

Lactate clearance as a target of therapy in sepsis: a flawed paradigm

PE Marik^{1*}, R Bellomo²

Abstract

Introduction

An increased blood lactate level is widely believed to be a marker of inadequate oxygen delivery and anaerobic metabolism. Furthermore, the rate of decline in lactate concentration (lactate clearance) has been recommended as an end-point of early goal directed therapy in critical ill patients with sepsis. We provide compelling data that an elevated lactate concentration is a consequence of increased aerobic glycolysis as part of the stress response and that titrating therapy to the rate of decline in lactate concentration is a potentially harmful endeavour. Furthermore, an increased lactate concentration may be an important adaptive survival response during critical illness.

Conclusion

An elevated lactate concentration in patients with sepsis is a marker of disease severity and not an indication of anaerobic metabolism. Increasing oxygen delivery to treat a non-existent oxygen debt may be a harmful undertaking. 'Lactate clearance' should not be used as the end-point of resuscitation in patients with sepsis.

Introduction

It is widely believed that in critically ill patients when oxygen delivery fails to meet oxygen demand an oxygen debt with global tissue hypoxia ensues¹⁻². This results in anaerobic metabolism and increased lactate production¹⁻².

An increased blood lactate concentration is therefore regarded as irrefutable evidence of anaerobic metabolism and tissue hypoxia¹. Nguyen reported that 'lactate clearance', defined as the percentage decrease in lactate from emergency department presentation to 6h, was an independent predictor of mortality¹. They concluded that 'lactate clearance in the early hospital course may indicate a resolution of global tissue hypoxia and that this is associated with decreased mortality rates.' This study popularized the concept of 'lactate clearance' and has led to a number of studies which have used 'lactate clearance' as the major end-point of haemodynamic resuscitation in critically ill patients with sepsis³⁻⁵. In this paper we demonstrate that this logic is scientifically flawed and that attempting to titrate therapy to a blood lactate level may be harmful. We suggest that the degree of elevation of the blood lactate is a reflection of the severity of illness and the degree of activation of the stress response

(and release of epinephrine) rather than being a marker of anaerobic metabolism. Furthermore, an increased lactate concentration may be an important adaptive survival response during critical illness.

Discussion

The authors have referenced some of their own studies in this review. These referenced studies have been conducted in accordance with the Declaration of Helsinki (1964) and the protocols of these studies have been approved by the relevant ethics committees related to the institution in which they were performed. All human subjects, in these referenced studies, gave informed consent to participate in these studies.

Lactate metabolism

Lactate is produced by glycolysis and metabolised by the liver and to a lesser degree by the kidney. Lactate is produced in the cytoplasm according to the following reaction (see Figure 1):

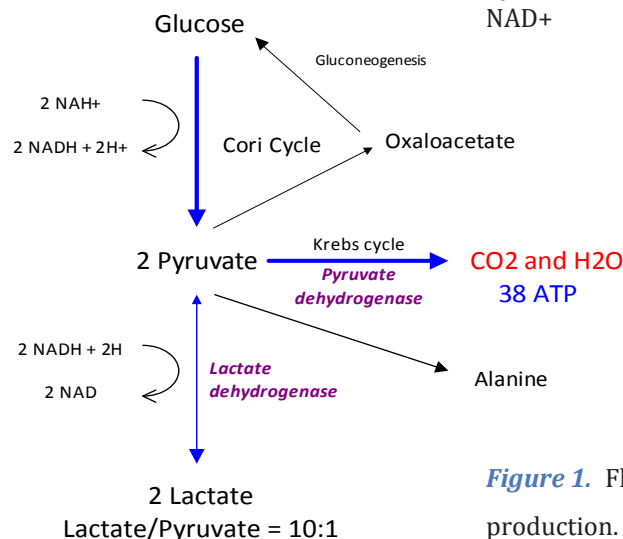
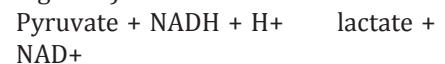


Figure 1. Flow chart of lactate production.

*Corresponding author
E-mail: marikpe@evms.edu

¹Division of Pulmonary and Critical Care Medicine, Eastern Virginia Medical School, Norfolk, VA 23507, USA.

²Australian and New Zealand Intensive Care Research Centre, Melbourne, Australia.

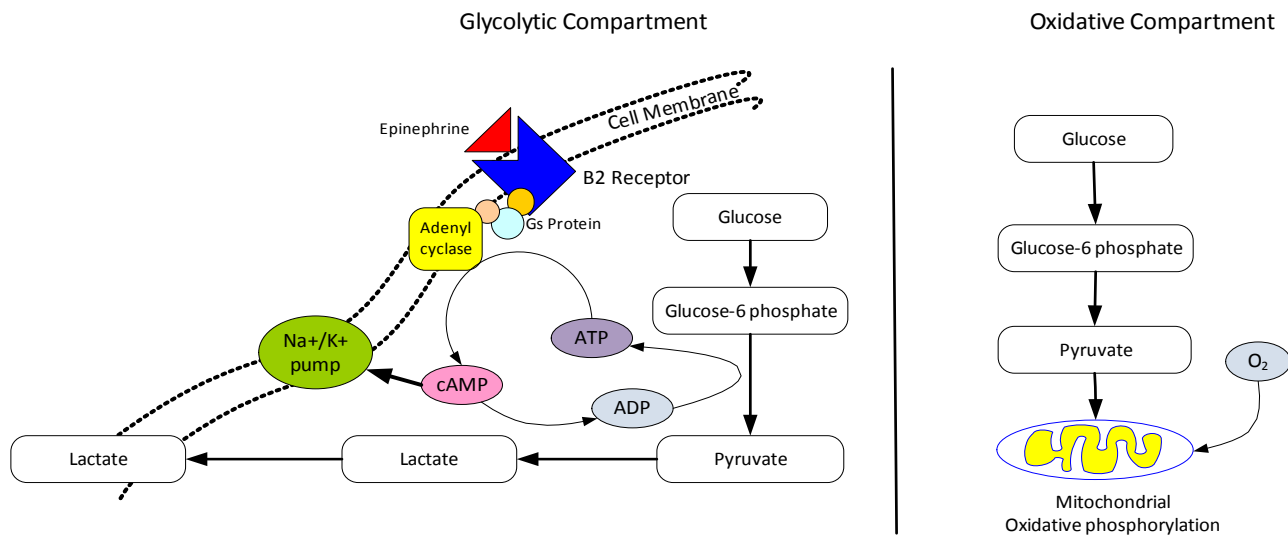


Figure 2. Glycolytic pathway. Epinephrine-increased glycolysis is coupled to Na^+/K^+ ATPase activity. From James *et al*³⁹.

This reaction favours lactate formation, yielding a ten-fold lactate/pyruvate ratio. In physiological conditions, lactate is produced by muscles (25%), skin (25%), brain (20%), intestine (10%) and red blood cells (20%)⁶. Increased glycolysis results in increased lactate formation. Arterial lactate concentration is dependent on the balance between its production and consumption. In general, this concentration is less than 2 mmol/l, although daily production of lactate is actually 1500 mmol/l⁶. Generated lactate can be transformed into oxaloacetate or alanine via the pyruvate pathway or can be utilized directly by periportal hepatocytes (60%) to produce glycogen and glucose (glycogenesis and glucogenesis; Cori cycle). The kidney also participates in the metabolism of lactate (30%), with the cortex classically acting as the metabolizer by glucogenesis and the medulla as a producer of lactate. Pyruvate is metabolized by the mitochondrial aerobic oxidation pathway via the Krebs cycle. This reaction leads to the production of large quantities of ATP (36 molecules of ATP for one molecule of pyruvate) (see Figure 2).

Hypoxia blocks mitochondrial oxidative phosphorylation, thereby

inhibiting ATP synthesis and reoxidation of NADH. This leads to a decrease in the ATP/ADP ratio and an increase in the NADH/NAD ratio. A decrease in the ATP/ADP ratio induces both an accumulation of pyruvate, which cannot be utilized by way of phosphofructokinase stimulation, and a decrease in pyruvate utilization by inhibiting pyruvate carboxylase, which converts pyruvate into oxaloacetate.⁶ Consequently, the increase in lactate production in an anaerobic setting is the result of an accumulation of pyruvate which is converted into lactate stemming from alterations in the redox potential; this results in an increase in the lactate/pyruvate ratio.

Classic teaching suggests that increased production of lactate results in acidosis, known widely as lactic acidosis.⁷ Close examination of glycolysis reveals that complete metabolism of glucose to lactate results in no net release of protons and, thus, does not contribute to acidosis. In fact, during the production of lactate from pyruvate, protons are consumed and acidosis is inhibited (Figure 1)⁸. Furthermore, lactate oxidation and lactate consumption via gluconeogenesis consume hydrogen ions and are alkalinizing processes.

This implies that 'lactic acidosis' is a condition that does not exist⁸.

Lactate as a marker of illness severity

It has been well established that an increased blood lactate concentration is a powerful predictor of mortality in critically ill patients. Over 50 years ago Weil and colleagues demonstrated an exponential increase in the mortality of critically ill patients with increasing blood lactate concentrations⁹⁻¹⁰. More recently, studies in both septic and trauma patients have demonstrated an independent association between increasing serum lactate concentration with organ failure and mortality¹¹⁻¹⁹. These studies suggest that the mortality increases linearly above a lactate concentration of approximately 1 mmol/l and that this association is independent of organ dysfunction or the presence of shock¹¹⁻²⁰. In patients with sepsis, a serum lactate concentration of more than 4 mmol/l is used as a marker of severe disease with an associated high risk of death^{2,14}.

Lactate clearance

A number of studies performed during the 1980s demonstrated that the ability to 'clear lactate' to normal in

patients suffering from both septic and cardiogenic shock was associated with an improved outcome. These authors coined the term 'lactate clearance'²¹⁻²³. In 1993 Abramson et al. reported that 'lactate clearance', defined as a decrease of lactate to less than 2mmol/l by 24h, was a predictor of survival following traumatic injury²⁴. These authors suggested increasing oxygen delivery in those patients in whom lactate fails to clear. The concept of lactate clearance was subsequently popularized by Nguyen and colleagues¹. While lactate clearance has been reported to be prognostic of outcome^{1,12,18,23-26}, not all studies have replicated this finding²⁷.

Lactate as a marker of metabolic stress

Cytosolic glycolytic flux is functionally divided into two distinct compartments. There are two distinctive glycolytic pathways utilizing separate glycolytic enzyme pools. The first pathway participates in oxidative metabolism via the Krebs cycle. The second pathway is linked to activity of the Na⁺/K⁺-ATPase pump (see Figure 2)⁶. ATP produced by this pathway is used to fuel this membrane pump. Numerous studies have demonstrated that epinephrine, via β 2-adrenoceptor stimulation, incAMP production, inducing the stimulation of glycogenolysis and glycolysis (ATP production) as well as activation of the Na⁺/K⁺-ATPase pump, which in turn will consume this ATP, thereby producing ADP²⁸⁻²⁹. This generated ADP via phosphofructokinase stimulation will reactivate glycolysis and hence generate more pyruvate and thereafter lactate.

Several studies performed over four decades ago provide strong evidence that hyperlactacidemia noted during shock states was unlikely to be caused by tissue hypoxia³⁰⁻³¹. These studies showed that hyperlactacidemia accompanying haemorrhage could be largely prevented by pretreatment with combined alpha and beta adrenergic-receptor blockade³².

Subsequent experimental studies confirmed that elevated arterial lactate in shock was not due to lack of oxygen but due to increased lactate production that could be mimicked by epinephrine infusion and blocked by adrenergic receptor blockade³³⁻³⁷. In these studies plasma lactate correlated well with plasma catecholamines concentrations. Furthermore, animal models of sepsis have demonstrated that despite shock and organ hypoperfusion tissue hypoxia is not a major pathophysiological finding³⁸. It has now been well established that epinephrine released as part of the stress response in patients with shock stimulates Na⁺/K⁺-ATPase activity. Increased activity of Na⁺/K⁺-ATPase leads to increased lactate production under well-oxygenated conditions in various cells, including erythrocytes, vascular smooth muscle, neurons, glia, and skeletal muscle^{29,39}. This concept was confirmed by Levy et al. who in patients with septic shock demonstrated that skeletal muscle was the leading source of lactate formation as a result of exaggerated *aerobic* glycolysis through Na⁺/K⁺-ATPase stimulation⁴⁰. Selective inhibition of Na⁺/K⁺-ATPase with ouabain infusion stopped overproduction of muscle lactate and pyruvate. This study demonstrated that increased aerobic glycolysis in skeletal muscle secondary to epinephrine-stimulated Na⁺/K⁺-ATPase activity and not anaerobic glycolysis (due to tissue hypoxia) is the major source of increased lactate in sepsis.

The hypermetabolic state with increased Na⁺/K⁺-ATPase activity results in accelerated glycolysis and generates pyruvate and lactate at an increased rate. If glycolysis occurs at a rate that exceeds that of oxidative metabolism, some pyruvate may not be oxidatively metabolized in the Krebs cycle and will be converted to lactate. The result will be a concomitant increase in both pyruvate and lactate with an unchanged lactate/pyruvate ratio (L/P). This observa-

tion has been demonstrated in patients with sepsis⁴¹. Gore measured lactate and pyruvate concentrations and the rates of pyruvate production and oxidation prior to and after dichloroacetate (DCA) administration in septic patients with severe lactic acidosis⁴². The patients in this report had significantly elevated levels of glucose, lactate and pyruvate (normal L/P ratio), with an increase in oxygen consumption and a significant decrease in glucose, lactate and pyruvate (unchanged L/P ratio) after the administration of DCA. This study confirmed the rate limiting effect of oxidative metabolism. Revelly et al. studied lactate kinetics in patients with severe sepsis. These authors demonstrated that hyperlactemia was related to increased production whereas lactate clearance was similar to that of healthy subjects⁴³.

These studies suggest that both increased lactate production and hyperglycemia are a consequence of activation of the stress response and are likely an essential evolutionary-preserved survival response⁴⁴. Under stable conditions the heart oxidizes free fatty acids for 70%-90% of its bioenergetic needs⁴⁵. However, the heart subjected to shock undergoes a shift in substrate utilisation such that it oxidizes lactate for the majority of its energy needs⁴⁶. Accelerated lactate clearance could therefore compromise cardiac performance during shock⁴⁷. In a rat endotoxin model, Levy et al. inhibited lactate production with a selective β 2-adrenergic blocker, enhanced its metabolism with dichloroacetate or studied a combination on both interventions³⁷. In this study lactate deprivation was associated with cardiovascular collapse and early death of the animals. Conversely, Revelly et al. demonstrated that an infusion of sodium lactate increased cardiac performance in patients with both cardiogenic and septic shock⁴³. These studies confirmed that lactate serves as an important energy source during acute haemo-

dynamic stress. Furthermore, while glucose serves as the major energy source for the brain increased brain lactate oxidation may occur with acute stress⁴⁸.

The concept that an epinephrine induced hypermetabolic state is responsible for increased lactate concentrations is supported by two randomized controlled trials, which investigated the haemodynamic effects and outcome of patients with septic shock treated with epinephrine or norepinephrine^{49–50}. In both these trials, epinephrine was associated with an initial increase in serum lactate concentration despite an increase in cardiac output and oxygen delivery. Furthermore, the magnitude of the increase in lactate and glucose following an infusion of epinephrine appears to be of prognostic importance with those patients with a blunted response having a significantly higher mortality³⁶. In addition to epinephrine induction increased lactate production may impair the activity of the pyruvate dehydrogenase enzyme complex, which in the setting of accelerated aerobic glycolysis, further increases lactate levels^{42,51}.

Increasing oxygen delivery may be harmful

Current evidence suggests that most of the increase in blood lactate levels in patients with severe sepsis is unrelated to poor tissue perfusion and is therefore unlikely to respond to iatrogenic attempts to increase oxygen delivery. Driving up oxygen delivery in patients without an oxygen debt will not increase oxygen consumption, and it is likely to be harmful. Hayes et al. performed a randomized controlled trial in which patients were randomized to 'supranormal oxygen delivery' or standard therapy⁵². Despite a significant increase in oxygen delivery in the supranormal group, oxygen consumption remained unchanged, while the mortality was significantly higher

than in the control group. Similarly, Marik and Sibbald demonstrated that blood transfusion in septic patients with an increased lactate concentration did not result in an increase in oxygen consumption⁵³. These data demonstrate that patients with sepsis and an increased lactate do not have an oxygen debt and that increasing oxygen delivery will not increase oxygen consumption and that such an approach is unlikely to be beneficial⁵⁴ and may be harmful⁵². Furthermore, we believe that the term 'lactate clearance' is scientifically incorrect. Clearance in medicine is expressed as milliliter per minute. What the authors who have popularized this term presumably mean is the rate of decline in the serum lactate concentration¹. Furthermore, it is impossible to know if the rate of decline is due to (i) increased removal (metabolism), (ii) decreased production, (iii) dilution due to fluid resuscitation or (iv) all of the above in variable combinations.

We believe that a fall in lactate concentration following the initiation of treatment for sepsis is due to an attenuation of the stress response and not due to correction of an oxygen debt. Furthermore, while a failure of blood lactate levels to decline after the initiation of treatment is an ominous sign, adequate lactate clearance does not guarantee survival. Our review demonstrates that the concept of 'lactate clearance' is fundamentally flawed, and as such, 'lactate clearance' should not be used as the end-point of resuscitation in patients with sepsis.

Conclusion

We believe that a fall in lactate concentration following the initiation of treatment for sepsis is due to an attenuation of the stress response and not due to correction of an oxygen debt. Furthermore, while a failure of blood lactate levels to decline after the initiation of treatment is an ominous sign, adequate lactate clearance does not guarantee survival. Our re-

view demonstrates that the concept of 'lactate clearance' is fundamentally flawed, and as such, 'lactate clearance' should not be used as the end-point of resuscitation in patients with sepsis

References

1. Nguyen HB, Rivers EP, Knoblich BP, Jacobsen G, Muzzin A, Ressler JA, et al. Early lactate clearance is associated with improved outcome in severe sepsis and septic shock. *Crit Care Med.* 2004 Aug;32(8):1637–42.
2. Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, et al. Surviving Sepsis Campaign: International Guidelines for Management of Severe Sepsis and Septic Shock: 2012. *Crit Care Med.* 2013 Feb;41(2):580–637.
3. Jones AE, Shapiro NI, Trzeciak S, Arnold RC, Claremont HA, Kline JA. Lactate clearance vs central venous oxygen saturation as goals of early sepsis therapy: a randomized clinical trial. *JAMA* 2010 Feb;303(8):739–46.
4. Jansen TC, van BJ, Schoonderbeek FJ, Smeetswijk Visser SJ, van der Klooster JM, Lima AP, et al. Early lactate-guided therapy in intensive care unit patients: a multicenter, open-label, randomized controlled trial. *Am J Respir Crit Care Med.* 2010 Sep;182(6):752–61.
5. Nguyen HB, Kuan WS, Batech M, Shrikhande P, Mahadevan M, Li CH, et al. Outcome effectiveness of the severe sepsis resuscitation bundle with addition of lactate clearance as a bundle item: a multi-national evaluation. *Crit Care* 2011 15(5):R229.
6. Levy B. Lactate and shock state: the metabolic view. *Curr Opin Crit Care* 2006 Aug;12(4):315–21.
7. Vernon C, LeTourneau JL. Lactic acidosis: Recognition, kinetics and associated prognosis. *Crit Care Clin.* 2010 Apr;26(2):255–83.
8. Robergs RA, Ghiasvand F, Parker D. Biochemistry of exercise-induced metabolic acidosis. *Am J Physiol Regul Integr Comp Physiol.* 2004 Sep;287(3):R502–16.
9. Broder G, Weil MH. Excess lactate: An index of reversibility of shock in human patients. *Science.* 1964 Mar;143(3613):1457–9.
10. Cady LD, Jr., Weil MH, Afifi AA, Michaels SF, Liu VY, Shubin H. et al. Quantitation of severity of critical illness with special reference to blood lactate.

Crit Care Med. 1973 Mar-Apr;1(2):75–80.

11. Regnier MA, Raux M, Le MY, Asencio Y, Gaillard J, Devilliers C, et al. Prognostic significance of blood lactate and lactate clearance in trauma patients. *Anesthesiol.* 2012 Dec;117(6):1276–88.

12. Manikis P, Jankowski S, Zhang H, Kahn RJ, Vincent JL. Correlation of serial blood lactate levels to organ failure and mortality after trauma. *Am J Emerg Med.* 1995 Nov;13(6):619–22.

13. Shapiro NI, Howell MD, Talmor D, Nathanson LA, Lisbon A, Wolfe RE, et al. Serum lactate as a predictor of mortality in emergency department patients with infection. *Ann Emerg Med.* 2005 May;45(5):524–8.

14. Trzeciak S, Dellinger RP, Chansky ME, Arnold RC, Schorr C, Milcarek B, et al. Serum lactate as a predictor of mortality in patients with infection. *Intensive Care Med.* 2007 Jun;33(6):970–7.

15. Bakker J, Coffernils M, Leon M, Gris P, Vincent JL. Blood lactate levels are superior to oxygen-derived variables in predicting outcome in human septic shock. *Chest* 1991 Apr;99(4):956–62.

16. Aduen J, Bernstein WK, Khastgir T, Miller J, Kerzner R, Bhatiani A, et al. The use and clinical importance of a substrate-specific electrode for rapid determination of blood lactate concentrations. *JAMA* 1994 Dec; 272(21):1678–85.

17. Varpula M, Tallgren M, Saukkonen K, Voipio-Pulkki LM, Pettila V. Hemodynamic variables related to outcome in septic shock. *Intensive Care Med.* 2005 Aug; 31(8):1066–71.

18. Bakker J, Gris P, Coffernils M, Kahn RJ, Vincent JL. Serial blood lactate levels can predict the development of multiple organ failure following septic shock. *Am J Surg* 1996 Feb;171(2):221–6.

19. Parker MM, Shelhamer JH, Natanson C, Alling DW, Parrillo JE. Serial cardiovascular variables in survivors and nonsurvivors of septic shock: Heart rate as an early predictor of prognosis. *Crit Care Med.* 1987 Oct;15(10):923–9.

20. Mikkelsen ME, Miltiades AN, Gaieski DF, Goyal M, Fuchs BD, Shah CV, et al. Serum lactate is associated with mortality in severe sepsis independent of organ failure and shock. *Crit Care Med.* 2009 May;37(5):1670–7.

21. Bock JS, Gottlieb SS. Cardiorenal syndrome: New perspectives. *Circulation.* 2010 Jun;121(23):2592–600.

22. Falk JL, Rackow EC, Leavy J, Astiz ME, Weil MH. Delayed lactate clearance in patients surviving circulatory shock. *Acute Care.* 1985 11(3–4):212–5.

23. Vincent JL, Dufaye P, Berre J, Leeman M, Degaute JP, Kahn RJ. Serial lactate determinations during circulatory shock. *Crit Care Med.* 1983; Jun;11(6):449–51.

24. Abramson D, Scalea TM, Hitchcock R, Trooskin SZ, Henry SM, Greenspan J. Lactate clearance and survival following injury. *J Trauma.* 1993 Oct;35(4):584–8.

25. Arnold RC, Shapiro NI, Jones AE, Schorr C, Pope J, Casner E, et al. Multicenter study of early lactate clearance as a determinant of survival in patients with presumed sepsis. *Shock* 2009 Jul;32(1):35–9.

26. McNelis J, Marini CP, Jurkiewicz A, Szomstein S, Simms HH, Ritter G, Lima AP, van der Hoven B, Rommes JH, et al. Prolonged lactate clearance is associated with increased mortality in the surgical intensive care unit. *Am J Surg.* 2001 Nov;182(5):481–5.

27. Jansen TC, van BJ, Mulder PG, et al. Prognostic value of blood lactate levels: does the clinical diagnosis at admission matter? *J Trauma* 2009 Feb; 66(2):377–85.

28. James JH, Wagner KR, King JK, Leffler RE, Upputuri RK, Balasubramaniam A, et al. Stimulation of both aerobic glycolysis and Na(+)-K(+)-ATPase activity in skeletal muscle by epinephrine or amylin. *Am J Physiol.* 1999 277(1pt1):E176–86.

29. James JH, Fang CH, Schrantz SJ, Hasselgren PO, Paul RJ, Fischer JE, et al. Linkage of aerobic glycolysis to sodium-potassium transport in rat skeletal muscle. Implications for increased muscle lactate production in sepsis. *J Clin Invest.* 1996 Nov;98(10):2388–97.

30. Irving MH. The sympatho-adrenal factor in haemorrhagic shock. *Ann R Coll Surg Engl.* 1968; Jun;42(6):367–86.

31. Daniel AM, Shizgal HM, MacLean LD. The anatomic and metabolic source of lactate in shock. *Surg Gynecol Obstet.* 1978 Nov;147(5):697–700.

32. Halmagyi DF, Kennedy M, Varga D. Combined adrenergic receptor blockade and circulating catecholamines in hemorrhagic shock. *Can J Surg.* 1971;3(6):378–88.

33. Liddell MJ, Daniel AM, MacLean LD, Shizgal HM. The role of stress hormones in the catabolic metabolism of shock. *Surg Gynecol Obstet.* 1979 Dec;149(6):822–30.

34. McCarter FD, James JH, Luchette FA, Wang L, Friend LA, King JK, et al. Adrenergic blockade reduces skeletal muscle

glycolysis and Na(+), K(+)-ATPase activity during hemorrhage. *J Surg Res.* 2001; Aug;99(2):235–44.

35. Halmagyi DF, Irving MH, Gillett DJ, Varga D. Effect of adrenergic blockade on consequences of sustained epinephrine infusion. *J Appl Physiol.* 1967 Aug;23(2):171–7.

36. Wutrich Y, Barraud D, Conrad M, Cravoisy-Popovic A, Nace L, Bollaert PE, et al. Early increase in arterial lactate concentration under epinephrine infusion is associated with a better prognosis during shock. *Shock.* 2010 Jul;3(1):4–9.

37. Levy B, Mansart A, Montemont C, Gibot S, Mallie JP, Regnault V, et al. Myocardial lactate deprivation is associated with decreased cardiovascular performance, decreased myocardial energetics, and early death in endotoxic shock. *Intensive Care Med.* 2007 Mar;33(3):495–502.

38. Regueira T, Djafarzadeh S, Brandt S, Gorrasi J, Borotto E, Porta F, et al. Oxygen transport and mitochondrial function in porcine septic shock, cardiogenic shock, and hypoxaemia. *Acta Anaesthesiol Scand.* 2012 Aug;56(7):846–59.

39. James JH, Luchette FA, McCarter FD, Fischer JE. Lactate is an unreliable indicator of tissue hypoxia in injury or sepsis. *Lancet.* 1999 354(9177):505–8.

40. Levy B, Gibot S, Franck P, Cravoisy A, Bollaert PE. Relation between muscle Na⁺K⁺ ATPase activity and raised lactate concentrations in septic shock: a prospective study. *Lancet.* 2005 Mar;365(9462):871–5.

41. Gore DC, Jahoor F, Hibbert JM, DeMaria EJ. Lactic acidosis during sepsis is related to increased pyruvate production, not deficits in tissue oxygen availability. *Ann Surg.* 1996 Jul;224(1):97–102.

42. Stacpoole PW, Wright EC, Baumgartner TG, Bersin RM, Buchalter S, Curry SH, et al. A controlled clinical trial of dichloroacetate for treatment of lactic acidosis in adults. The Dichloroacetate-Lactic Acidosis Study Group. *N Engl J Med.* 1992 Nov;327(22):1564–69.

43. Revelly JP, Tappy L, Martinez A, Bollmann M, Cayeux MC, Berger MM, et al. Lactate and glucose metabolism in severe sepsis and cardiogenic shock. *Crit Care Med.* 2005 Oct;33(10):2235–40.

44. Marik PE, Bellomo R. Stress hyperglycemia: an essential survival response? *Crit Care.* 2013 Mar;17(2):305.

45. Beadle RM, Frenneaux M. Modification of myocardial substrate utilisation: a new therapeutic paradigm in

cardiovascular disease. *Heart*. 2010 Jun;96(11):824–30.

46. Spitzer JJ, Spitzer JA. Myocardial metabolism in dogs during hemorrhagic shock. *Am J Physiol*. 1972 222(1):101–5.

47. Barbee RW, Kline JA, Watts JA. Depletion of lactate by dichloroacetate reduces cardiac efficiency after hemorrhagic shock. *Shock*. 2000 Aug;14(2):208–14.

48. Wyss MT, Jolivet R, Buck A, Magistretti PJ, Weber B. *In vivo* evidence for lactate as a neuronal energy source of Neuron. 2011 May;31(20):7477–85.

49. Myburgh JA, Higgins A, Jovanovska A, Lipman J, Ramakrishnan N, Santamaria J.

A comparison of epinephrine and norepinephrine in critically ill patients. *Intensive Care Med*. 2008 Dec;34(12):2226–34.

50. Annane D, Vignon P, Renault A, Bollaert PE, Charpentier C, Martin C, et al. Norepinephrine plus dobutamine versus epinephrine alone for management of septic shock: a randomised trial. *Lancet*. 2007; Aug;370(9588):676–84.

51. Alamdari N, Constantin-Teodosiu D, Murton AJ, Gardiner SM, Bennett T, Layfield R, et al. Temporal changes in the involvement of pyruvate dehydrogenase complex in muscle lactate accumulation during lipopolysaccha-

ride infusion in rats of *Physiology* 2008 Mar;586(6):1767–75.

52. Hayes MA, Timmins AC, Yau E, Palazzo M, Hinds CJ, Watson D. Elevation of systemic oxygen delivery in the treatment of critically ill patients. *N Engl J Med*. 1994 Jun;330(24):1717–22.

53. Marik PE, Sibbald WJ. Effect of stored-blood transfusion on oxygen delivery in patients with sepsis. *JAMA*. 1993 Jun;269(23):3024–9.

54. Gattinoni L, Brazzi L, Pelosi P, Latini R, Tognoni G, Pesenti A. A trial of goal-oriented hemodynamic therapy in critically ill patients. *N Engl J Med*. 1995 Oct;333(16):1025–32.