis	31	6.3%	14 <sup>a</sup>	4.7%	15 <sup>a</sup>	9.1%	
Trauma	94	19.1%	74 <sup>a</sup>	24.7%	13 <sup>b</sup>	7.9%	
Other	33	6.7%	15 <sup>a</sup>	5.0%	15 <sup>a</sup>	9.1%	
Length of hospital stay before ICU admission, days	0	0–4	0	0–4	0	0–6	0.166
Time from ICU admission to CT scan, days	0	0–1	0	0–1	0	0–1	0.277
Skeletal muscle area, cm <sup>2</sup>	136.5	±39.0	143.5	±38.9	120.3	±33.0	<b>&lt;0.001</b>
Skeletal muscle index, cm <sup>2</sup> /m <sup>2</sup>	44.8	±11.0	46.6	±10.6	40.4	±9.9	<b>&lt;0.001</b>
Low skeletal muscle area <sup>3</sup> , n (%)	312	63.5%	163	54.5%	137	83.0%	<b>&lt;0.001</b>
SMD, HU	29.9	±11.7	31.4	±11.7	25.1	±9.4	<b>&lt;0.001</b>
IMAT, cm <sup>2</sup>	13.6	8.4–24.3	13.3	7.9–23.2	17.1	10.5–27.1	<b>0.004</b>
Visceral adipose tissue, cm <sup>2</sup> (n = 337)	96.7	49.3–170.6	95.8	50.9–178.1	108.1	54.1–177.5	0.593
Subcutaneous adipose tissue, cm <sup>2</sup> (n = 337)	132.7	90.2–182.4	133.7	89.8–189.2	127.7	95.7–176.2	0.440

<sup>1</sup>Survivors and non-survivors 6 months after ICU admission<sup>2</sup>WHO categories: underweight, BMI <18.5; normal weight: BMI 18.5–24.9; overweight: BMI 25–29.9; obesity: BMI ≥30 [42]<sup>3</sup>Defined by skeletal muscle area: <170 cm<sup>2</sup> for males and <110 cm<sup>2</sup> for females [9]<sup>a, b</sup>Values in the same row not sharing the same superscript letter are significantly different in a post-hoc Bonferroni analysis

Values in bold indicate statistically significant p values

APACHE Acute Physiological, Age, and Chronic Health Evaluation, BMI, body mass index, CT computed tomography, HU Hounsfield Units, ICU intensive care unit, IMAT intermuscular adipose tissue, IQR interquartile range, SD standard deviation, SMD skeletal muscle density

Cox regression analysis showed that higher SMD was associated with lower 6-month mortality (hazard ratio (HR) per 10 HU, 0.640; 95% confidence interval (CI), 0.552–0.742;  $p < 0.001$ ; Table 3). This association was still apparent when SMD was adjusted for the confounders

APACHE II score, BMI, and skeletal muscle area (HR per 10 HU, 0.774; 95% CI, 0.643–0.931;  $p = 0.006$ ).

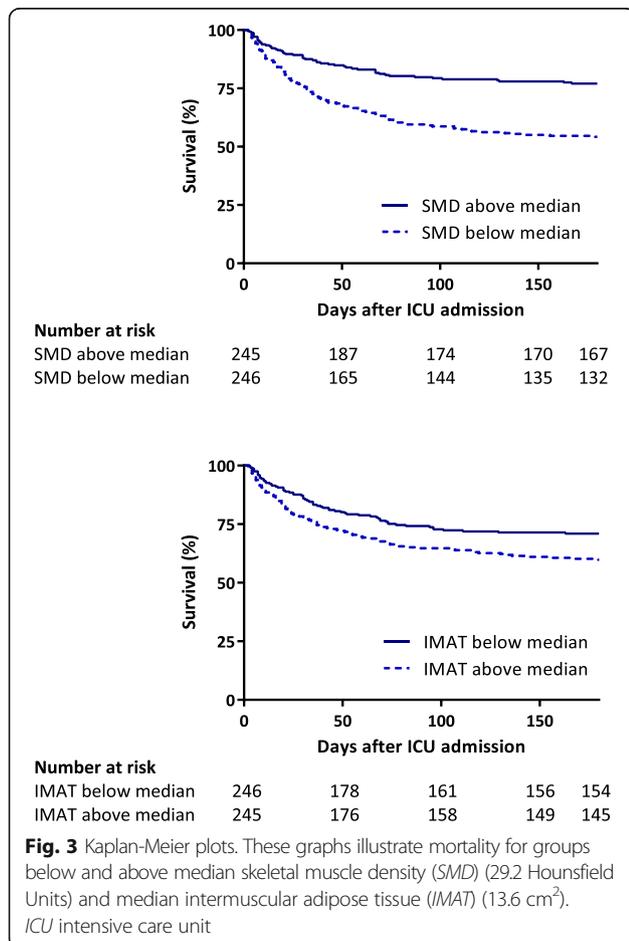
Cox regression analysis showed that higher IMAT was associated with higher 6-month mortality (HR per 10 cm<sup>2</sup>, 1.153; 95% CI, 1.042–1.277;  $p = 0.006$ ). However, when

**Table 2** Primary and secondary outcome measures

	n	%	Days	IQR
Six-month mortality	165	35.6%		
ICU mortality	84	17.1%		
Hospital mortality	132	26.9%		
Length of ventilation			11	6–20
ICU length of stay			13	7–23
Hospital length of stay			35	19–59
Destination after discharge				
Home	144	40.7%		
Other hospital	80	22.6%		
Nursing home	76	21.5%		
Rehabilitation unit	45	12.7%		
Other	9	2.5%		

ICU intensive care unit, IQR interquartile range

adjusted for APACHE II score alone or the confounders APACHE II score, BMI, and skeletal muscle area the association between IMAT and 6-month mortality was not significant (HR per 10 cm<sup>2</sup>, 1.092; 95% CI, 0.966–1.236; *p* = 0.159).



**Fig. 3** Kaplan-Meier plots. These graphs illustrate mortality for groups below and above median skeletal muscle density (SMD) (29.2 Hounsfield Units) and median intermuscular adipose tissue (IMAT) (13.6 cm<sup>2</sup>). ICU intensive care unit

**Analyses in the subgroup with visceral and subcutaneous adipose tissue**

Additional Cox regression analyses were performed in the subgroup of patients with available data on visceral and subcutaneous adipose tissue (*n* = 337, Table 3). Patients in this subgroup were significantly different from patients in whom visceral and/or subcutaneous adipose tissue could not be analysed. They were younger (56 vs. 64 years; *p* < 0.001), more often male (66 vs. 55%; *p* = 0.021), and had a lower BMI (24.8 vs. 25.9 kg/m<sup>2</sup>; *p* = 0.026). In this subgroup, we found both SMD (HR per 10 HU, 0.623; 95% CI 0.524–0.739; *p* < 0.001) and IMAT (HR per 10 cm<sup>2</sup>, 1.245; 95% CI, 1.106–1.401; *p* < 0.001) were significantly associated with 6-month mortality. In multivariable analyses both SMD (HR per 10 HU, 0.728; 95% CI, 0.571–0.928; *p* = 0.010) and IMAT (HR per 10 cm<sup>2</sup>, 1.244; 95% CI, 1.048–1.476; *p* = 0.012) remained significantly associated with 6-month mortality, adjusted for APACHE II score, skeletal muscle area, and visceral and subcutaneous adipose tissue.

**Secondary outcome measures in survivors**

Higher SMD was significantly associated with shorter hospital-LOS after adjustment for APACHE II score, BMI, and skeletal muscle area (Table 4). After re-transformation we found that 10 HU higher SMD was associated with a 14% shorter hospital-LOS. IMAT was not associated with hospital LOS. Neither SMD nor IMAT were significantly associated with the odds of being discharged to home, LOV, or ICU-LOS.

**Discussion**

This retrospective study in mechanically ventilated patients admitted to the ICU for 4 days or longer shows that low skeletal muscle quality at ICU admission, as assessed by skeletal muscle density on CT scans, is associated with higher 6-month mortality independent of muscle quantity, APACHE II score, and BMI. A lower SMD was also associated with a longer hospital stay in survivors. This is the first study investigating the relation between CT-derived markers for muscle quality and outcome in ventilated critically ill patients. Intermuscular adipose tissue was also associated with mortality but not independently, suggesting that SMD is a stronger marker of muscle quality for 6-month mortality or that IMAT is better represented by confounders than SMD.

**Muscle quality and quantity**

Previously we have found that low muscle quantity as assessed by skeletal muscle area on CT scans at ICU admission is a risk factor for hospital mortality, independent of sex and APACHE II score [9]. These findings were in line with a study by Moisey et al. in elderly injured ICU patients, who found low skeletal muscle area

**Table 3** Cox regression: association between skeletal muscle density or intermuscular adipose tissue and mortality

6-month mortality	Univariable N = 491			Model 2 N = 491			Model 3 N = 491			Model 4 (n = 337)		
	HR	95% CI	P value	HR	95% CI	P value	HR	95% CI	P value	HR	95% CI	P value
SMD (per 10 HU)	0.640	0.552–0.742	<b>&lt;0.001</b>	0.703	0.605–0.818	<b>&lt;0.001</b>	0.774	0.643–0.931	<b>0.006</b>	0.728	0.571–0.928	<b>0.010</b>
IMAT (per 10 cm <sup>2</sup> )	1.153	1.042–1.277	<b>0.006</b>	1.092	0.980–1.217	0.110	1.092	0.966–1.236	0.159	1.244	1.048–1.476	<b>0.012</b>

Model 2: adjusted for APACHE II score

Model 3: adjusted for APACHE II score, skeletal muscle area, and BMI

Model 4 (subgroup analysis): adjusted for APACHE II score, skeletal muscle area, visceral adipose tissue, and subcutaneous adipose tissue

Values in bold indicate statistically significant *p* values

APACHE Acute Physiological, Age, and Chronic Health Evaluation, CI confidence interval, HR hazard ratio, HU Hounsfield Units, IMAT intermuscular adipose tissue, SMD skeletal muscle density

to be associated with higher mortality and less ventilator-free and ICU-free days [10]. In the present study, we found that the quality of muscle appeared to be important for survival in addition to quantity.

The APACHE II score is the best validated prognostic ICU score for hospital mortality incorporating age, comorbidities, and acute illness. However, it appears that, independently of APACHE II score, a poor health status as reflected by low muscle quantity and quality (whether due to inactivity, comorbidity, or high age) are important prognostic markers. Unfortunately, the updated APACHE III and IV scores were not available for all patients.

Of interest, IMAT was independently associated with 6-month mortality in a subgroup, but not in the entire cohort. The patients in the subgroup were younger, more often male, and had a lower BMI. Apparently,

visceral tissue on CT scans can more often not be analysed in older patients with high BMI, mostly because a part of the scan is often cut-off in the windowing process.

#### Causes and consequences of myosteatorsis

Previous studies have shown that inactivity, as seen in pre-existing illness and advancing age, can cause an increase in myosteatorsis (as seen by a decrease in SMD and an increase in IMAT) and that these changes are associated with decreased muscle strength [24–26]. During inactivity there is a decrease in lipoprotein lipase activity, the rate-limiting enzyme in triglyceride metabolism, which hydrolyses triglycerides into lipoproteins [27, 28]. Additionally, during bed rest a decrease in 3-hydroxyacyl-CoA-dehydrogenase concentration is seen, which impairs

**Table 4** Logistic and linear regression: association between skeletal muscle density or intermuscular adipose tissue and secondary outcomes

	Univariable			Model 2			Model 3		
	OR/B	95% CI	P value	OR/B	95% CI	P value	OR/B	95% CI	P value
Discharge to home									
SMD (per 10 HU)	1.039	0.864 to 1.250	0.683	0.990	0.816 to 1.200	0.915	0.926	0.718 to 1.195	0.556
IMAT (per 10 cm <sup>2</sup> )	0.886	0.741 to 1.059	0.182	0.912	0.761 to 1.093	0.317	0.884	0.715 to 1.093	0.254
Length of ventilation									
SMD (per 10 HU)	-0.038	-0.107 to 0.032	0.292	-0.003	-0.075 to 0.069	0.936	-0.018	-0.111 to 0.075	0.705
IMAT (per 10 cm <sup>2</sup> )	0.050	-0.014 to 0.115	0.126	0.029	-0.036 to 0.094	0.384	0.026	-0.049 to 0.101	0.499
Length of ICU stay									
SMD (per 10 HU)	-0.051	-0.123 to 0.020	0.158	-0.020	-0.092 to 0.053	0.598	-0.032	-0.128 to 0.063	0.506
IMAT (per 10 cm <sup>2</sup> )	0.064	-0.003 to 0.130	0.059	0.043	-0.023 to 0.110	0.199	0.041	-0.036 to 0.119	0.292
Length of hospital stay									
SMD (per 10 HU)	-0.123	-0.192 to -0.054	<b>0.001</b>	-0.112	-0.184 to -0.041	<b>0.002</b>	-0.134	-0.228 to -0.040	<b>0.005</b>
IMAT (per 10 cm <sup>2</sup> )	0.075	0.010 to 0.140	<b>0.023</b>	0.065	-0.001 to 0.131	0.052	0.064	-0.012 to 0.141	0.100

Model 2: adjusted for APACHE II score

Model 3: adjusted for APACHE II score, skeletal muscle area, and BMI

Discharge to home results are given as OR; length of ventilation, ICU, and hospital stay are given as B values

Values in bold indicate statistically significant *p* values

APACHE Acute Physiological, Age, and Chronic Health Evaluation, B beta coefficient, CI confidence interval, HU Hounsfield Units, ICU intensive care unit, IMAT intermuscular adipose tissue, OR odds ratio, SMD skeletal muscle density

the muscle's ability to metabolize free fatty acids to acyl-CoA [29, 30]. Finally, denervation causes an increase in malonyl-CoA concentrations, which in turn inhibits the rate-limiting enzyme responsible for transporting acyl-CoA into the mitochondria [31]. These altered metabolic mechanisms associated with inactivity decrease the ability of muscles to oxidise lipids and promotes a shift in muscle fuel utilisation from lipids towards glucose, causing accumulation of lipids in the muscle [26, 32]. Manini et al. found that 4 weeks of lower limb immobilisation in healthy adults caused an increase in IMAT and a loss in muscle strength independent of a decrease in muscle mass [26]. Their findings support the idea that myosteatosis is related to decreased muscle quality.

Adipose tissue has been noted as a major endocrine organ. To date, hundreds of adipokines, cytokines secreted by adipose tissue, have been identified [33]. Myosteatosis is associated with an upregulation of macrophage and T-cell expression [34]. These inflammatory cells produce pro-inflammatory cytokines such as tumour necrosis factor- $\alpha$  (TNF $\alpha$ ) and interleukin-6 (IL-6) [35] which mediate contractile dysfunction [36, 37] and create a low-grade inflammatory environment in which the metabolic syndrome, cardiovascular disease, and insulin resistance are prone to develop [16, 17, 34].

### Muscle wasting and long-term outcome

Previous studies have shown that muscle wasting as occurring *during* critical illness has a large impact on survival, successful weaning from ventilation, and long-term functioning [3–8, 38]. Herridge et al. found functional disability in survivors of acute respiratory distress syndrome as much as 5 years after admission to the ICU [7] and Iwashyna et al. found functional limitations up to 8 years after severe sepsis [8]. A decrease in muscle quality as assessed by CT scans has been described in 15 patients in a small substudy of the EPaNIC trial where a substantial decrease in skeletal muscle area and SMD, and an increase in IMAT developing over a 7-day period during the early stage of critical illness was found [39]. In two observational studies including 136 and 115 patients requiring at least 5 and 7 days of mechanical ventilation, respectively, muscle weakness acquired during critical illness was associated with increased ICU and hospital mortality [3, 38]. Our study found that low muscle quality present at the beginning of critical illness was already associated with poor outcome, before the devastating effects of critical illness on muscle wasting.

### Strengths and limitations

Our study has strengths and limitations. This is the first study up to now investigating the relation between muscle quality assessed with CT scans and clinical outcomes in a large group of critically ill ventilated patients. However,

we only included patients who had a CT scan made and the resulting selection bias might limit the generalizability of our findings to the overall ICU population. The APACHE II score of the study population was higher than the overall ICU population, all patients were ventilated, and had an ICU length of stay of at least 4 days, indicating that the study patients were severely ill. Low muscle quality at admission likely has greater impact in the more severely ill patients, because the effect of additional critical illness-related muscle wasting is greater in this population.

Muscle quality is typically defined as muscle strength per unit of muscle mass or cross-sectional area. However, measuring muscle strength in ventilated critically ill patients is not feasible. Therefore, we used SMD and IMAT as proxy markers for muscle quality [40]. To date, SMD on CT scans has been related to myosteatosis [14, 23]. However, in recent studies in ICU patients using ultrasound, a relation between ultrasound echogenicity and myonecrosis in muscle biopsies has been found [2, 41]. Changes in SMD on CT scans might therefore not only reflect myosteatosis, but also myonecrosis. A prospective study using CT scans and muscle biopsies will have to further elucidate which changes in muscle are reflected by SMD in ICU patients.

A further limitation to our study is its observational design, precluding any deduction of causality. In addition, the complexity of critical illness may obscure residual confounding. Finally, the focus of our study was the prediction of long-term mortality at ICU admission, e.g. whether muscle quality at admission is a predictor of long-term mortality independent of muscle mass and of the best validated predictive score (APACHE). Further studies are needed to determine the risk factors for poor muscle quality and to determine the additional impact of ICU-acquired weakness on long-term mortality.

### Conclusions

Low skeletal muscle quality at ICU admission, as assessed by skeletal muscle density on CT scans, is associated with higher 6-month mortality in mechanically ventilated patients, independent of muscle quantity, APACHE II score, and BMI. Low muscle quality was also associated with longer hospital length of stay in survivors. Therefore, muscle quality appears to be as important for outcome as muscle quantity. Future intervention studies, including nutrition and early exercise, should not only focus on preventing further deterioration of muscle quantity, but also of muscle quality.

### Abbreviations

APACHE: Acute Physiological, Age, and Chronic Health Evaluation; BMI: Body mass index; CI: Confidence interval; CT: Computed tomography; Hospital-LOS: Hospital length of stay; HR: Hazard ratio; HU: Hounsfield Units; ICU: Intensive care unit; ICU-LOS: ICU length of stay; IMAT: Intermuscular adipose tissue; LOV: Length of ventilation; SMD: Skeletal muscle density

### Acknowledgements

We thank Ronald Driessen from the Department of Intensive Care Medicine for his contribution in the collection of data.

### Funding

A research grant provided by Baxter Healthcare was used for acquisition of CT scan analysis software and for a part of CT scan analysis. The funding source was not involved in any aspect of the design of the study, nor in collection, analysis, and interpretation of data, nor in manuscript preparation.

### Availability of data and materials

The datasets generated and analysed during the current study are available from the corresponding author on reasonable request.

### Authors' contributions

WGPML, HMO-vS and PJMW designed research; WGPML and IMD collected data; ARJG and SNS provided essential resources; WGPML, PJMW, HMO-vS, and JWRT analysed the data; WGPML, HMO-vS, and PJMW wrote the paper; WGPML had primary responsibility for final content. All authors read and approved the final manuscript.

### Competing interests

The authors declare that they have no competing interests.

### Consent for publication

All images in this manuscript are entirely unidentifiable and do not include any personal details, therefore no consent for publication was obtained.

### Ethics approval and consent to participate

The study was approved by the Institutional Review Board of the VU University Medical Center (identification number 2012/243). The need for informed consent was waived because of the retrospective nature of the study using only data obtained from standard care.

### Author details

<sup>1</sup>Department of Intensive Care Medicine, VU University Medical Center Amsterdam, De Boelelaan 1117, Amsterdam, The Netherlands. <sup>2</sup>Institute for Cardiovascular Research, VU University Medical Center Amsterdam, De Boelelaan 1117, Amsterdam, The Netherlands. <sup>3</sup>Department of Nutrition and Dietetics, Internal Medicine, VU University Medical Center Amsterdam, De Boelelaan 1117, 1081 HV Amsterdam, The Netherlands. <sup>4</sup>Department of Epidemiology and Biostatistics, VU University Medical Center Amsterdam, van der Boerhorststraat 7, Amsterdam, The Netherlands. <sup>5</sup>Department of Nutrition and Dietetics, Amsterdam University of Applied Sciences, Dr. Meurerlaan 8, Amsterdam, The Netherlands. <sup>6</sup>VU University Medical Center Amsterdam, Room ZH 7D174, P.O. Box 7057, 1007 MB Amsterdam, The Netherlands.

Received: 26 July 2016 Accepted: 8 November 2016

Published online: 01 December 2016

### References

- Hermans G, Van den Berghe G. Clinical review: intensive care unit acquired weakness. *Crit Care*. 2015;19:274.
- Puthuchery ZA, Rawal J, McPhail M, Connolly B, Ratnayake G, Chan P, et al. Acute skeletal muscle wasting in critical illness. *JAMA*. 2013;310:1591–600.
- Ali NA, O'Brien Jr JM, Hoffmann SP, Phillips G, Garland A, Finley JC, et al. Acquired weakness, handgrip strength, and mortality in critically ill patients. *Am J Respir Crit Care Med*. 2008;178:261–8.
- De Jonghe B, Sharshar T, Lefaucheur JP, Authier FJ, Durand-Zaleski I, Boussarsar M, et al. Paresis acquired in the intensive care unit: a prospective multicenter study. *JAMA*. 2002;288:2859–67.
- De Jonghe B, Bastuji-Garin S, Durand MC, Malissin I, Rodrigues P, Cerf C, et al. Respiratory weakness is associated with limb weakness and delayed weaning in critical illness. *Crit Care Med*. 2007;35:2007–15.
- Wieske L, Dettling-Ihnenfeldt DS, Verhamme C, Nollet F, van Schaik IN, Schultz MJ, et al. Impact of ICU-acquired weakness on post-ICU physical functioning: a follow-up study. *Crit Care*. 2015;19:196.
- Herridge MS, Tansey CM, Matte A, Tomlinson G, Diaz-Granados N, Cooper A, et al. Functional disability 5 years after acute respiratory distress syndrome. *N Engl J Med*. 2011;364:1293–304.
- Iwashyna TJ, Ely EW, Smith DM, Langa KM. Long-term cognitive impairment and functional disability among survivors of severe sepsis. *JAMA*. 2010;304:1787–94.
- Weijs PJ, Looijaard WG, Dekker IM, Stapel SN, Girbes AR, Oudemans-van Straaten HM, et al. Low skeletal muscle area is a risk factor for mortality in mechanically ventilated critically ill patients. *Crit Care*. 2014;18:R12.
- Moisey LL, Mourtzakis M, Cotton BA, Premji T, Heyland DK, Wade CE, et al. Skeletal muscle predicts ventilator-free days, ICU-free days, and mortality in elderly ICU patients. *Crit Care*. 2013;17:R206.
- Goodpaster BH, Carlson CL, Visser M, Kelley DE, Scherzinger A, Harris TB, et al. Attenuation of skeletal muscle and strength in the elderly: The Health ABC Study. *J Appl Physiol* (1985). 2001;90:2157–65.
- Martin L, Birdsell L, Macdonald N, Reiman T, Clandinin MT, McCargar LJ, et al. Cancer cachexia in the age of obesity: skeletal muscle depletion is a powerful prognostic factor, independent of body mass index. *J Clin Oncol*. 2013;31:1539–47.
- Antoun S, Lanoy E, Iacovelli R, Albiges-Sauvin L, Liorot Y, Merad-Taoufik M, et al. Skeletal muscle density predicts prognosis in patients with metastatic renal cell carcinoma treated with targeted therapies. *Cancer*. 2013;119:3377–84.
- Goodpaster BH, Kelley DE, Thaete FL, He J, Ross R. Skeletal muscle attenuation determined by computed tomography is associated with skeletal muscle lipid content. *J Appl Physiol* (1985). 2000;89:104–10.
- Parry SM, El-Ansary D, Cartwright MS, Sarwal A, Berney S, Koopman R, et al. Ultrasonography in the intensive care setting can be used to detect changes in the quality and quantity of muscle and is related to muscle strength and function. *J Crit Care*. 2015;30:1151. e9–14.
- Addison O, Marcus RL, Lastayo PC, Ryan AS. Intermuscular fat: a review of the consequences and causes. *Int J Endocrinol*. 2014;2014:309570.
- Coen PM, Goodpaster BH. Role of intramyocellular lipids in human health. *Trends Endocrinol Metab*. 2012;23:391–8.
- MacDonald AJ, Greig CA, Baracos V. The advantages and limitations of cross-sectional body composition analysis. *Curr Opin Support Palliat Care*. 2011;5:342–9.
- Shen W, Punyanitya M, Wang Z, Gallagher D, St-Onge MP, Albu J, et al. Total body skeletal muscle and adipose tissue volumes: estimation from a single abdominal cross-sectional image. *J Appl Physiol* (1985). 2004;97:2333–8.
- Schweitzer L, Geisler C, Pourhassan M, Braun W, Gluer CC, Bosy-Westphal A, et al. What is the best reference site for a single MRI slice to assess whole-body skeletal muscle and adipose tissue volumes in healthy adults? *Am J Clin Nutr*. 2015;102:58–65.
- Ruan XY, Gallagher D, Harris T, Albu J, Heymsfield S, Kuznia P, et al. Estimating whole body intermuscular adipose tissue from single cross-sectional magnetic resonance images. *J Appl Physiol* (1985). 2007;102:748–54.
- Mourtzakis M, Prado CM, Liefers JR, Reiman T, McCargar LJ, Baracos VE. A practical and precise approach to quantification of body composition in cancer patients using computed tomography images acquired during routine care. *Appl Physiol Nutr Metab*. 2008;33:997–1006.
- Aubrey J, Esfandiari N, Baracos VE, Buteau FA, Frenette J, Putman CT, et al. Measurement of skeletal muscle radiation attenuation and basis of its biological variation. *Acta Physiol (Oxf)*. 2014;210:489–97.
- Baracos V, Kazemi-Bajestani SM. Clinical outcomes related to muscle mass in humans with cancer and catabolic illnesses. *Int J Biochem Cell Biol*. 2013;45:2302–8.
- Delmonico MJ, Harris TB, Visser M, Park SW, Conroy MB, Velasquez-Mieyer P, et al. Longitudinal study of muscle strength, quality, and adipose tissue infiltration. *Am J Clin Nutr*. 2009;90:1579–85.
- Manini TM, Clark BC, Nalls MA, Goodpaster BH, Ploutz-Snyder LL, Harris TB. Reduced physical activity increases intermuscular adipose tissue in healthy young adults. *Am J Clin Nutr*. 2007;85:377–84.
- Zderic TW, Hamilton MT. Physical inactivity amplifies the sensitivity of skeletal muscle to the lipid-induced downregulation of lipoprotein lipase activity. *J Appl Physiol* (1985). 2006;100:249–57.
- Bey L, Hamilton MT. Suppression of skeletal muscle lipoprotein lipase activity during physical inactivity: a molecular reason to maintain daily low-intensity activity. *J Physiol*. 2003;551:673–82.
- Hikida RS, Gollnick PD, Dudley GA, Convertino VA, Buchanan P. Structural and metabolic characteristics of human skeletal muscle following 30 days of simulated microgravity. *Aviat Space Environ Med*. 1989;60:664–70.
- Ferretti G, Antonutto G, Denis C, Hoppeler H, Minetti AE, Narici MV, et al. The interplay of central and peripheral factors in limiting maximal O<sub>2</sub> consumption in man after prolonged bed rest. *J Physiol*. 1997;501(Pt 3):677–86.

31. Wagenmakers AJ. A malonyl-CoA fuel sensing mechanism in muscle: effects of insulin, glucose and denervation. *Clin Nutr.* 1996;15:144–5.
32. Stein TP, Wade CE. Metabolic consequences of muscle disuse atrophy. *J Nutr.* 2005;135:1824s–8s.
33. Lehr S, Hartwig S, Sell H. Adipokines: a treasure trove for the discovery of biomarkers for metabolic disorders. *Proteomics Clin Appl.* 2012;6:91–101.
34. Khan IM, Perrard XY, Brunner G, Lui H, Sparks LM, Smith SR, et al. Intermuscular and perimuscular fat expansion in obesity correlates with skeletal muscle T cell and macrophage infiltration and insulin resistance. *Int J Obes (Lond).* 2015;39:1607–18.
35. Lee DE, Kehlenbrink S, Lee H, Hawkins M, Yudkin JS. Getting the message across: mechanisms of physiological cross talk by adipose tissue. *Am J Physiol Endocrinol Metab.* 2009;296:E1210–29.
36. Reid MB, Lannergren J, Westerblad H. Respiratory and limb muscle weakness induced by tumor necrosis factor- $\alpha$ : involvement of muscle myofilaments. *Am J Respir Crit Care Med.* 2002;166:479–84.
37. Wilcox P, Osborne S, Bressler B. Monocyte inflammatory mediators impair in vitro hamster diaphragm contractility. *Am Rev Respir Dis.* 1992;146:462–6.
38. Sharshar T, Bastuji-Garin S, Stevens RD, Durand MC, Malissin I, Rodriguez P, et al. Presence and severity of intensive care unit-acquired paresis at time of awakening are associated with increased intensive care unit and hospital mortality. *Crit Care Med.* 2009;37:3047–53.
39. Casaer MP, Langouche L, Coudyzer W, Vanbeckevoort D, De Dobbelaer B, Guiza FG, et al. Impact of early parenteral nutrition on muscle and adipose tissue compartments during critical illness. *Crit Care Med.* 2013;41:2298–309.
40. McGregor RA, Cameron-Smith D, Poppitt SD. It is not just muscle mass: a review of muscle quality, composition and metabolism during ageing as determinants of muscle function and mobility in later life. *Longev Healthspan.* 2014;3:9.
41. Puthuchery ZA, Phadke R, Rawal J, McPhail MJ, Sidhu PS, Rowleron A, et al. Qualitative ultrasound in acute critical illness muscle wasting. *Crit Care Med.* 2015;43:1603–11.
42. World Health Organization. Obesity: preventing and managing the global epidemic: report of a WHO consultation. *World Health Organ Tech Rep Ser.* 2000;894:i-253.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

