

Pharmacokinetic issues for antibiotics in the critically ill patient

Jason A. Roberts, B Pharm (Hons); Jeffrey Lipman, FJFICM, MD

LEARNING OBJECTIVES

On completion of this article, the reader should be able to:

1. Explain principles which influence pharmacokinetics in critically ill patients.
2. Describe rational dosing regimens for antibiotics in critically ill patients.
3. Use this information in a clinical setting.

Dr. Roberts has disclosed that he is the recipient of an education grant to the research center from Astra-Zeneca. Dr. Lipman has disclosed that he is the recipient of an education grant to the research center from Astra-Zeneca; is a consultant/advisor for Astra-Zeneca and Janssen-Cilag; and is on the speaker's bureau for Wyeth Australia.

All faculty and staff in a position to control the content of this CME activity have disclosed that they have no financial relationship with, or financial interests in, any commercial companies pertaining to this educational activity.

Lippincott CME Institute, Inc., has identified and resolved all faculty conflicts of interest regarding this educational activity.

Visit the *Critical Care Medicine* Web site (www.ccmjournal.org) for information on obtaining continuing medical education credit.

Objective: To discuss the altered pharmacokinetic properties of selected antibiotics in critically ill patients and to develop basic dose adjustment principles for this patient population.

Data Sources: PubMed, EMBASE, and the Cochrane-Controlled Trial Register.

Study Selection: Relevant papers that reported pharmacokinetics of selected antibiotic classes in critically ill patients and antibiotic pharmacodynamic properties were reviewed. Antibiotics and/or antibiotic classes reviewed included aminoglycosides, β -lactams (including carbapenems), glycopeptides, fluoroquinolones, tigecycline, linezolid, lincosamides, and colistin.

Data Synthesis: Antibiotics can be broadly categorized according to their solubility characteristics which can, in turn, help describe possible altered pharmacokinetics that can be caused by the pathophysiological changes common to critical illness. Hydrophilic antibiotics (e.g., aminoglycosides, β -lactams, glycopeptides, and colistin) are mostly affected with the pathophysiological changes observed in critically ill patients with increased volumes

of distribution and altered drug clearance (related to changes in creatinine clearance). Lipophilic antibiotics (e.g., fluoroquinolones, macrolides, tigecycline, and lincosamides) have lesser volume of distribution alterations, but may develop altered drug clearances. Using antibiotic pharmacodynamic bacterial kill characteristics, altered dosing regimens can be devised that also account for such pharmacokinetic changes.

Conclusions: Knowledge of antibiotic pharmacodynamic properties and the potential altered antibiotic pharmacokinetics in critically ill patients can allow the intensivist to develop individualized dosing regimens. Specifically, for renally cleared drugs, measured creatinine clearance can be used to drive many dose adjustments. Maximizing clinical outcomes and minimizing antibiotic resistance using individualized doses may be best achieved with therapeutic drug monitoring. (*Crit Care Med* 2009; 37: 840–851)

KEY WORDS: pharmacokinetics; critically ill; pharmacodynamics; antibiotic; dosing

Research Fellow (JR), the University of Queensland, Pharmacy Department, Royal Brisbane and Women's Hospital, Herston, Australia; and Professor (JL), University of Queensland and Royal Brisbane and Women's Hospital, Herston, Australia.

Supported, in part, by the Australian and New Zealand College of Anesthetists, Society of Hospital Pharmacists of Australia, the Queensland State Government—

Smart State Initiative, Royal Brisbane and Women's Hospital Research Foundation and the Australian National Health and Medical Research Council (519702). Supported by the Australian National Health and Medical Research Council (JAR) (409931).

Dr. Lipman holds consultancies with AstraZeneca and Wyeth. Mr. Roberts has not disclosed any potential conflicts of interest.

For information regarding this article, E-mail: j.lipman@uq.edu.au

Copyright © 2009 by the Society of Critical Care Medicine and Lippincott Williams & Wilkins

DOI: 10.1097/CCM.0b013e3181961bff

Antibiotic treatment of critically ill patients remains a significant challenge to intensivists world-wide with persisting high mortality and morbidity rates. The importance of effective therapy continues to grow with increasing numbers of patients with increasingly levels of sickness severity being admitted to intensive care units (ICUs) (1). Compelling evidence suggests that in infected critically ill patients, source control of the pathogen and early and appropriate antibiotic therapy remains the most important intervention that the clinician can implement for such patients (1–6). Therefore, optimizing antibiotic therapy should be a priority in the management of critically ill patients.

A vast array of pathophysiological changes can occur in critically ill patients that can complicate antibiotic dosing. Knowledge of the pharmacokinetic and pharmacodynamic properties of the antibiotics used for the management of critically ill patients is essential for selecting the antibiotic dosing regimens, which will optimize patient outcomes (7). Changes in volume of distribution (Vd) and clearance (CL) of antibiotics have been noted in these patients, which may affect the antibiotic concentration at the target site. It follows that the pharmacodynamic parameters that determine antibiotic efficacy, which can vary between antibiotic classes, may also be affected. Optimization of these parameters is necessary to maximize the rate of response through patient recovery and minimized antibiotic resistance (7–9).

The aim of this review is to identify the pathophysiological changes that occur in critically ill patients and the effect that they have on the pharmacokinetic behavior, and furthermore, the pharmacodynamic effect of commonly used antibiotics. Furthermore, we seek to develop general principles of dosage adjustment of these antibiotics in critically ill patients. Because of the spectrum of different patient presentations and different levels of organ function that critically ill patients may present with, it is not the intention of this article to provide definitive dose adjustment recommendations for each of the cited antibiotic classes. However, we aim to provide information that empowers the clinician to individualize antibiotic dosing by considering the factors that are most likely to affect antibiotic pharmacokinetics.

Search Strategy and Selection Criteria

Data for this review were identified by searches of PubMed (1966 to February 2008), EMBASE (1966 to February 2008) and the Cochrane Controlled Trial Registry as well as references from relevant articles. Search terms were “antibiotic” or “antibacterial,” “intensive care unit,” or “critically ill” or “critical illness,” and “pharmacokinetics” or “pharmacodynamics.” English language papers were reviewed. Numerous articles were identified through searches of the extensive files of the authors. All relevant papers that described antibiotic pharmacodynamics and/or antibiotic pharmacokinetics in critically ill patients were reviewed.

General Concepts

Kill Characteristics of Antibiotics. For antibiotics, pharmacodynamic parameters relate pharmacokinetic parameters

to the ability of the antibiotic to kill or inhibit the growth of the infective organism (10). Different antibiotic classes have been shown to have different kill characteristics on bacteria (Fig. 1 and Table 1).

Developing dosing regimens that maximize the rate of response in ICU patients is important for optimizing patient outcomes and minimizing the development of antibiotic resistance (2, 3, 5, 9).

Pharmacokinetic Changes Observed in Critically Ill Patients. The changes to the pharmacokinetic parameters of antibiotics in critically ill patients are driven by both drug and disease factors. From a drug perspective, the hydrophilicity and lipophilicity of the molecule will influence Vd and CL of a drug. Figure 2 summarizes these effects diagrammatically.

Changes in Vd. The pathogenesis of infections in critically ill patients appears highly complex (1, 11–13). Endotoxins from bacteria or fungi may stimulate the production of various endogenous mediators that may affect the vascular endothelium.

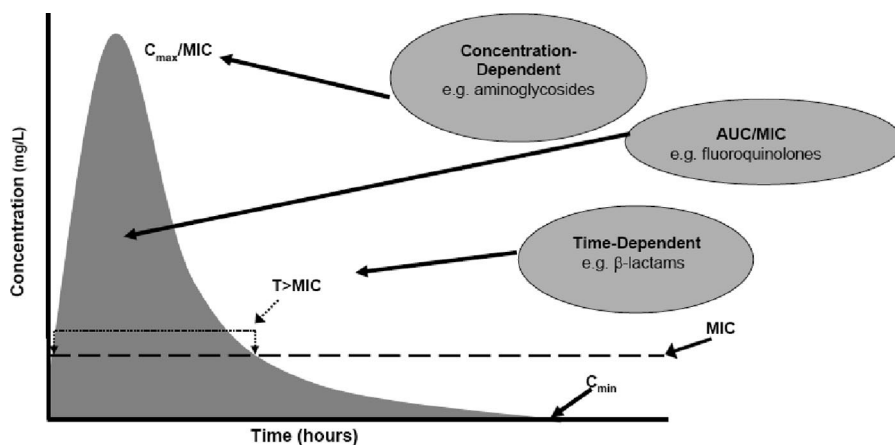


Figure 1. Pharmacokinetic and pharmacodynamic parameters of antibiotics on a concentration vs. time curve. Key: $T > MIC$ —The time for which a drug’s plasma concentration remains above the minimum inhibitory concentration (MIC) for a dosing period; C_{max}/MIC , the ratio of the maximum plasma antibiotic concentration (C_{max}) to MIC; AUC/MIC , the ratio of the area under the concentration time curve during a 24-hour time period (AUC_{0-24}) to MIC.

Table 1. Pharmacodynamic properties that correlate with efficacy of selected antibiotics

Antibiotics	β -lactams	Aminoglycosides	Fluoroquinolones
	Carbapenems	Metronidazole	Aminoglycosides
	Linezolid	Fluoroquinolones	Azithromycin
	Erythromycin	Telithromycin	Tetracyclines
	Clarithromycin	Daptomycin	Glycopeptides
	Lincosamides	Quinupristin/dalfopristin	Tigecycline
			Quinupristin/dalfopristin
			Linezolid
PD kill characteristics	Time-dependent	Concentration-dependent	Concentration-dependent with time-dependence
Optimal PD parameter	$T > MIC$	$C_{max}:MIC$	$AUC_{0-24}:MIC$

MIC, minimum inhibitory concentration; AUC, area under curve; PD, pharmacodynamics; C_{max} , maximum concentration.

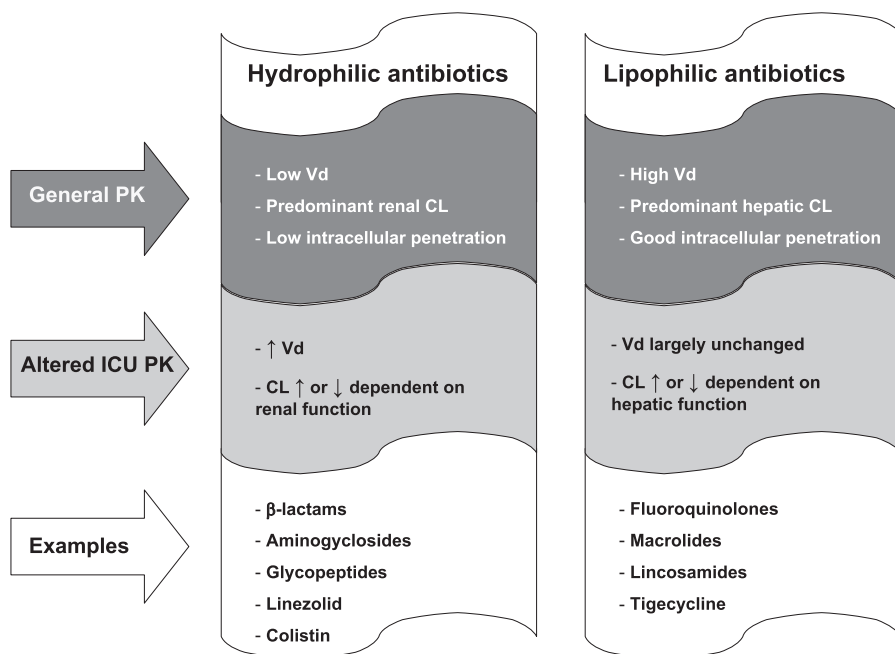


Figure 2. The interrelationship of hydrophilicity and lipophilicity of antibiotic molecules on the pharmacokinetic characteristics in general ward patients (General pharmacokinetics [PK]) and the altered PK observed in critically ill patients in intensive care unit (ICU). CL, clearance; Vd, volume of distribution.

lium resulting in either vasoconstriction or vasodilatation with maldistribution of blood flow, endothelial damage, and increased capillary permeability (14). This capillary leak syndrome results in fluid shifts from the intravascular compartment to the interstitial space (15, 16). This would increase the Vd of hydrophilic drugs which decreases their plasma drug concentration. Vd of hydrophilic drugs may also be increased by the presence of mechanical ventilation, hypoalbuminaemia (increased capillary leakage) extracorporeal circuits (e.g., plasma exchange, cardiopulmonary bypass), postsurgical drains, or in patients with significant burn injuries (17–20). Lipophilic drugs typically have a large Vd because of their partitioning into adipose tissue, and as such the increased Vd that results from third-spacing is likely to cause insignificant increases in drug Vd.

Changes in Antibiotic Half-Life. Drug elimination half-life ($T_{1/2}$) is directly related to antibiotic CL and Vd. $T_{1/2}$ is represented by the equation (21):

$$T_{1/2} = \frac{0.693 \times Vd}{CL}$$

It follows that an increased drug CL is likely to reduce $T_{1/2}$, whereas an increased Vd is likely to increase $T_{1/2}$.

CL, and therefore $T_{1/2}$, can be affected by the disease process that occurs in crit-

ically ill patients and from interventions of the intensivist. Standard initial management of hypotension that critically ill patients may develop is administration of intravenous fluids. When hypotension persists, vasopressor agents are prescribed. It is, therefore, not surprising that critically ill patients often have higher than normal cardiac indices (13, 22). Some information suggests that mechanical ventilation may cause decreased antibiotic CL (19). In the absence of significant organ dysfunction, there is often an increased renal perfusion and consequently increased creatinine clearance and elimination of hydrophilic antibiotics (23–25). It follows that dose adjustment for hydrophilic antibiotics can be guided by measures of creatinine clearance even in patients with significant burn injuries (19). Strong evidence suggests that the most effective way to calculate renal function remains using an 8, 12, or 24-hour creatinine clearance collection (26, 27), although recent work has suggested that a 2-hour creatinine clearance may be an adequate substitute (28). It must be emphasized that equations such as the Cockcroft-Gault (29) and Modified Diet in Renal Disease (30) equations are likely to be unreliable and, if possible, should not be substituted for urinary creatinine clearance data (31).

Further evidence suggests that critically ill patients may have higher creatinine clearances even in the presence of normal plasma creatinine concentrations (32, 33). A subsequent higher CL of renally eliminated drugs may result in a decreased $T_{1/2}$.

Hypoalbuminemia. Protein binding is a factor that may influence the Vd and CL of many antibiotics. A notable example of this pharmacokinetic alteration exists for ceftriaxone, which is 95% bound to albumin in normal ward patients (34, 35). In hypoalbuminemic states, as common in critically ill patients, this can result in a higher unbound concentration that has a 100% increased CL and 90% greater Vd (36). Other highly protein-bound antibiotics that probably develop altered pharmacokinetics from hypoalbuminaemia include oxacillin and teicoplanin.

Development of End-Organ Dysfunction. With further deterioration in the health status of the patient, significant myocardial depression can occur, which leads to a decrease in organ perfusion and failure of the microvascular circulation (37). This may then progress to multiple organ dysfunction syndrome, which may include renal and/or hepatic dysfunction (38). This will result in decreased antibiotic CL, prolonged $T_{1/2}$, and potential toxicity from elevated antibiotic concentrations and/or accumulation of metabolites. For some drugs, if dysfunction of the primary eliminating organ occurs, other organs may increase their intrinsic CL causing little change in expected plasma concentration (e.g., in renal dysfunction, ciprofloxacin transintestinal CL can increase, resulting in only a small decrease in total body CL) (39). Preliminary data also support increased biliary CL of ticarcillin and piperacillin in renal dysfunction (40, 41).

Figure 3 schematically identifies the pharmacokinetic changes that can occur because of the altered physiology in critical illness.

When renal dysfunction is present or if the patient needs renal replacement therapy, standard texts or review articles should be used as a guide for altered dosing (42–44).

Tissue Penetration. Antibiotic pharmacokinetics at the target site, which is usually tissue (45), are important to predict antibiotic-bacteria interactions. Microdialysis is an *in vivo* sampling technique that is the subject of an increasing number of research publications, particularly in critically ill patients (46–49).

The current data suggest that antibiotic penetration into tissues of patients with septic shock is impaired, possibly up to five to ten times lower than in healthy

volunteers, although in other patients with sepsis but without shock there seems to be a less significant effect on tissue concentrations (47–49). Therefore,

dosing of antibiotics at high doses is probably required to maximize antibiotic penetration, particularly in patients with shock, although data to support this is currently lacking.

The potential pharmacokinetic variability for many antibiotics requires the clinician to develop dosing strategies that account for altered pharmacokinetics and pathogen susceptibility studies in each patient. Such individualized dosing may facilitate optimized patient outcomes. Ongoing evaluations of sickness severity can facilitate timely adjustment of antibiotic dosing.

Specific Antibiotic Classes

General pharmacokinetic and pharmacodynamic characteristics will be considered for aminoglycosides, β -lactams, glycopeptides, fluoroquinolones, lincosamides as well as tigecycline, linezolid, and colistin. The clinical application and dosing implications of these properties for critically ill patients will also be addressed. Table 2 describes the potential altered pharmacokinetics of these antibiotics in critically ill patients.

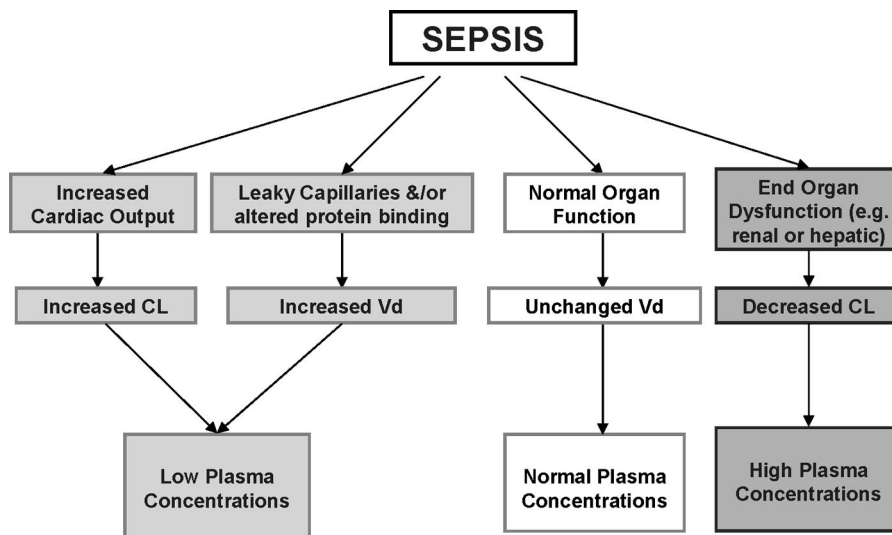


Figure 3. Schematic representation of the basic pathophysiological changes that can occur during sepsis and their subsequent pharmacokinetic effects. Note that there can be significant overlap between the groups above enabling multiple permutations for altered drug pharmacokinetics, e.g. patients with mild-to-moderate renal failure may develop increased transintestinal clearance of ciprofloxacin resulting in relatively normal plasma concentrations (39). *CL*, clearance; *Vd*, volume of distribution.

Table 2. General PK characteristics of various antibiotics and possible changes that can occur during fluid shifts in critically ill patients

Antibiotic Class	Vd (L/kg)	Increased Vd with Fluid Shifts?	Decreased C_{max} with Fluid Shifts?	Plasma $T_{1/2}$ (hrs)	Protein Binding	Altered CL in Critically Ill?	TDM Required?
Aminoglycosides (61, 62, 67)	0.2–0.3 (consistent with extracellular water)	Yes	Yes	2–3	Low	Varies proportionately with renal function	Yes, to ensure high C_{max} and adequate CL
β -lactams (33, 70, 155, 156)	Variable but consistent with extracellular water	Yes	Yes	0.5–2 (except ceftriaxone 6–9 hrs)	Low (except ceftriaxone and oxacillin)	Varies proportionately with renal function (some exceptions)	No
Carbapenems (90, 91)	Variable but consistent with extracellular water	Yes	Yes	1 (except ertapenem 4 hrs)	Low (except ertapenem)	Varies proportionately with renal function	No
Glycopeptides (17, 105)	0.2–1.6 (consistent with extracellular water)	Yes	Yes	4–6 (vancomycin) 80–160 (teicoplanin)	30% to 55% (vancomycin) 90% (teicoplanin)	Varies proportionately with renal function. Increased teicoplanin CL in hypoalbuminemia	Yes, to ensure plasma $C_{min} > 15$ mg/mL
Tigecycline (132–134)	7–10	Unlikely	Unlikely	37–66	73% to 79%	May decrease with cholestasis	No
Clindamycin (138, 140)	0.6–1.2	No	Yes	1.5–5	65% to 90%	Decreased hepatic CL	No
Linezolid (130)	0.5–0.6	Yes	Yes	3.5–7	31%	PK changes in critical illness probably not clinically significant	No
Colistin (143, 146, 147) ^a	0.18–1.5 (assuming 60 kg patient)	Likely	Likely	2–7.4	Unknown	Varies proportionately with renal function	No

Vd, volume of distribution; CL, clearance; PK, pharmacokinetic; TDM, therapeutic drug monitoring.

^aVery little accurate pharmacokinetic data exists for colistin because of a lack of reliable analytical methods (141).

Aminoglycosides. Dosing of aminoglycoside antibiotics has been vigorously debated in the literature because of the narrow therapeutic index of these drugs. The kill characteristic of the aminoglycosides is concentration dependent (50–54), with a significant postantibiotic effect that can prevent bacterial regrowth for prolonged periods should drug concentrations fall below the minimum inhibitory concentration (MIC) (51–54). Such pharmacodynamic properties have stimulated research that has supported once daily administration as opposed to small, multiple doses (50, 55–58). It is considered that high minimum concentration (C_{\min}), or more specifically the area under the concentration-time curve (AUC), are more closely correlated with the well-documented adverse renal and ototoxic effects of these hydrophilic drugs (50, 55, 56), although recent data suggest a genetic predisposition to ototoxicity (59).

Aminoglycosides often have increased Vd in critically ill patients that can result in a decreased maximum concentration (C_{\max}) (60–65). This increased Vd has been shown to increase proportionally with increasing levels of sickness severity (66). Maximal weight-based dosing (e.g., tobramycin/gentamicin 7 mg/kg) consistently achieves adequate C_{\max} :MIC ratios (62). In burn patients (61) and patients undergoing mechanical ventilation (65), an increased Vd has been shown to prolong $T_{1/2}$. However, creatinine clearance is likely to be more descriptive of aminoglycoside CL (67). Such pharmacokinetic variability and potential for adverse effects mandates that monitoring of plasma aminoglycoside concentrations is essential. Although Bayesian dosing methods may be used, use of dosing nomograms should be avoided as they have been invalidated in critically ill patients (62, 68). To optimize aminoglycoside:bacterial effectiveness in critically ill patients, extended-interval dosing with C_{\max} monitoring and MIC determination of the pathogen remains ideal practice. However, given the apparent success of mg/kg dosing (62), a low C_{\min} (preferably undetectable concentration) should be obtained to minimize aminoglycoside toxicity. Multiple doses per day should only be considered for the treatment of endocarditis or in neutropenic patients.

β -Lactam Antibiotics. The β -lactam group of antibiotics consists of penicillins, cephalosporins, and monobactams. Although these antibiotics are generally

hydrophilic molecules that are renally cleared with moderate-to-low protein binding, there is variability within this group [e.g., ceftriaxone has a longer $T_{1/2}$ —5.8 to 8.7 hours in adults—and high protein binding—>95% (34, 35)]. In conventional bolus dosing regimens, plasma concentrations of these antibiotics may fall to low levels between doses (33, 69, 70).

In vivo animal experiments have demonstrated that β -lactams have a slow continuous kill characteristic that is almost entirely related to $T > \text{MIC}$ (71). Data from a recent study by McKinnon et al (72) suggest that maintaining a $T > \text{MIC}$ of 100% is associated with significantly greater clinical cure (82% vs. 33% $p = 0.002$) and bacteriologic eradication (97% vs. 44%; $p < 0.001$) in patients with severe infections. Other studies have demonstrated maximum killing of bacteria at four to five times MIC (73, 74). As such, concentrations of β -lactam antibiotics should be maintained at four to five times the MIC for extended periods during each dosing period (51, 52, 54). This would be especially appropriate in patient groups likely to have compromised host-defenses, including critically ill patients. Research shows that an improved pharmacodynamic profile is obtained with either more frequent dosing or extended or continuous infusions (70, 73, 75–80). This mode of administration is likely to be of high value if the patient develops a high glomerular filtration rate and/or increased volume of distribution, which commonly occurs in critically ill patients receiving β -lactams (33, 70, 79, 81–83). Some data exist to suggest that some β -lactams, such as piperacillin and ticarcillin, are likely to have increased biliary CL in the absence of renal CL (40, 41). The clinical implications of this situation are likely to be significant when a patient develops moderate renal and hepatic dysfunction leading to greatly reduced antibiotic CL (84–86). Seizures have been noted with high β -lactam exposures but are relatively uncommon.

Support for reduced mortality from extended infusions (4-hour infusion every 8 hours) of piperacillin-tazobactam was described in a recent cohort study of 194 seriously ill patients with *Pseudomonas aeruginosa* infection by Lodise et al (87). In this study, patients receiving extended infusions with an Acute Physiology and Chronic Health Evaluation II score ≥ 17 had a significantly lower 14-day mortality rate (12.2% vs. 31.6%; $p = 0.04$) than

those receiving bolus infusions. Data on clinical cure superiority of continuous infusions of β -lactam antibiotics also exist. In a retrospective cohort study in patients with ventilator-associated pneumonia, Lorente et al (88) described superior clinical cure when given a continuous infusion (90.5%) compared with extended infusion (over 30 minutes; 59.6%). Roberts et al (89) described advantages for clinical cure in patients receiving ceftriaxone by continuous infusion when 4 or more days of therapy was required. No difference was found in the intention-to-treat analysis of this article, though. Further research is required to quantify the clinical utility of administering β -lactams as a continuous infusion.

Carbapenems. Carbapenems have very similar pharmacokinetic properties to β -lactams (Table 2). Pharmacodynamically, they are time-dependent antibiotics that have been reported to have maximal bactericidal activity when $T > \text{MIC}$ is maintained for a minimum of 40% of the dosing interval. In critical illness, carbapenems are likely to develop increased Vd and higher CL (90, 91). Continuous infusion of these antibiotics has been studied, as has administration by extended infusion, which is thought to be appropriate given the low % $T > \text{MIC}$ to optimize activity. Pharmacodynamic advantages to this method of dosing have been well described (92–94) and appear highly appropriate for use in critically ill patients.

Glycopeptides. Glycopeptides are relatively hydrophilic antibiotics that include vancomycin and teicoplanin. The optimal pharmacodynamic properties of glycopeptides have not been completely elucidated. Some *in vitro* and animal data suggest that the bactericidal activity of vancomycin is time dependent (95–97), whereas other data from a nonneutropenic mouse model found C_{\max} :MIC to correlate with efficacy (98). Other studies have proposed that AUC:MIC is the pharmacokinetic and pharmacodynamic parameter correlated with efficacy (10, 99).

As such there is little consensus on whether C_{\max} :MIC or $T > \text{MIC}$ should be maximized in dosing regimens. Previous studies examining continuous infusion of vancomycin have not provided conclusive results. Wsocki et al (100) found no clinical advantages for continuous infusion of vancomycin compared with intermittent dosing in 160 patients. However, recently Rello et al (101), described a suggestion of clinical superiority for continuous infusion of vancomycin in a subset of pa-

Table 3. General PK characteristics of fluoroquinolone antibiotics and possible changes that can occur during fluid shifts in critically ill patients

Fluoroquinolone	Vd (L/kg)	Increased Vd with Fluid Shifts?	Decreased C _{max} with Fluid Shifts?	Plasma T _{1/2} (hrs)	Protein Binding	Altered CL with Renal Dysfunction?	Normal Dose	Dose Adjustment in Renal Dysfunction?
Ciprofloxacin (157–159)	1.2–2.7	No	Yes	3 (4–5 hrs in the elderly)	20% to 40%	No	IV 400 mg 8 hourly	Yes
Levofloxacin (109–111)	0.92–1.36	No	Yes	6–8.9	24% to 38%	Yes	500–750 mg daily (May increase to 1000 mg daily in critically ill patients with sepsis)	a) CrCL = 20–49 mL/min → 250–500 mg daily; b) CrCL is 10–19 mL/min → 250–500 mg 48-hourly
Moxifloxacin (160–162)	2.45–3.55	No	Yes	9.3–15.6	39% to 52%	No	400 mg daily	No
Gatifloxacin (163, 164)	1.98–2.31	No	Yes	6.5–9.6	20%	Yes	400 mg daily	CrCL ≤40 mL/min → 400 mg initial dose followed by 200 mg 24-hourly

PK, pharmacokinetic; CL, clearance; CrCL, creatinine CL; C_{max}, maximum concentration; IV, intravenous.

tients treated for ventilator-associated pneumonia caused by methicillin-resistant *Staphylococcus aureus*.

Like most β-lactams, glycopeptide CL is closely related to creatinine clearance. Nonrenal CL of vancomycin has been well described and shown to increase in patients with acute renal failure, although it displays significant variability among patients (102). In obese patients, weight-based dosing (~30 mg/kg) that uses total body weight appears appropriate although such patients may require more frequent dosing (103). Therefore, empirical dosing based on creatinine clearance data with subsequent therapeutic drug monitoring of C_{min} plasma concentrations (suggested C_{min} 15–20 mg/L) is recommended (104–106). It should be noted that recent data report higher rates of nephrotoxicity with high vancomycin dosing when higher C_{min} concentrations (≥15 mg/L) are present (107). Nephrotoxicity will be potentiated by coadministration with other nephrotoxic drugs such as aminoglycosides or amphotericin.

Fluoroquinolones. Fluoroquinolones are lipophilic antibiotics that include ciprofloxacin, moxifloxacin, levofloxacin, and gatifloxacin. All fluoroquinolones have extensive distribution characteristics and achieve good extracellular and intracellular concentrations with excellent penetration of neutrophils and lymphocytes (108). The Vd of most fluoroquinolones is minimally affected in the critically ill patient, although levofloxacin requires increased dosing in critically

ill patients because of a decreased T_{1/2} (resulting in an AUC reduced by 30% to 40%) (109–111). The pharmacokinetics of selected fluoroquinolones are described in Table 3.

Fluoroquinolones not only display largely concentration-dependent kill characteristics, but also some time-dependent effects. Previous research has suggested that achieving a C_{max}:MIC ratio of 10 for ciprofloxacin is the critical variable in predicting bacterial eradication (112–114). Forrest et al studied ciprofloxacin in critically ill patients and concluded that achieving an AUC:MIC greater than 125 is associated with a successful clinical outcome (115). This result is necessary for Gram-negative organisms with Gram-positive organisms requiring an AUC:MIC of 30 (115–118). Inappropriate low dosing of ciprofloxacin has also been associated with the emergence of resistant bacterial strains (particularly enterococci, pseudomonas and methicillin-resistant *Staphylococcus aureus*) (9, 119–121). For Gram-negative bacteria, this may occur when AUC:MIC <100 (122, 123). Therefore, AUC:MIC and C_{max}:MIC are pharmacodynamic variables that require close attention for optimal fluoroquinolone usage. Dosing should seek to maximize C_{max}:MIC as this will drive adequate AUC:MIC exposures. The principal adverse effects that may occur with drug toxicity include QT-interval prolongation as well as confusion and dizziness. The latter two effects may affect any cognition evalu-

ations by the healthcare staff of critically ill patients.

Linezolid. Linezolid belongs to a new class of antimicrobials called the *oxazolidinone*. Although linezolid is quite hydrophilic, it distributes widely into tissues and is mostly metabolized hepatically before being cleared renally (124, 125). At this time, no dose adjustment is recommended in renal dysfunction or hepatic dysfunction (125, 126). From a pharmacodynamic perspective, maintaining a T > MIC of 40% to 80% is thought to be the major predictor of efficacy (10, 127–129). A 600-mg 12-hourly dose should achieve this ratio in humans against susceptible organisms with MICs up to 2–4 mg/L. Linezolid T_{1/2} has been shown to be shorter and Vd is larger in critically ill patients, although these are probably not significant (130).

A significant area of interest for the intensivist should be the potential for adverse effects associated with linezolid and drug interaction with other agents that may inhibit monoamine oxidase (125). Although linezolid is generally safe and well tolerated for up to 28 days at 600 mg twice daily (131), evidence exists that therapy longer than 14 days can cause reversible myelosuppression (132). As such, as part of individualized patient-specific therapy, patients prescribed linezolid may require complete blood counts ordered [up to weekly (131)] to monitor for hematologic adverse effects.

Tigecycline. Tigecycline is a member of the glycylcyclines that are novel tetracyclines with Gram-positive and Gram-

negative activity. Pharmacokinetically, tigecycline possesses lipophilic characteristics that enable rapid and extensive penetration into body tissues (133). It is primarily eliminated by biliary excretion with only 15% of the dose eliminated unchanged in urine (134). There are few data to support potentially altered pharmacokinetics in critically ill patients. Pharmacodynamically, although tigecycline displays time-dependent killing against some bacteria (135), AUC:MIC is more likely to be correlated with efficacy (132, 133). This is because of its long $T_{1/2}$ and prolonged postantibiotic effect.

Lincosamides. The lincosamide antibiotics include clindamycin and lincomycin. This lipophilic class of antibiotics achieves wide distribution throughout the body and achieves therapeutic concentrations in most body compartments (136–138). $T > MIC$ has been determined to be the pharmacodynamic factor correlated with efficacy. Free drug levels of

lincosamides should exceed the MIC of the infective pathogen for at least 40% to 50% of the dosing interval (139). Hepatic CL of clindamycin is documented to decrease in critically ill patients with sepsis (140). Antibiotic-associated diarrhea is a significant adverse effect for this class of antibiotics.

Colistin. The polymyxin antibiotics (e.g., colistin) were first used in the 1960s and subsequently lost appeal because of associated nephro- and neurotoxicities (141). With the escalation of antibiotic multi-drug resistance, it is now being increasingly used as an alternate antibiotic. Colistin is administered typically as colistimethate sodium (sodium colistin methanesulphate). This molecule is hydrolyzed to sulfomethylated derivatives and colistin (142). It is a hydrophilic molecule for which little pharmacokinetic information exists (143). Pharmacodynamically, it is thought to have predominantly concentration-dependent bacterial killing activity (141, 144, 145).

Colistin is available as two formulations Colomycin Injection (40, 80, and 160 mg colistimethate per vial) and Colymycin Parenteral (400 mg colistimethate per vial, Forest Laboratories, Bexley, UK), which contain differing amounts of colistimethate sodium per vial; ERFA, Montreal, Canada. Patient dosing should follow weight and renal dysfunction considerations where possible (146). Comparative data between critically ill patients and other patients is currently lacking and dosing remains a difficult issue with little consensus on appropriate dosing strategies existing (143, 146, 147).

General Dosing Considerations. The importance of effective antibiotic therapy in the ICU mandates intensivists to select effective dosing regimens for critically ill patients. Table 4 proposes some general dosing recommendations that could be considered to this end. However, because of the large spectrum of different ICU admission diagnoses and the effect of the

Table 4. Broad guidelines that can be used to assist antibiotic dosing adjustment for critically ill patients

Antibiotic Class	Suggested Dosing Adjustment for Critically Ill Patients	
	Normal Renal Function	Moderate to Severe Renal Dysfunction Comments
Aminoglycosides	Use high doses (e.g., gentamicin 7 mg/kg) where possible to target C_{max} :MIC ratio of 10; monitor C_{min} and aim for undetectable plasma concentrations ^a	Use high doses where possible and monitor C_{min} thereafter (36 to 48 hourly extended interval dosing acceptable); dosing can be guided by MIC data if available if dose reductions are essential
β -lactams	Consider extended or continuous infusion or more frequent dosing to ensure $T > MIC$; therapeutic drug monitoring may be useful if available	If intermittent dosing used, dosing can occur at reduced dose or frequency (not both); err toward larger doses as β -lactams have large therapeutic window
Carbapenems		
Glycopeptides	Dosing at 30–40 mg/kg/day (vancomycin), which can be increased according to C_{min} plasma concentrations (aim for 15–20 mg/L); continuous infusions should be used when difficulty obtaining therapeutic C_{min}	High dosing on day 1 may be required to ensure adequate distribution; dose adjustments should occur according to C_{min} concentrations
Fluoroquinolones	Doses that achieve high C_{max} :MIC ratio should be targeted (e.g. ciprofloxacin 1200 mg/day); levofloxacin may require 500 mg 12-hourly in some patients with high creatinine clearance; where high doses used, monitor for toxicity (seizures)	Dose adjustment is probably only required in renal impairment for levofloxacin, gatifloxacin, and ciprofloxacin; where possible reduce frequency and maintain dose
Tigecycline	Use 100 mg loading dose then 50 mg 12 hourly	No dose adjustment required in renal failure or dialysis ^b
Linezolid	Use 600 mg 12 hourly	No dosage adjustment required in renal failure or dialysis
Lincosamides	Use 600–900 mg 8 hourly	Decreased lincomycin dose or frequency in renal or hepatic dysfunction; decrease clindamycin dose or frequency in hepatic dysfunction
Colistin	Use 5 mg/kg/day of colistin base (75,000 international units/kg/day colistimethate sodium) ^c intravenously in 3 divided doses	Reduce dose or frequency (not both)

MIC, minimum inhibitory concentration; C_{max} , maximum concentration; C_{min} , minimum concentration.

^aAminoglycoside levels should be undetectable for no more than the post-antibiotic effect. We recommend a maximum of 4 hrs before redosing as any longer delay may enable bacterial regrowth; ^bif severe cholestasis present then tigecycline should be dosed with 50-mg loading dose, then 25 mg 12 hourly; ^c1 mg colistimethate sodium is equivalent to 12,500 international units (165).

different levels of organ function and pathophysiological changes that may be observed in these patients, it is not possible to provide specific dosing recommendations for each potential patient. Standard considerations of potential for adverse effects and drug interactions should always be considered as part of the antibiotic-prescribing process and any ongoing monitoring performed by the clinician or associated healthcare staff.

Appropriate Dosing May Reduce Development of Antibiotic Resistance. Antibiotic resistance continues to escalate worldwide with the ICU being a particular focus on further development. There is now sufficient data to suggest that inappropriately low antibiotic dosing may be contributing to the increasing rate of antibiotic resistance (9, 123, 148–154). Developing dosing regimens that adhere to pharmacodynamic principles and maximize antibiotic exposure appears to be essential to reduce the development of antibiotic resistance. This is probably best achieved by administering the highest recommended dose to the patient.

CONCLUSION

In summary, the solubility characteristics of antibiotics can help determine where dose adjustment may be necessary for individual critically ill patients. Hydrophilic concentration-dependent antibiotics may possess a higher V_d in critically ill patients leading to a reduced C_{max} . It follows that hydrophilic time-dependent antibiotics may develop a low C_{min} that may reduce antibiotic efficacy. Common increases in V_d need to be contrasted against potential increased or reduced antibiotic CL that can occur in these patients. Antibiotic underdosing can occur, which may in turn lead to the development of antibiotic resistance and/or therapeutic failure, if appropriate dosing adjustments are not made. For renally cleared compounds, dose prescription based on measured creatinine clearance should enable appropriate initial dosing in critically ill patients. Where possible thereafter, therapeutic drug monitoring should be considered to ensure target plasma concentrations are being achieved.

Given that most antibiotic regimens have been derived from trials with patients who are not critically ill, the intensivist must adapt his/her dosing to account for the potential altered pathophysiology of this patient group. To

optimize dosing, the antibiotic's pharmacodynamic properties, as well as the potential altered antibiotic pharmacokinetics, need to be considered by the clinician. Such a process will enable dose selection that is more appropriate for use in the individual patient.

REFERENCES

- Rice TW, Bernard GR: Therapeutic intervention and targets for sepsis. *Annu Rev Med* 2005; 56:225–248
- Garnacho-Montero J, Garcia-Garmendia JL, Barrero-Almodovar A, et al: Impact of adequate empirical antibiotic therapy on the outcome of patients admitted to the intensive care unit with sepsis. *Crit Care Med* 2003; 31:2742–2751
- Harbarth S, Garbino J, Pugin J, et al: Inappropriate initial antimicrobial therapy and its effect on survival in a clinical trial of immunomodulating therapy for severe sepsis. *Am J Med* 2003; 115:529–535
- Hugonnet S, Harbarth S, Ferriere K, et al: Bacteremic sepsis in intensive care: Temporal trends in incidence, organ dysfunction, and prognosis. *Crit Care Med* 2003; 31:390–394
- Kollef MH, Sherman G, Ward S, et al: Inadequate antimicrobial treatment of infections: A risk factor for hospital mortality among critically ill patients. *Chest* 1999; 115:462–474
- Kumar A, Roberts D, Wood KE, et al: Duration of hypotension before initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. *Crit Care Med* 2006; 34:1589–1596
- Nicolau DP: Optimizing antimicrobial therapy and emerging pathogens. *Am J Manag Care* 1998; 4(Suppl 2):S525–S530
- Nicolau DP: Optimizing outcomes with antimicrobial therapy through pharmacodynamic profiling. *J Infect Chemother* 2003; 9:292–296
- Roberts JA, Kruger P, Paterson DL, et al: Antibiotic resistance—What's dosing got to do with it? *Crit Care Med* 2008; 36:2433–2440
- Craig WA: Basic pharmacodynamics of antibacterials with clinical applications to the use of β -lactams, glycopeptides, and linezolid. *Infect Dis Clin North Am* 2003; 17:479–501
- Calandra T, Cohen J: The international sepsis forum consensus conference on definitions of infection in the intensive care unit. *Crit Care Med* 2005; 33:1538–1548
- Grocott-Mason RM, Shah AM: Cardiac dysfunction in sepsis: New theories and clinical implications. *Intensive Care Med* 1998; 24:286–295
- Parrillo JE: Pathogenetic mechanisms of septic shock. *N Engl J Med* 1993; 328:1471–1477
- van der Poll T: Immunotherapy of sepsis. *Lancet Infect Dis* 2001; 1:165–174
- Gosling P, Sanghera K, Dickson G: Generalized vascular permeability and pulmonary function in patients following serious trauma. *J Trauma* 1994; 36:477–481
- Nuytink HK, Offermans XJ, Kubat K, et al: Whole-body inflammation in trauma patients. An autopsy study. *Arch Surg* 1988; 123:1519–1524
- Barbot A, Venisse N, Rayeh F, et al: Pharmacokinetics and pharmacodynamics of sequential intravenous and subcutaneous teicoplanin in critically ill patients without vasopressors. *Intensive Care Med* 2003; 29:1528–1534
- Buijk SL, Gyssens IC, Mouton JW, et al: Pharmacokinetics of ceftazidime in serum and peritoneal exudate during continuous versus intermittent administration to patients with severe intra-abdominal infections. *J Antimicrob Chemother* 2002; 49:121–128
- Conil JM, Georges B, Lavit M, et al: A population pharmacokinetic approach to ceftazidime use in burn patients: Influence of glomerular filtration, gender and mechanical ventilation. *Br J Clin Pharmacol* 2007; 64:27–35
- Roberts JA, Roberts MS, Robertson TA, et al: A novel way to investigate the effects of plasma exchange on antibiotic levels: Use of microdialysis. *Int J Antimicrob Agents* 2008; 31:240–244
- Rowland M, Tozer TN: Clinical Pharmacokinetics—Concepts and Applications. Third Edition. Philadelphia, Lippincott Williams & Wilkins, 1995
- Pea F, Porreca L, Baraldo M, et al: High vancomycin dosage regimens required by intensive care unit patients cotreated with drugs to improve haemodynamics following cardiac surgical procedures. *J Antimicrob Chemother* 2000; 45:329–335
- Di Giantomasso D, Bellomo R, May CN: The haemodynamic and metabolic effects of epinephrine in experimental hyperdynamic septic shock. *Intensive Care Med* 2005; 31:454–462
- Di Giantomasso D, May CN, Bellomo R: Norepinephrine and vital organ blood flow. *Intensive Care Med* 2002; 28:1804–1809
- Di Giantomasso D, May CN, Bellomo R: Norepinephrine and vital organ blood flow during experimental hyperdynamic sepsis. *Intensive Care Med* 2003; 29:1774–1781
- Pong S, Seto W, Abdollell M, et al: 12-hour versus 24-hour creatinine clearance in critically ill pediatric patients. *Pediatr Res* 2005; 58:83–88
- Wells M, Lipman J: Measurements of glomerular filtration in the intensive care unit are only a rough guide to renal function. *S Afr J Surg* 1997; 35:20–23
- Herrera-Gutierrez ME, Sellar-Perez G, Banderas-Bravo E, et al: Replacement of 24-h creatinine clearance by 2-h creatinine clearance in

- intensive care unit patients: A single-center study. *Intensive Care Med* 2007; 33:1900–1906
29. Cockcroft DW, Gault MH: Prediction of creatinine clearance from serum creatinine. *Nephron* 1976; 16:31–41
 30. Levey AS, Bosch JP, Lewis JB, et al: A more accurate method to estimate glomerular filtration rate from serum creatinine: A new prediction equation. Modification of Diet in Renal Disease Study Group. *Ann Intern Med* 1999; 130:461–470
 31. Hoste EA, Damen J, Vanholder RC, et al: Assessment of renal function in recently admitted critically ill patients with normal serum creatinine. *Nephrol Dial Transplant* 2005; 20:747–753
 32. Lipman J, Gous AG, Mathivha LR, et al: Ciprofloxacin pharmacokinetic profiles in paediatric sepsis: How much ciprofloxacin is enough? *Intensive Care Med* 2002; 28: 493–500
 33. Lipman J, Wallis SC, Rickard CM, et al: Low ceftiofome levels during twice daily dosing in critically ill septic patients: Pharmacokinetic modelling calls for more frequent dosing. *Intensive Care Med* 2001; 27:363–370
 34. McNamara PJ, Gibaldi M, Stoeckel K: Volume of distribution terms for a drug (ceftriaxone) exhibiting concentration-dependent protein binding. II. Physiological significance. *Eur J Clin Pharmacol* 1983; 25: 407–412
 35. McNamara PJ, Gibaldi M, Stoeckel K: Volume of distribution terms for a drug (ceftriaxone) exhibiting concentration-dependent protein binding. I. Theoretical considerations. *Eur J Clin Pharmacol* 1983; 25: 399–405
 36. Joynt GM, Lipman J, Gomersall CD, et al: The pharmacokinetics of once-daily dosing of ceftriaxone in critically ill patients. *J Antimicrob Chemother* 2001; 47:421–429
 37. Parrillo JE, Parker MM, Natanson C, et al: Septic shock in humans. Advances in the understanding of pathogenesis, cardiovascular dysfunction, and therapy. *Ann Intern Med* 1990; 113:227–242
 38. Marshall JC: Inflammation, coagulopathy, and the pathogenesis of multiple organ dysfunction syndrome. *Crit Care Med* 2001; 29(Suppl 7):S99–S106
 39. Rohwedder R, Bergan T, Thorsteinsson SB, et al: Transintestinal elimination of ciprofloxacin. *Chemotherapy* 1990; 36:77–84
 40. Brogard JM, Jehl F, Blicke JF, et al: Biliary elimination of ticarcillin plus clavulanic acid (Claventin): Experimental and clinical study. *Int J Clin Pharmacol Ther Toxicol* 1989; 27:135–144
 41. Brogard JM, Jehl F, Blicke JF, et al: Biliary pharmacokinetic profile of piperacillin: experimental data and evaluation in man. *Int J Clin Pharmacol Ther Toxicol* 1990; 28: 462–470
 42. Livornese LL, Slavin D, Benz RL, et al: Use of antibacterial agents in renal failure. *Infect Dis Clin North Am* 2001; 15:983–1002
 43. Livornese LL, Slavin D, Gilbert B, et al: Use of antibacterial agents in renal failure. *Infect Dis Clin North Am* 2004; 18:551–579
 44. Trotman RL, Williamson JC, Shoemaker DM, et al: Antibiotic dosing in critically ill adult patients receiving continuous renal replacement therapy. *Clin Infect Dis* 2005; 41:1159–1166
 45. Ryan DM: Pharmacokinetics of antibiotics in natural and experimental superficial compartments in animals and humans. *J Antimicrob Chemother* 1993; 31(Suppl D):1–16
 46. Dahyot C, Marchand S, Bodin M, et al: Application of basic pharmacokinetic concepts to analysis of microdialysis data: Illustration with imipenem muscle distribution. *Clin Pharmacokinet* in press
 47. Joukhadar C, Frossard M, Mayer BX, et al: Impaired target site penetration of beta-lactams may account for therapeutic failure in patients with septic shock. *Crit Care Med* 2001; 29:385–391
 48. Roberts JA, Roberts MS, Robertson TA, et al: Piperacillin penetration into tissue of critically ill patients with sepsis—Bolus vs continuous administration? *Crit Care Med* 2009; 37:926–933
 49. Sauermann R, Delle-Karth G, Marsik C, et al: Pharmacokinetics and pharmacodynamics of ceftiofome in subcutaneous adipose tissue of septic patients. *Antimicrob Agents Chemother* 2005; 49:650–655
 50. Olsen KM, Rudis MI, Rebeck JA, et al: Effect of once-daily dosing vs. multiple daily dosing of tobramycin on enzyme markers of nephrotoxicity. *Crit Care Med* 2004; 32: 1678–1682
 51. Roosendaal R, Bakker-Woudenberg IA, van den Bergh-van Raffe M, et al: Impact of the dosage schedule on the efficacy of ceftazidime, gentamicin and ciprofloxacin in *Klebsiella pneumoniae* pneumonia and septicemia in leukopenic rats. *Eur J Clin Microbiol Infect Dis* 1989; 8:878–887
 52. Vogelmann B, Craig WA: Kinetics of antimicrobial activity. *J Pediatr* 1986; 108: 835–840
 53. Vogelmann B, Gudmundsson S, Leggett J, et al: Correlation of antimicrobial pharmacokinetic parameters with therapeutic efficacy in an animal model. *J Infect Dis* 1988; 158: 831–847
 54. Vogelmann BS, Craig WA: Postantibiotic effects. *J Antimicrob Chemother* 1985; 15 (Suppl A):37–46
 55. Ali MZ, Goetz MB: A meta-analysis of the relative efficacy and toxicity of single daily dosing versus multiple daily dosing of aminoglycosides. *Clin Infect Dis* 1997; 24: 796–809
 56. Bailey TC, Little JR, Littenberg B, et al: A meta-analysis of extended-interval dosing versus multiple daily dosing of aminoglycosides. *Clin Infect Dis* 1997; 24:786–795
 57. Marik PE, Lipman J, Kobalski S, et al: A prospective randomized study comparing once-versus twice-daily amikacin dosing in critically ill adult and paediatric patients. *J Antimicrob Chemother* 1991; 28:753–764
 58. Prins JM, Buller HR, Kuijper EJ, et al: Once versus thrice daily gentamicin in patients with serious infections. *Lancet* 1993; 341: 335–339
 59. Bitner-Grindzic M, Rahman S: Ototoxicity caused by aminoglycosides. *Br Med J* 2007; 335:784–785
 60. Beckhouse MJ, Whyte IM, Byth PL, et al: Altered aminoglycoside pharmacokinetics in the critically ill. *Anaesth Intensive Care* 1988; 16:418–422
 61. Bracco D, Landry C, Dubois MJ, et al: Pharmacokinetic variability of extended interval tobramycin in burn patients. *Burns* 2008; 34:791–796
 62. Buijk SE, Mouton JW, Gyssens IC, et al: Experience with a once-daily dosing program of aminoglycosides in critically ill patients. *Intensive Care Med* 2002; 28: 936–942
 63. Moller JC, Gilman JT, Kearns GL, et al: Effect of extracorporeal membrane oxygenation on tobramycin pharmacokinetics in sheep. *Crit Care Med* 1992; 20:1454–1458
 64. Trigriner C, Izquierdo I, Fernandez R, et al: Gentamicin volume of distribution in critically ill septic patients. *Intensive Care Med* 1990; 16:303–306
 65. Trigriner C, Izquierdo I, Fernandez R, et al: Changes in gentamicin pharmacokinetic profiles induced by mechanical ventilation. *Eur J Clin Pharmacol* 1991; 40:297–302
 66. Marik PE: Aminoglycoside volume of distribution and illness severity in critically ill septic patients. *Anaesth Intensive Care* 1993; 21:172–173
 67. Tang GJ, Tang JJ, Lin BS, et al: Factors affecting gentamicin pharmacokinetics in septic patients. *Acta Anaesthesiol Scand* 1999; 43:726–730
 68. Toschlog EA, Blount KP, Rotondo MF, et al: Clinical predictors of subtherapeutic aminoglycoside levels in trauma patients undergoing once-daily dosing. *J Trauma* 2003; 55:255–260; discussion 260–252
 69. Lipman J, Crewe-Brown HH, Saunders GL, et al: Subtleties of antibiotic dosages—Do doses and intervals make a difference in the critically ill? *S Afr J Surg* 1996; 34:160–162
 70. Lipman J, Wallis SC, Rickard C: Low plasma ceftiofome levels in critically ill septic patients: Pharmacokinetic modeling indicates improved troughs with revised dosing. *Antimicrob Agents Chemother* 1999; 43: 2559–2561
 71. Craig WA: Pharmacokinetic/pharmacodynamic parameters: Rationale for antibacterial dosing of mice and men. *Clin Infect Dis* 1998; 26:1–10
 72. McKinnon PS, Paladino JA, Schentag JJ: Evaluation of area under the inhibitory curve (AUC) and time above the minimum inhibitory concentration ($T > MIC$) as predictors of outcome for ceftiofome and ceftazidime in serious bacterial infections. *Int J Antimicrob Agents* 2008; 31:345–351
 73. Angus BJ, Smith MD, Suputtamongkol Y, et al: Pharmacokinetic-pharmacodynamic

- evaluation of ceftazidime continuous infusion vs intermittent bolus injection in septicemic melioidosis. *Br J Clin Pharmacol* 2000; 50:184–191
74. Mouton JW, den Hollander JG: Killing of *Pseudomonas aeruginosa* during continuous and intermittent infusion of ceftazidime in an in vitro pharmacokinetic model. *Antimicrob Agents Chemother* 1994; 38:931–936
 75. Boselli E, Breilh D, Cannesson M, et al: Steady-state plasma and intrapulmonary concentrations of piperacillin/tazobactam 4 g/0.5 g administered to critically ill patients with severe nosocomial pneumonia. *Intensive Care Med* 2004; 30:976–979
 76. Burgess DS, Waldrep TP: Pharmacokinetics and pharmacodynamics of piperacillin/tazobactam when administered by continuous infusion and intermittent dosing. *Clin Ther* 2002; 24:1090–1104
 77. Georges B, Conil JM, Cougot P, et al: Cefepime in critically ill patients: Continuous infusion vs. an intermittent dosing regimen. *Int J Clin Pharmacol Ther* 2005; 43:360–369
 78. Jaruratanasirikul S, Sriwiriyan S, Ingviya N: Continuous infusion versus intermittent administration of cefepime in patients with Gram-negative bacilli bacteraemia. *J Pharm Pharmacol* 2002; 54:1693–1696
 79. Lipman J, Gomersall CD, Gin T, et al: Continuous infusion ceftazidime in intensive care: A randomized controlled trial. *J Antimicrob Chemother* 1999; 43:309–311
 80. Mouton JW, Vinks AA: Pharmacokinetic/pharmacodynamic modelling of antibacterials in vitro and in vivo using bacterial growth and kill kinetics: The minimum inhibitory concentration versus stationary concentration. *Clin Pharmacokinet* 2005; 44:201–210
 81. Bakker-Woudenberg IA, Roosendaal R: Impact of dosage schedule of antibiotics on the treatment of serious infections. *Intensive Care Med* 1990; 16:S229–S234
 82. Gomez CM, Cordingly JJ, Palazzo MG: Altered pharmacokinetics of ceftazidime in critically ill patients. *Antimicrob Agents Chemother* 1999; 43:1798–1802
 83. Hanes SD, Wood GC, Herring V, et al: Intermittent and continuous ceftazidime infusion for critically ill trauma patients. *Am J Surg* 2000; 179:436–440
 84. Cooper BE, Nester TJ, Armstrong DK, et al: High serum concentrations of mezlocillin in a critically ill patient with renal and hepatic dysfunction. *Clin Pharm* 1986; 5:764–766
 85. Green L, Dick JD, Goldberger SP, et al: Prolonged elimination of piperacillin in a patient with renal and liver failure. *Drug Intell Clin Pharm* 1985; 19:427–429
 86. Hoffman TA, Cestero R, Bullock WE: Pharmacokinetics of carbenicillin in patients with hepatic and renal failure. *J Infect Dis* 1970; 122(Suppl):S75–S77
 87. Lodise TP Jr., Lomaestro B, Drusano GL: Piperacillin-tazobactam for *Pseudomonas aeruginosa* infection: Clinical implications of an extended-infusion dosing strategy. *Clin Infect Dis* 2007; 44:357–363
 88. Lorente L, Lorenzo L, Martin MM, et al: Meropenem by continuous versus intermittent infusion in ventilator-associated pneumonia due to gram-negative bacilli. *Ann Pharmacother* 2006; 40:219–223
 89. Roberts JA, Boots R, Rickard CM, et al: Is continuous infusion ceftriaxone better than once-a-day dosing in intensive care? A randomized controlled pilot study. *J Antimicrob Chemother* 2007; 59:285–291
 90. Kitzes-Cohen R, Farin D, Piva G, et al: Pharmacokinetics and pharmacodynamics of meropenem in critically ill patients. *Int J Antimicrob Agents* 2002; 19:105–110
 91. Novelli A, Adembri C, Livi P, et al: Pharmacokinetic evaluation of meropenem and imipenem in critically ill patients with sepsis. *Clin Pharmacokinet* 2005; 44:539–549
 92. Jaruratanasirikul S, Sriwiriyan S, Punyo J: Comparison of the pharmacodynamics of meropenem in patients with ventilator-associated pneumonia following administration by 3-hour infusion or bolus injection. *Antimicrob Agents Chemother* 2005; 49:1337–1339
 93. Li C, Kuti JL, Nightingale CH, et al: Population pharmacokinetic analysis and dosing regimen optimization of meropenem in adult patients. *J Clin Pharmacol* 2006; 46:1171–1178
 94. Lomaestro BM, Drusano GL: Pharmacodynamic evaluation of extending the administration time of meropenem using a Monte Carlo simulation. *Antimicrob Agents Chemother* 2005; 49:461–463
 95. Chambers HF, Kennedy S: Effects of dosage, peak and trough concentrations in serum, protein binding, and bacterial rate on efficacy of teicoplanin in a rabbit model of endocarditis. *Antimicrob Agents Chemother* 1990; 47:2018–2021
 96. Larsson AJ, Walker KJ, Raddatz JK, et al: The concentration-independent effect of monoexponential and biexponential decay in vancomycin concentrations on the killing of *Staphylococcus aureus* under aerobic and anaerobic conditions. *J Antimicrob Chemother* 1996; 38:589–597
 97. Lowdin E, Odenholt I, Cars O: In vitro studies of pharmacodynamic properties of vancomycin against *Staphylococcus aureus* and *Staphylococcus epidermidis*. *Antimicrob Agents Chemother* 1998; 42:2739–2744
 98. Knudsen JD, Fuursted K, Raber S, et al: Pharmacodynamics of glycopeptides in the mouse peritonitis model of *Streptococcus pneumoniae* or *Staphylococcus aureus* infection. *Antimicrob Agents Chemother* 2000; 44:1247–1254
 99. Rybak MJ: The pharmacokinetic and pharmacodynamic properties of vancomycin. *Clin Infect Dis* 2006; 42(Suppl 1):S35–S39
 100. Wysocki M, Delatour F, Faurisson F, et al: Continuous versus intermittent infusion of vancomycin in severe Staphylococcal infections: Prospective multicenter randomized study. *Antimicrob Agents Chemother* 2001; 45:2460–2467
 101. Rello J, Sole-Violan J, Sa-Borges M, et al: Pneumonia caused by oxacillin-resistant *Staphylococcus aureus* treated with glycopeptides. *Crit Care Med* 2005; 33:1983–1987
 102. Macias WL, Mueller BA, Scarim SK: Vancomycin pharmacokinetics in acute renal failure: Preservation of nonrenal clearance. *Clin Pharmacol Ther* 1991; 50:688–694
 103. Bauer LA, Black DJ, Lill JS: Vancomycin dosing in morbidly obese patients. *Eur J Clin Pharmacol* 1998; 54:621–625
 104. Fernandez de Gatta MD, Calvo MV, Hernandez JM, et al: Cost-effectiveness analysis of serum vancomycin concentration monitoring in patients with hematologic malignancies. *Clin Pharmacol Ther* 1996; 60:332–340
 105. Llopis-Salvia P, Jimenez-Torres NV: Population pharmacokinetic parameters of vancomycin in critically ill patients. *J Clin Pharm Ther* 2006; 31:447–454
 106. MacGowan AP: Pharmacodynamics, pharmacokinetics, and therapeutic drug monitoring of glycopeptides. *Ther Drug Monit* 1998; 20:473–477
 107. Hidayat LK, Hsu DI, Quist R, et al: High-dose vancomycin therapy for methicillin-resistant *Staphylococcus aureus* infections: Efficacy and toxicity. *Arch Intern Med* 2006; 166:2138–2144
 108. Smith RP, Baltch AL, Franke MA, et al: Levofloxacin penetrates human monocytes and enhances intracellular killing of *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *J Antimicrob Chemother* 2000; 45:483–488
 109. Kiser TH, Hoody DW, Obritsch MD, et al: Levofloxacin pharmacokinetics and pharmacodynamics in patients with severe burn injury. *Antimicrob Agents Chemother* 2006; 50:1937–1945
 110. Pea F, Di Qual E, Cusenza A, et al: Pharmacokinetics and pharmacodynamics of intravenous levofloxacin in patients with early-onset ventilator-associated pneumonia. *Clin Pharmacokinet* 2003; 42:589–598
 111. Rebuck JA, Fish DN, Abraham E: Pharmacokinetics of intravenous and oral levofloxacin in critically ill adults in a medical intensive care unit. *Pharmacotherapy* 2002; 22:1216–1225
 112. Blaser J, Stone BB, Groner MC, et al: Comparative study with enoxacin and netilmicin in a pharmacodynamic model to determine importance of ratio of antibiotic peak concentration to MIC for bactericidal activity and emergence of resistance. *Antimicrob Agents Chemother* 1987; 31:1054–1060
 113. Dudley MN, Blaser J, Gilbert D, et al: Combination therapy with ciprofloxacin plus azlocillin against *Pseudomonas aeruginosa*:

- Effect of simultaneous versus staggered administration in an in vitro model of infection. *J Infect Dis* 1991; 164:499–506
114. Preston SL, Drusano GL, Berman AL: Pharmacodynamics of levofloxacin: A new paradigm for early clinical trials. *JAMA* 1998; 279:125–129
 115. Forrest A, Nix DE, Ballow CH, et al: Pharmacodynamics of intravenous ciprofloxacin in seriously ill patients. *Antimicrob Agents Chemother* 1993; 37:1073–1081
 116. Goss TF, Forrest A, Nix DE, et al: Mathematical examination of dual individualization principles (II): The rate of bacterial eradication at the same area under the inhibitory curve is more rapid for ciprofloxacin than for cefmenoxime. *Ann Pharmacother* 1994; 28:863–868
 117. Nix DE, Spivey JM, Norman A, et al: Dose-ranging pharmacokinetic study of ciprofloxacin after 200-, 300-, and 400-mg intravenous doses. *Ann Pharmacother* 1992; 26: 8–10
 118. Piddock LJ, Dalhoff A: Should quinolones be used in the treatment of bacterial infections in neutropenic patients? *J Antimicrob Chemother* 1993; 32:771–774
 119. Hyatt JM, Schentag JJ: Pharmacodynamic modeling of risk factors for ciprofloxacin resistance in *Pseudomonas aeruginosa*. *Infect Control Hosp Epidemiol* 2000; 21(Suppl 1):S9–S11
 120. MacGowan A, Rogers C, Bowker K: The use of in vitro pharmacodynamic models of infection to optimize fluoroquinolone dosing regimens. *J Antimicrob Chemother* 2000; 46:163–170
 121. Zhou J, Dong Y, Zhao X, et al: Selection of antibiotic-resistant bacterial mutants: Allelic diversity among fluoroquinolone-resistant mutations. *J Infect Dis* 2000; 182: 517–525
 122. Schentag JJ: Antimicrobial action and pharmacokinetics/pharmacodynamics: The use of AUC to improve efficacy and avoid resistance. *J Chemother* 1999; 11:426–439
 123. Thomas JK, Forrest A, Bhavnani SM, et al: Pharmacodynamic evaluation of factors associated with the development of bacterial resistance in acutely ill patients during therapy. *Antimicrob Agents Chemother* 1998; 42:521–527
 124. Boselli E, Breil D, Rimmel T, et al: Pharmacokinetics and intrapulmonary concentrations of linezolid administered to critically ill patients with ventilator-associated pneumonia. *Crit Care Med* 2005; 33: 1529–1533
 125. MacGowan AP: Pharmacokinetic and pharmacodynamic profile of linezolid in healthy volunteers and patients with Gram-positive infections. *J Antimicrob Chemother* 2003; 51 (Suppl 2):ii17–25
 126. Brier ME, Stalker DJ, Aronoff GR, et al: Pharmacokinetics of linezolid in subjects with renal dysfunction. *Antimicrob Agents Chemother* 2003; 47:2775–2780
 127. Andes D, van Ogtrop ML, Craig W: Pharmacodynamic activity of a new oxazolidinone linezolid in an animal infection model. *In: 38th Interscience Conference on Antimicrobial Agents and Chemotherapy*: 1998. San Diego, CA, American Society for Microbiology, 1998, p 3
 128. Andes D, van Ogtrop ML, Peng J, et al: In vivo pharmacodynamics of a new oxazolidinone (linezolid). *Antimicrob Agents Chemother* 2002; 46:3484–3489
 129. Gentry-Nielsen MJ, Olsen KM, Preheim LC: Pharmacodynamic activity and efficacy of linezolid in a rat model of pneumococcal pneumonia. *Antimicrob Agents Chemother* 2002; 46:1345–1351
 130. Buerger C, Plock N, Dehghanyar P, et al: Pharmacokinetics of unbound linezolid in plasma and tissue interstitium of critically ill patients after multiple dosing using microdialysis. *Antimicrob Agents Chemother* 2006; 50:2455–2463
 131. French G: Safety and tolerability of linezolid. *J Antimicrob Chemother* 2003; 51 (Suppl 2):ii45–53
 132. Meagher AK, Ambrose PG, Grasela TH et al: The pharmacokinetic and pharmacodynamic profile of tigecycline. *Clin Infect Dis* 2005; 41 (Suppl 5):S333–S340
 133. Meagher AK, Ambrose PG, Grasela TH, et al: Pharmacokinetic/pharmacodynamic profile for tigecycline—a new glycolcylcine antimicrobial agent. *Diagn Microbiol Infect Dis* 2005; 52:165–171
 134. Muralidharan G, Micalizzi M, Speth J, et al: Pharmacokinetics of tigecycline after single and multiple doses in healthy subjects. *Antimicrob Agents Chemother* 2005; 49: 220–229
 135. Petersen PJ, Jacobus NV, Weiss WJ, et al: In vitro and in vivo antibacterial activities of a novel glycolcylcine, the 9-t-butylglycylamido derivative of minocycline (GAR-936). *Antimicrob Agents Chemother* 1999; 43: 738–744
 136. Fass RJ, Saslaw S: Clindamycin: clinical and laboratory evaluation of parenteral therapy. *Am J Med Sci* 1972; 263:368–382
 137. Mueller SC, Henkel KO, Neumann J, et al: Perioperative antibiotic prophylaxis in maxillofacial surgery: Penetration of clindamycin into various tissues. *J Craniomaxillofac Surg* 1999; 27:172–176
 138. Nagar H, Berger SA, Hammar B, et al: Penetration of clindamycin and metronidazole into the appendix and peritoneal fluid in children. *Eur J Clin Pharmacol* 1989; 37: 209–210
 139. Craig WA: Does the dose matter? *Clin Infect Dis* 2001; 33 (Suppl 3):S233–S237
 140. Mann HJ, Townsend RJ, Fuhs DW, et al: Decreased hepatic clearance of clindamycin in critically ill patients with sepsis. *Clin Pharm* 1987; 6:154–159
 141. Li J, Nation RL, Turnidge JD, et al: Colistin: The re-emerging antibiotic for multidrug-resistant Gram-negative bacterial infections. *Lancet Infect Dis* 2006; 6:589–601
 142. Falagas ME, Kasiakou SK: Colistin: The revival of polymyxins for the management of multidrug-resistant gram-negative bacterial infections. *Clin Infect Dis* 2005; 40: 1333–1341
 143. Reed MD, Stern RC, O’Riordan MA, et al: The pharmacokinetics of colistin in patients with cystic fibrosis. *J Clin Pharmacol* 2001; 41:645–654
 144. Catchpole CR, Andrews JM, Brenwald N, et al: A reassessment of the in-vitro activity of colistin sulphomethate sodium. *J Antimicrob Chemother* 1997; 39:255–260
 145. Tam VH, Schilling AN, Vo G, et al: Pharmacodynamics of polymyxin B against *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2005; 49:3624–3630
 146. Nation RL, Li J: Optimizing use of colistin and polymyxin B in the critically ill. *Sem Respir Crit Care Med* 2007; 28:604–614
 147. Markou N, Markantonis SL, Dimitrakis E, et al: Colistin serum concentrations after intravenous administration in critically ill patients with serious multidrug-resistant, gram-negative bacilli infections: A prospective, open-label, uncontrolled study. *Clin Ther* 2008; 30:143–151
 148. Chastre J, Wunderink R, Prokocimer P, et al: Efficacy and safety of intravenous infusion of doripenem versus imipenem in ventilator-associated pneumonia: A multicenter, randomized study. *Crit Care Med* 2008; 36:1089–1096
 149. Daikos GL, Lolans VT, Jackson GG: First-exposure adaptive resistance to aminoglycoside antibiotics in vivo with meaning for optimal clinical use. *Antimicrob Agents Chemother* 1991; 35:117–123
 150. Gumbo T, Louie A, Deziel MR, et al: Selection of a moxifloxacin dose that suppresses drug resistance in *Mycobacterium tuberculosis*, by use of an in vitro pharmacodynamic infection model and mathematical modeling. *J Infect Dis* 2004; 190:1642–1651
 151. Henderson-Begg SK, Livermore DM, Hall LM: Effect of subinhibitory concentrations of antibiotics on mutation frequency in *Streptococcus pneumoniae*. *J Antimicrob Chemother* 2006; 57:849–854
 152. Olofsson SK, Geli P, Andersson DI, et al: Pharmacodynamic model to describe the concentration-dependent selection of cefotaxime-resistant *Escherichia coli*. *Antimicrob Agents Chemother* 2005; 49: 5081–5091
 153. Tam VH, Schilling AN, Neshat S, et al: Optimization of meropenem minimum concentration/MIC ratio to suppress in vitro resistance of *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2005; 49: 4920–4927
 154. Tsuji B, Rybak M: The influence of *Staphylococcus aureus* accessory gene regulator function on the development of glycopeptide hetero-resistance in an in vitro phar-

- macodynamic model moxifloxacin. *In*: 15th European Congress on Clinical Microbiology and Infectious Diseases: April 2–5 2005, Nice, France; Copenhagen, Denmark, 2005, p 1590
155. Lipman J, Wallis SC, Boots RJ: Cefepime versus ceftazidime: The importance of creatinine clearance (table of contents). *Anesth Analg* 2003; 97:1149–1154
 156. Roos JF, Bulitta J, Lipman J, et al: Pharmacokinetic-pharmacodynamic rationale for cefepime dosing regimens in intensive care units. *J Antimicrob Chemother* 2006; 58: 987–993
 157. Bennett WM, Aronoff GR, Golper TA, et al: Drug Prescribing in Renal Failure. Third Edition. Philadelphia, American College of Physicians, 1994
 158. Brittain DC, Scully BE, McElrath MJ, et al: The pharmacology of orally administered ciprofloxacin. *Drugs Exp Clin Res* 1985; 11:339–341
 159. Micromedex Healthcare Series, Micromedex, Date accessed June 2008
 160. Chien SC, Chow AT, Natarajan J, et al: Absence of age and gender effects on the pharmacokinetics of a single 500-milligram oral dose of levofloxacin in healthy subjects. *Antimicrob Agents Chemother* 1997; 41: 1562–1565
 161. Stass H: Distribution and tissue penetration of moxifloxacin. *Drugs* 1999; 58(Suppl 2): 229–230
 162. Stass H, Dalhoff A, Kubitzka D, et al: Pharmacokinetics, safety, and tolerability of ascending single doses of moxifloxacin, a new 8-methoxy quinolone, administered to healthy subjects. *Antimicrob Agents Chemother* 1998; 42:2060–2065
 163. MIMS Australia, Ciproxin(R) Product Information
 164. Nakashima M, Uematsu T, Kosuge K, et al: Single- and multiple-dose pharmacokinetics of AM-1155, a new 6-fluoro-8-methoxy quinolone, in humans. *Antimicrob Agents Chemother* 1995; 39:2635–2640
 165. Sweetman SC (Ed): Martindale. Thirty-fourth Edition. London, The Pharmaceutical Press, 2005